# Positive Feedback Between Synthetic Biology and Natural Learning

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University of Alaska - Fairbanks March 2, 2012

#### Outline of Presentation

- 1. Introduce synthetic biology
- 2. Applications of synthetic biology
- 3. Synthetic biology research at Davidson College
- 4. Why make biological computers?
- 5. How do we prepare undergraduates for research?

#### What is Synthetic Biology?

Implementation of engineering principles and mathematical modeling to the design and construction of biological parts, devices, and systems with applications in energy, medicine, and technology.

www.bio.davidson.edu/projects/gcat/Synthetic/What\_Is\_SynBio.html

## Synthetic Biology: Win-Win

Win #1: your design functions as expected.



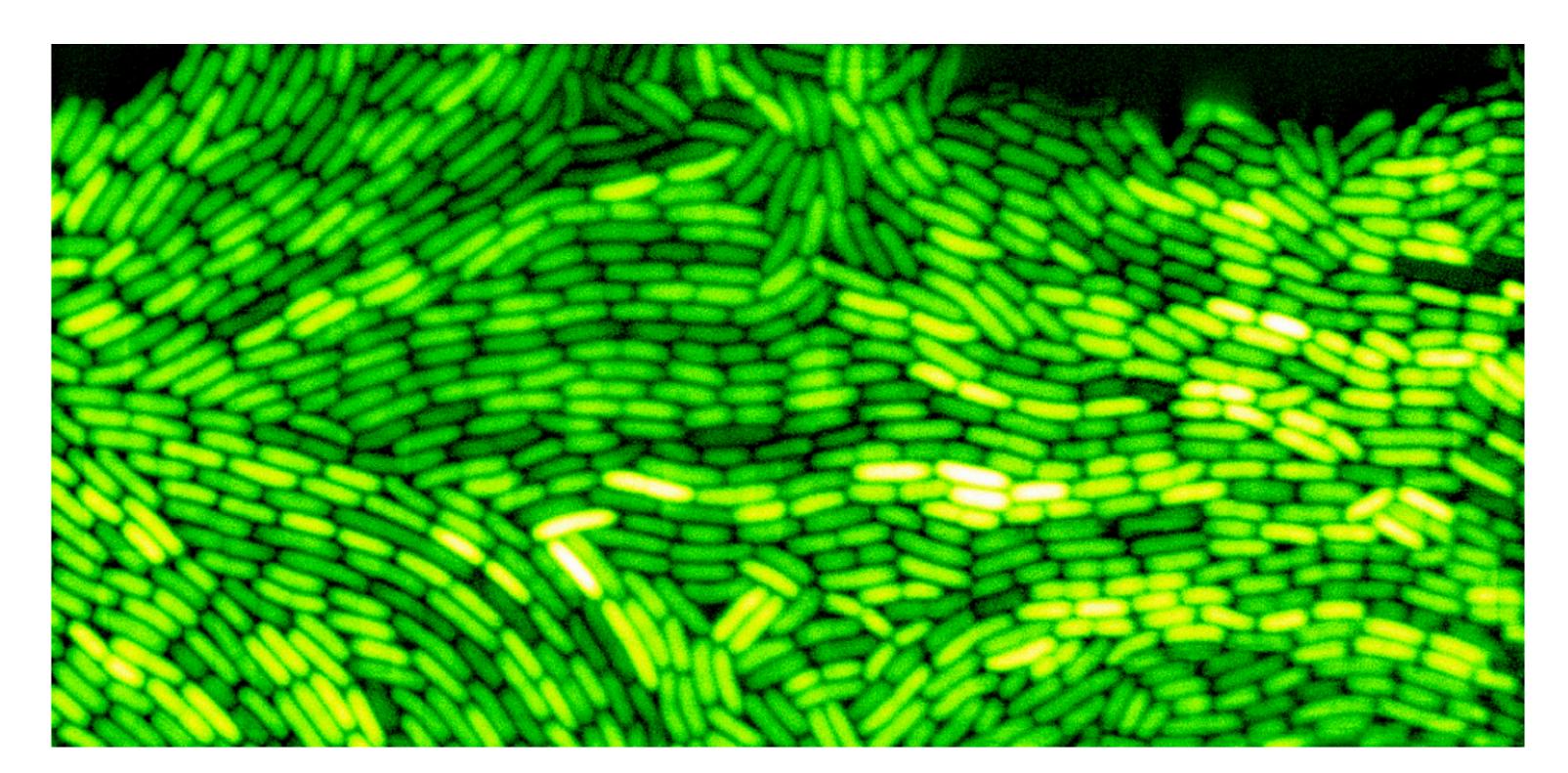


#### Synthetic Biology: Win-Win Research



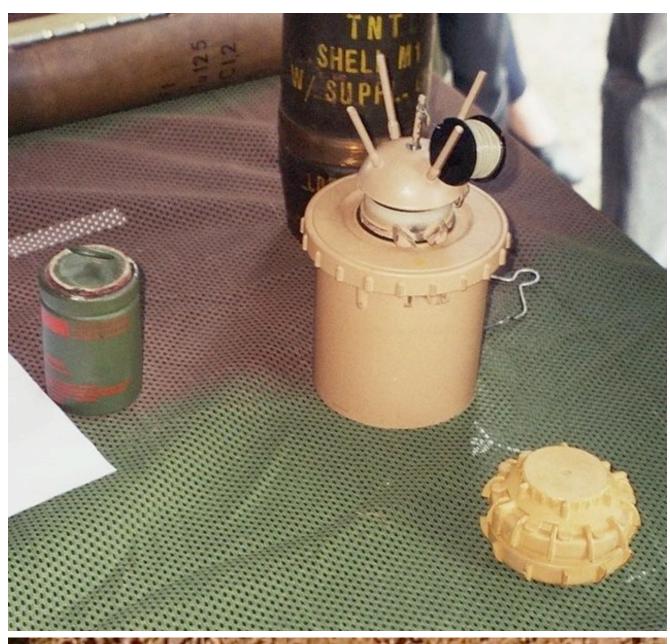
Win #1: your design functions as expected.



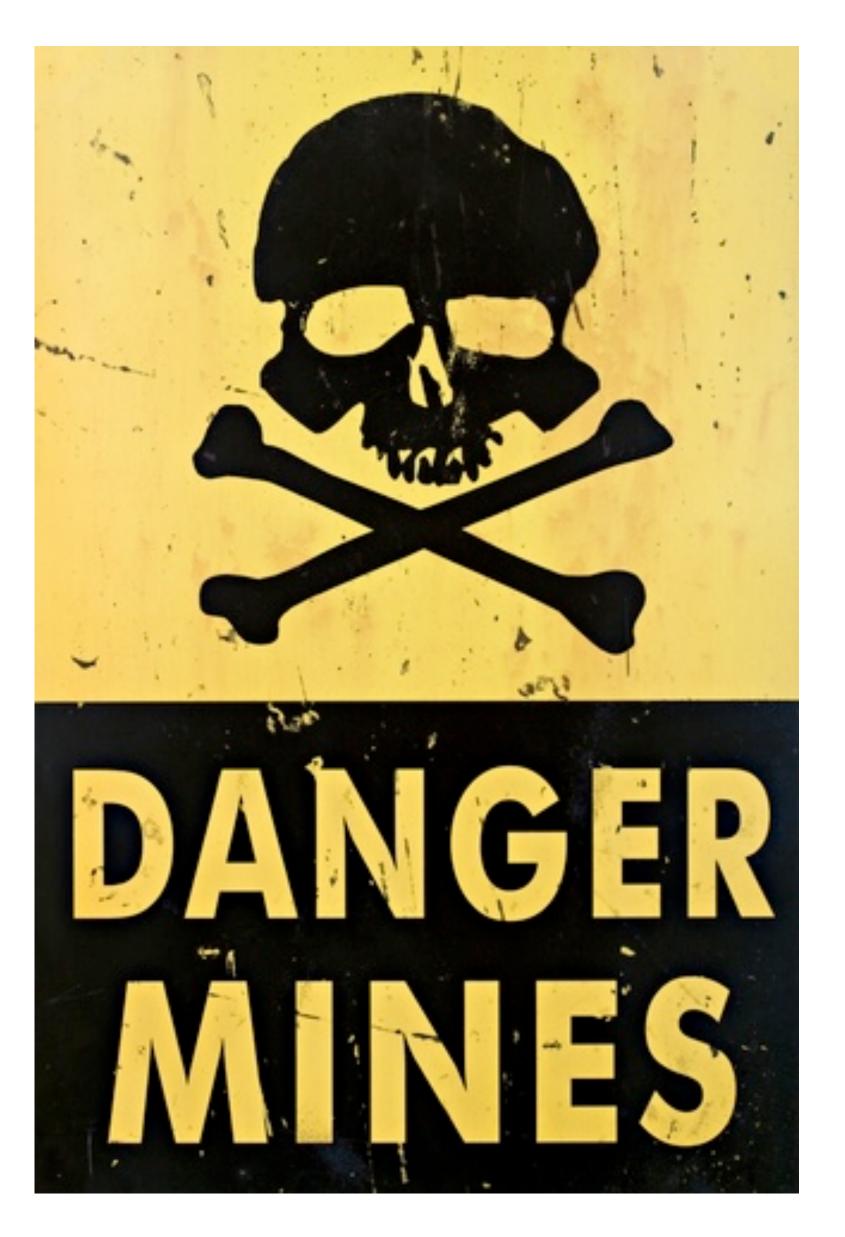


# Real World Applications of Synthetic Biology

#### Land Mine Detection

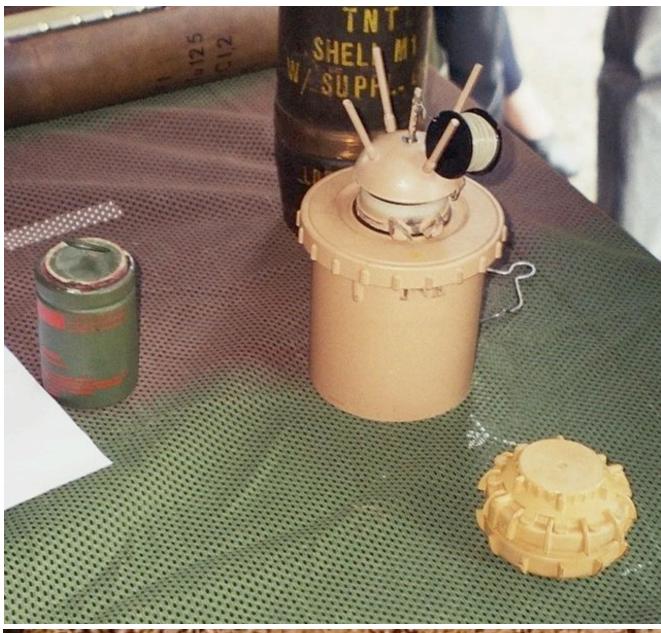






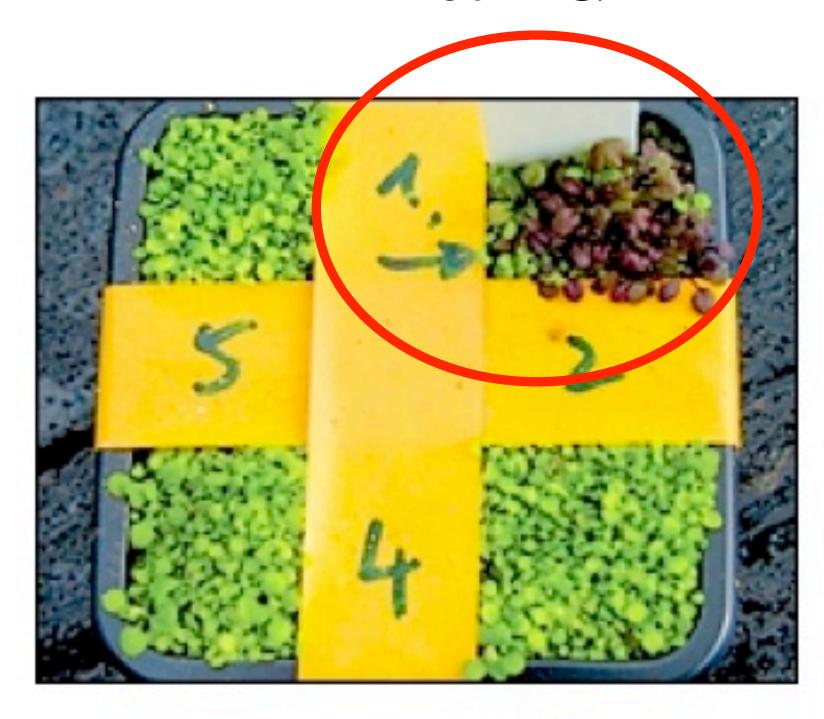
#### Land Mine Detection







# Synthetic Biology Land Mine Detection



#### WARNING SIGN: The

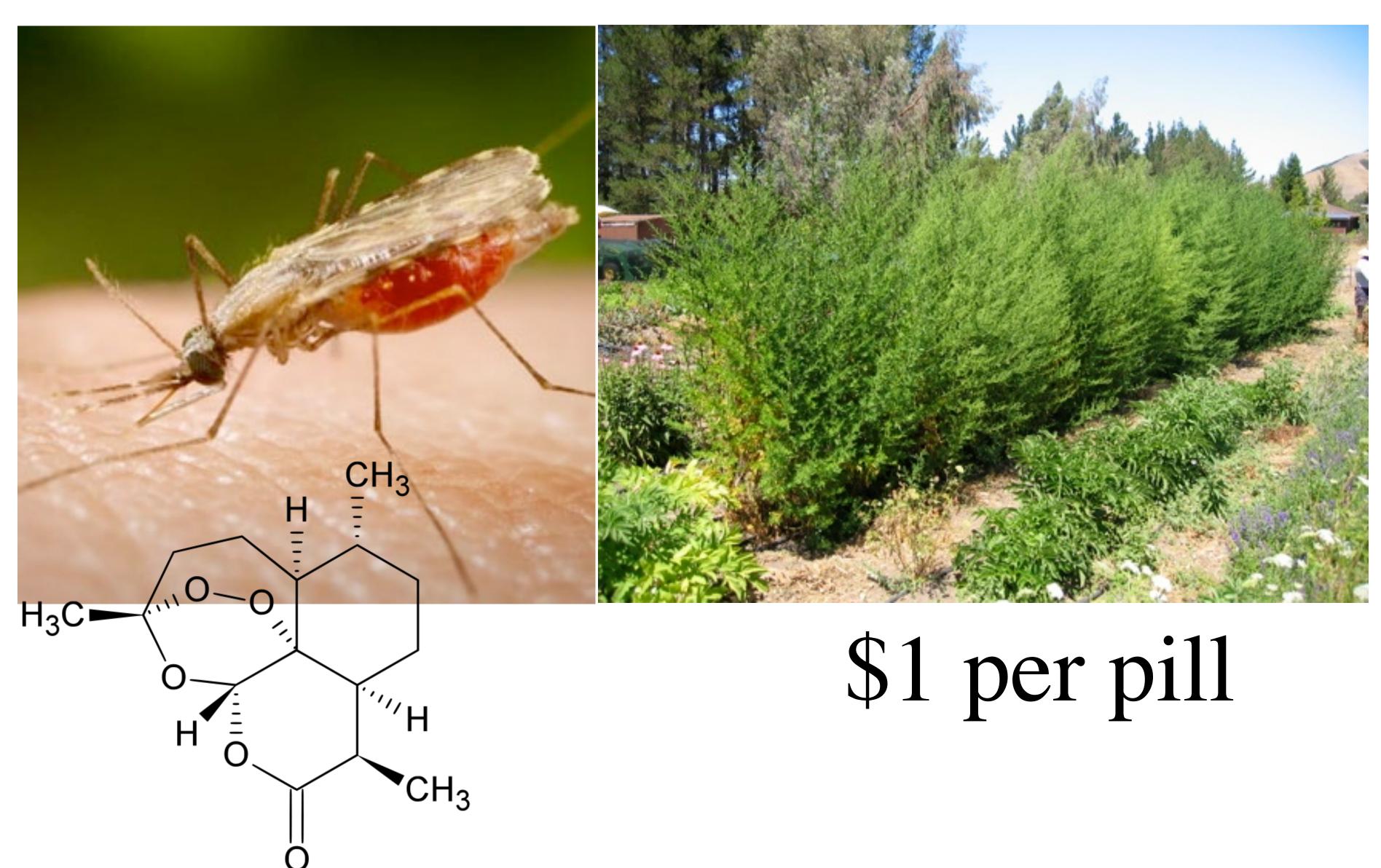
bioengineered Thales cress turns red when exposed to a mine byproduct.

COURTESY OF ARESA BIODETECTION

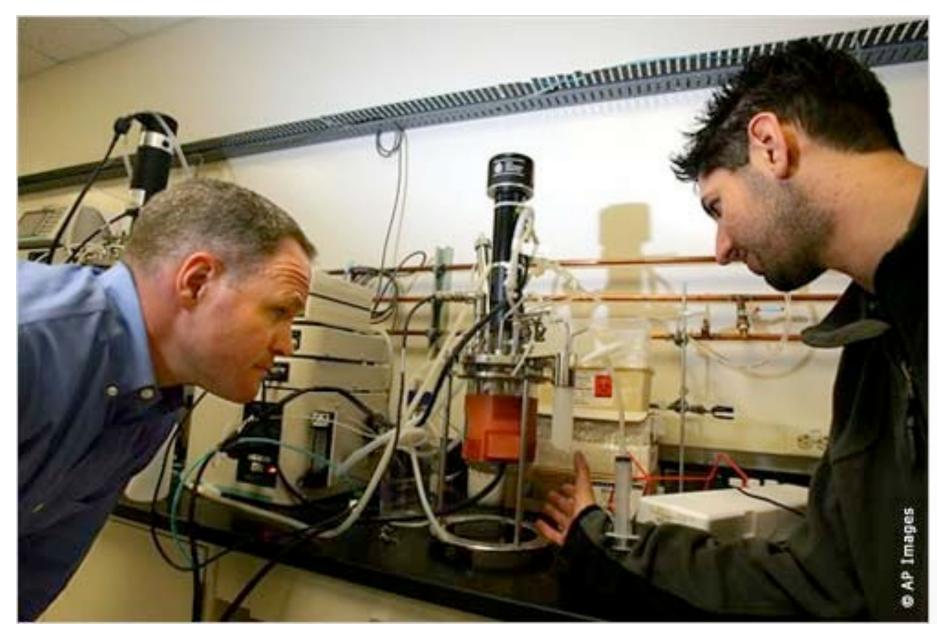
## New weed may flag land mines

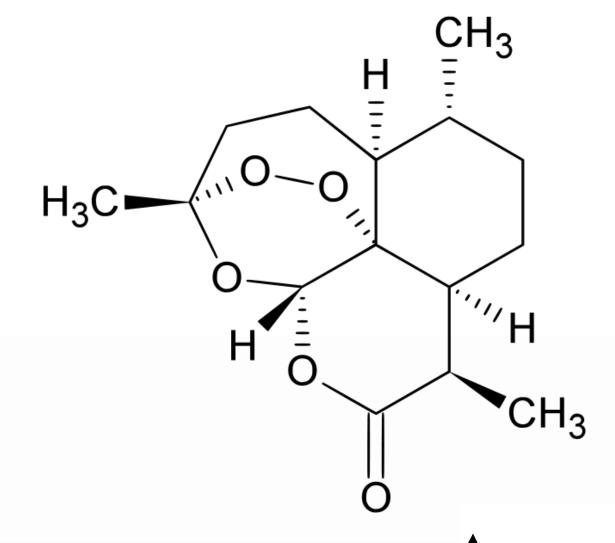
By John K. Borchardt | Contributor to The Christian Science Monitor

#### Production of Medicines

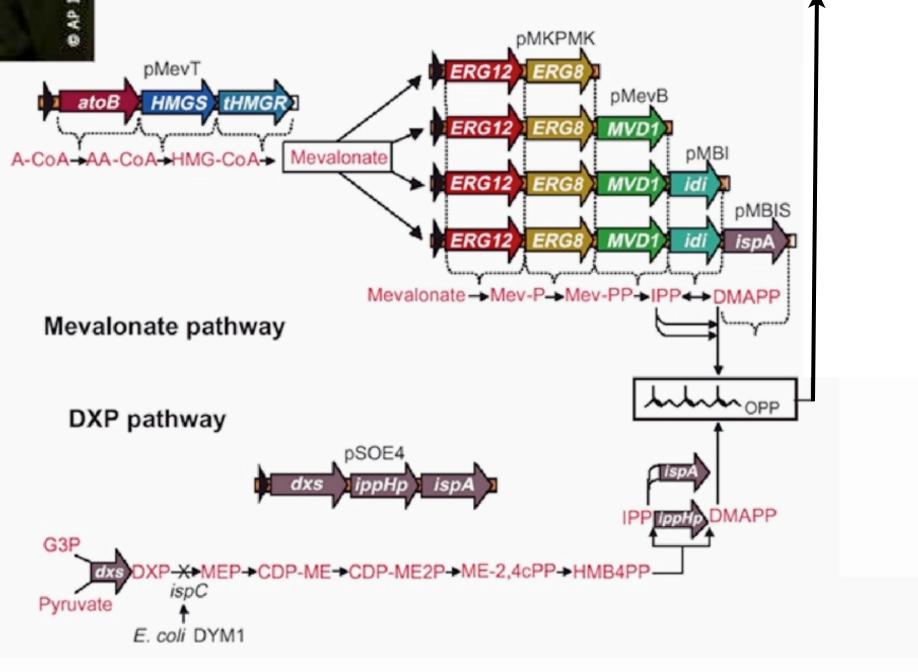


#### Production of Medicines





10¢ per pill



Biofuels from Algae





CO<sub>2</sub>-neutral 1,000,000 gallons in 2008

# Synthetic Biology at Davidson College



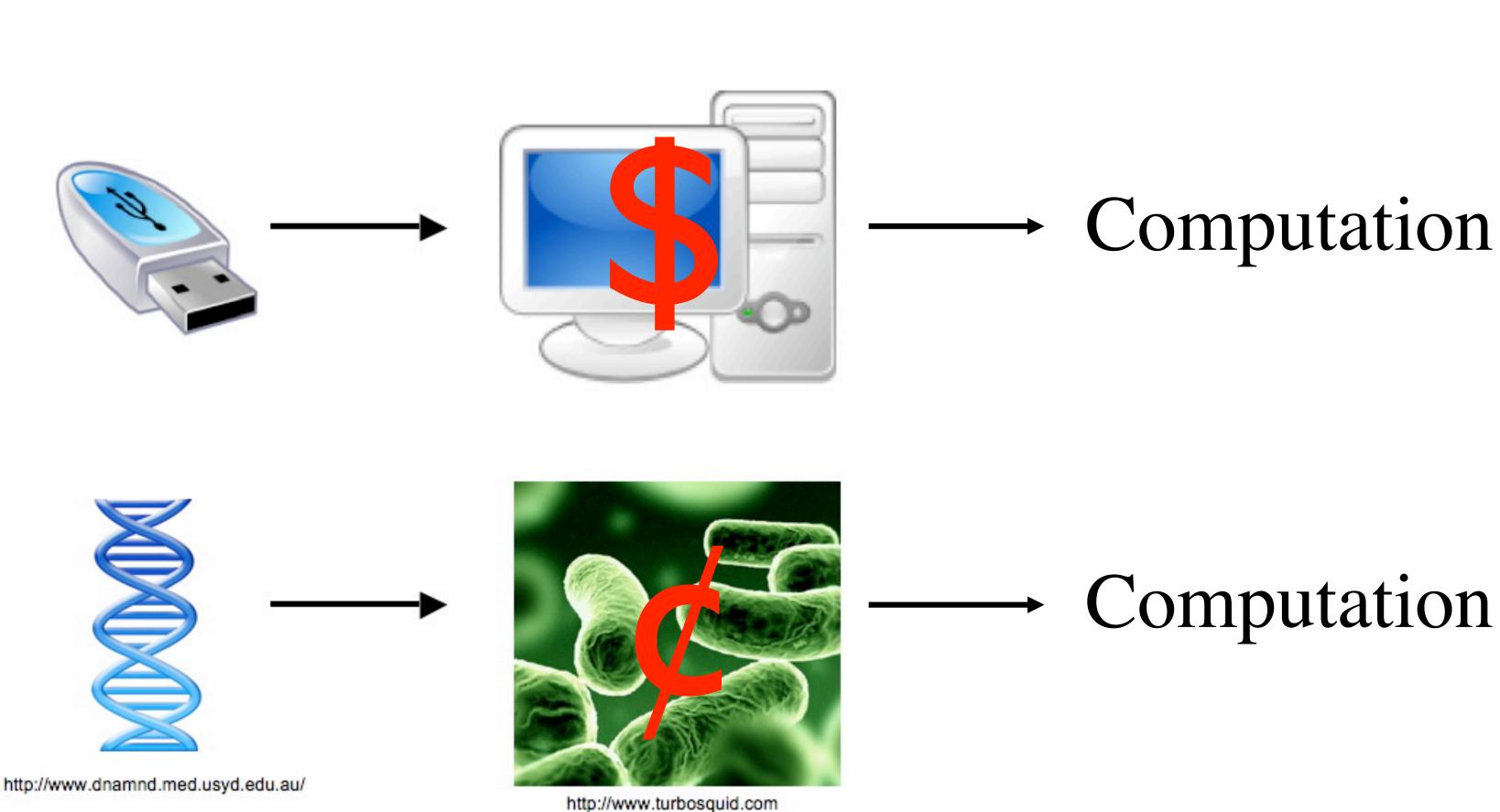


Laurie Heyer, Todd Eckdahl & Jeff Poet

Building Bacterial Computers

#### Advantages of Bacterial Computation

Software — Hardware — Computation



#### Advantages of Biological Computers

go anywhere - arctic, thermal vents, inside organisms

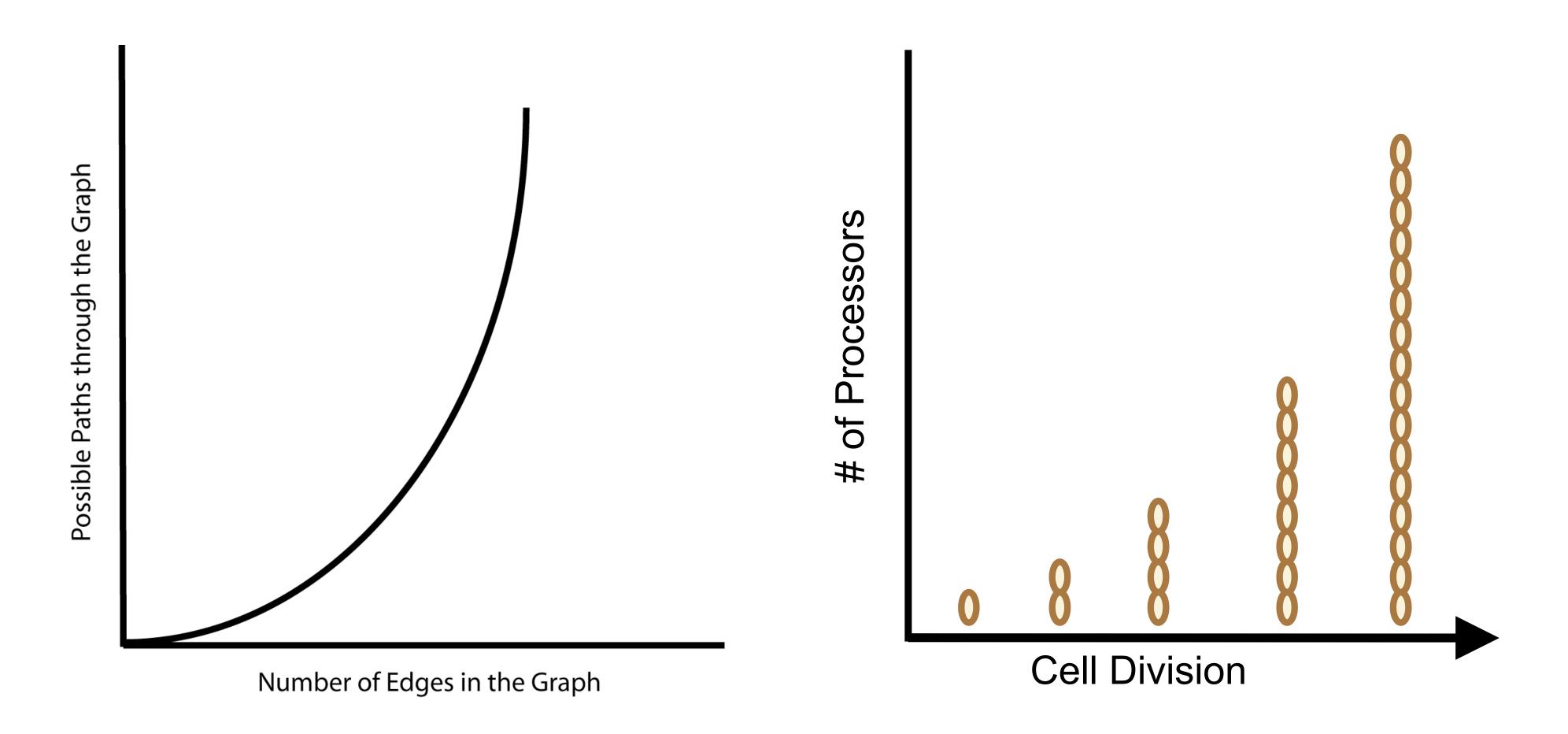
no electricity

self-replicating

no immune rejection



### Self-replicating Computers



# Two Undergraduate Research Projects

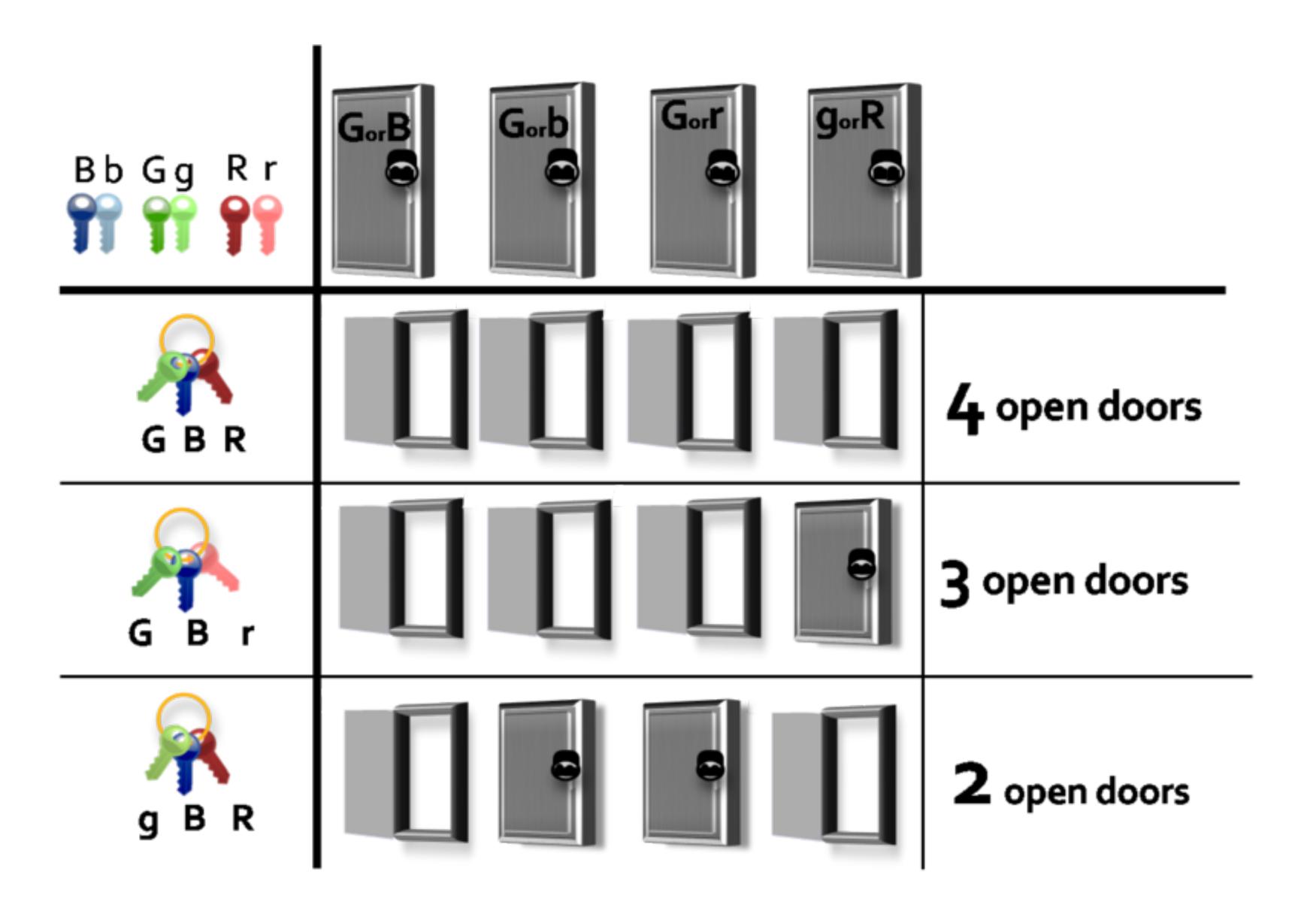
(G or B) & (G or b) & (G or r) & (g or R)

G, g, B, b, R, r

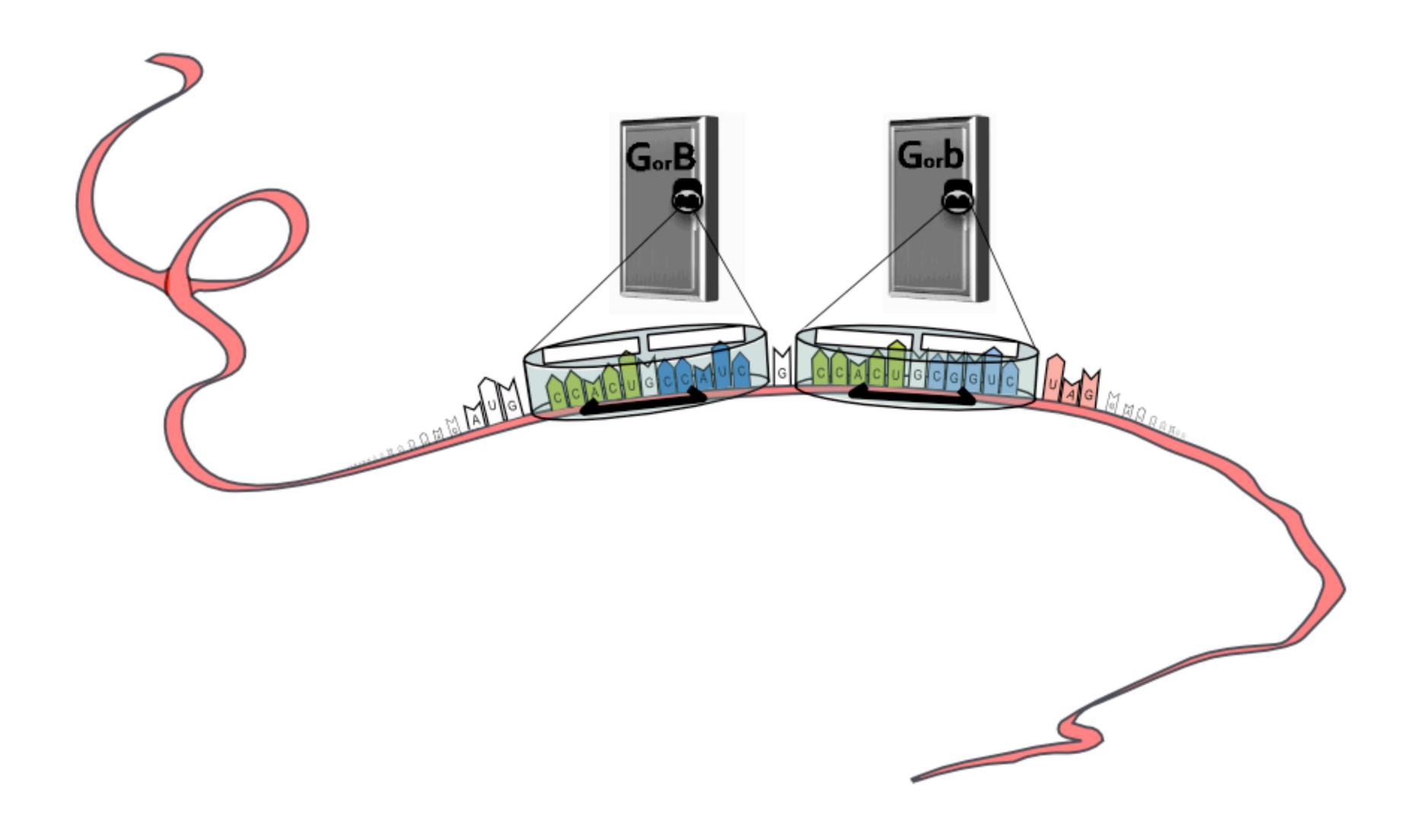


(G or B) & (G or b) & (G or r) & (g or R)

| Bb Gg Rr | G <sub>o</sub> rB | Gorb | Gort | gorR |  |
|----------|-------------------|------|------|------|--|
| G B R    |                   |      |      |      |  |
| G B      |                   |      |      |      |  |
| g B R    |                   |      |      |      |  |



## Converting Math to Biology



#### Central Dogma

DNA atgccctactcactactatagcgcat



transcription

mRNA aug ccc uac uca cua ccu aua ccg cau

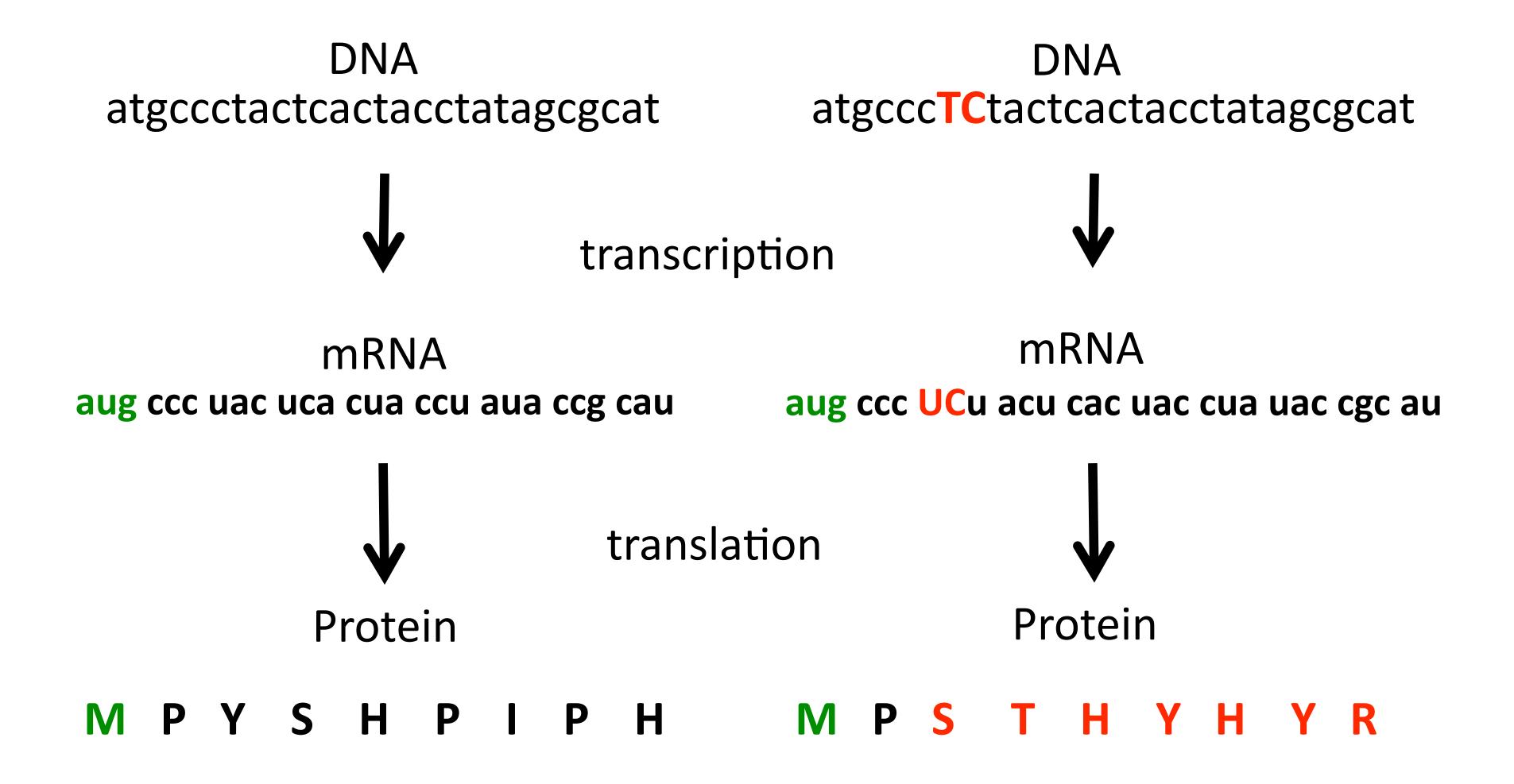


translation

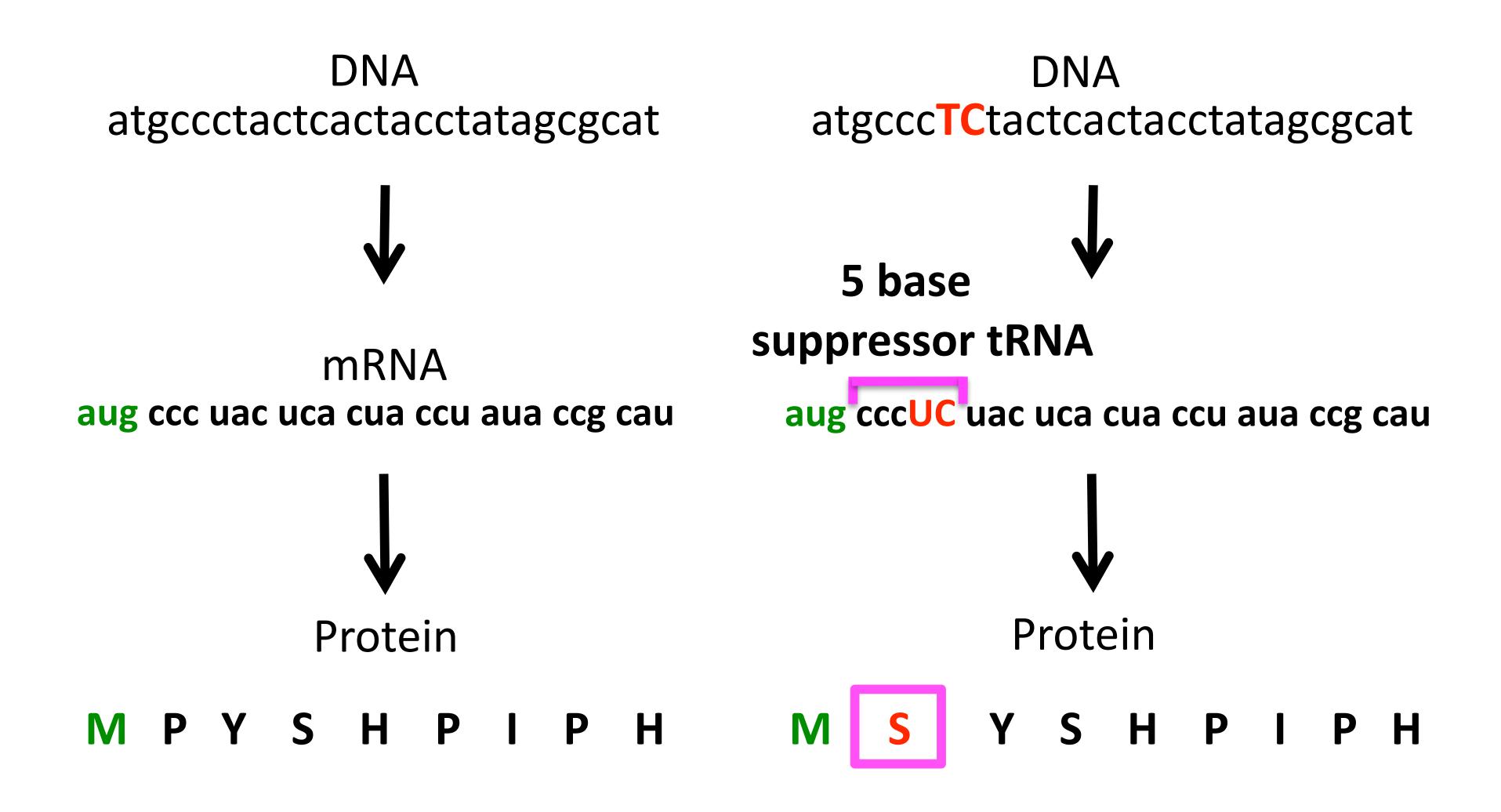
Protein

M P Y S H P I P H

#### Frameshift Mutation



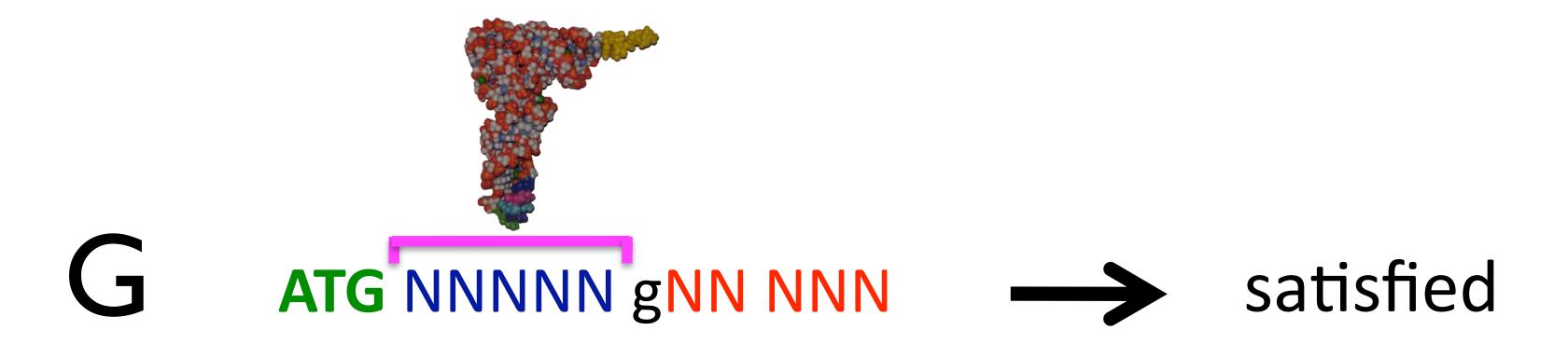
#### Frameshift Suppression



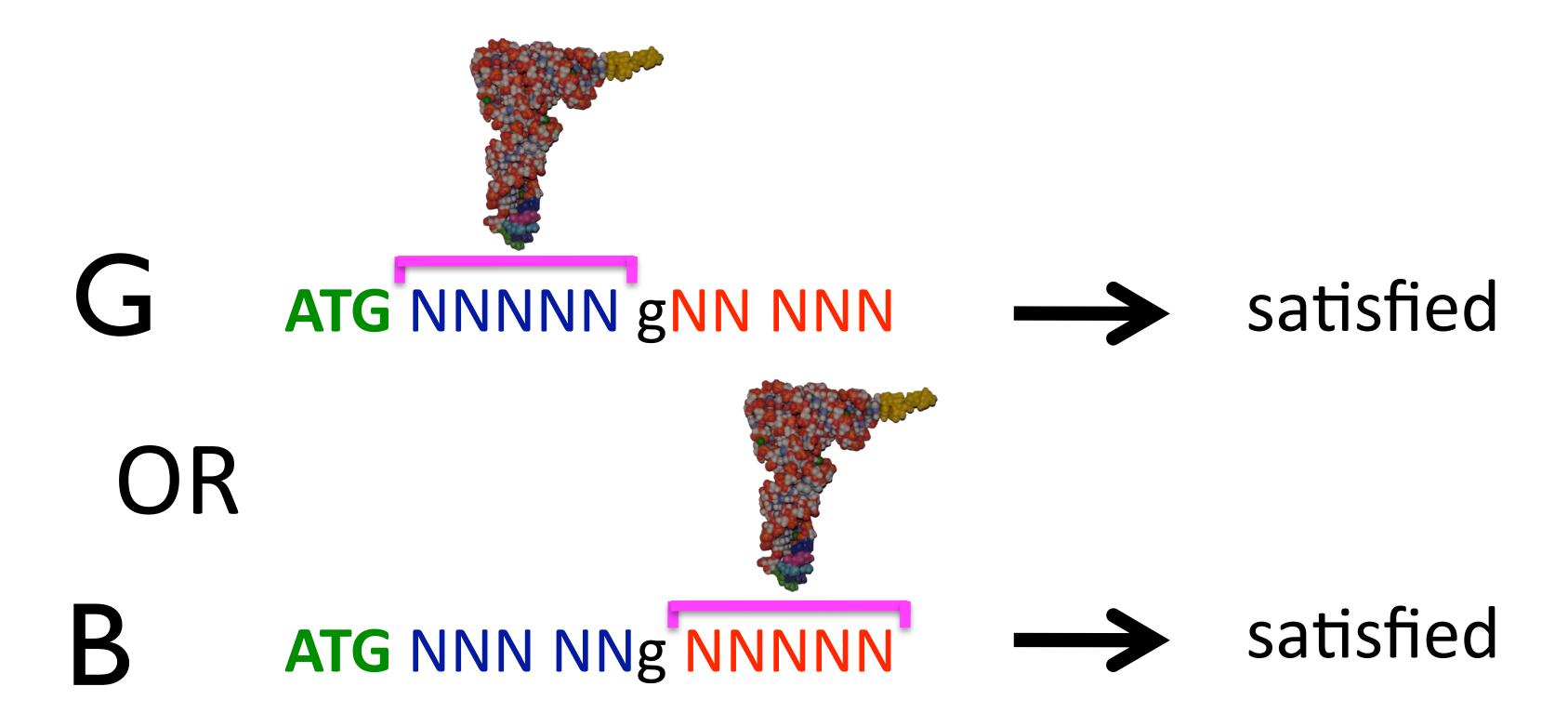
#### Suppressor tRNA

core tRNA nucleotides serine 5 base anticodon

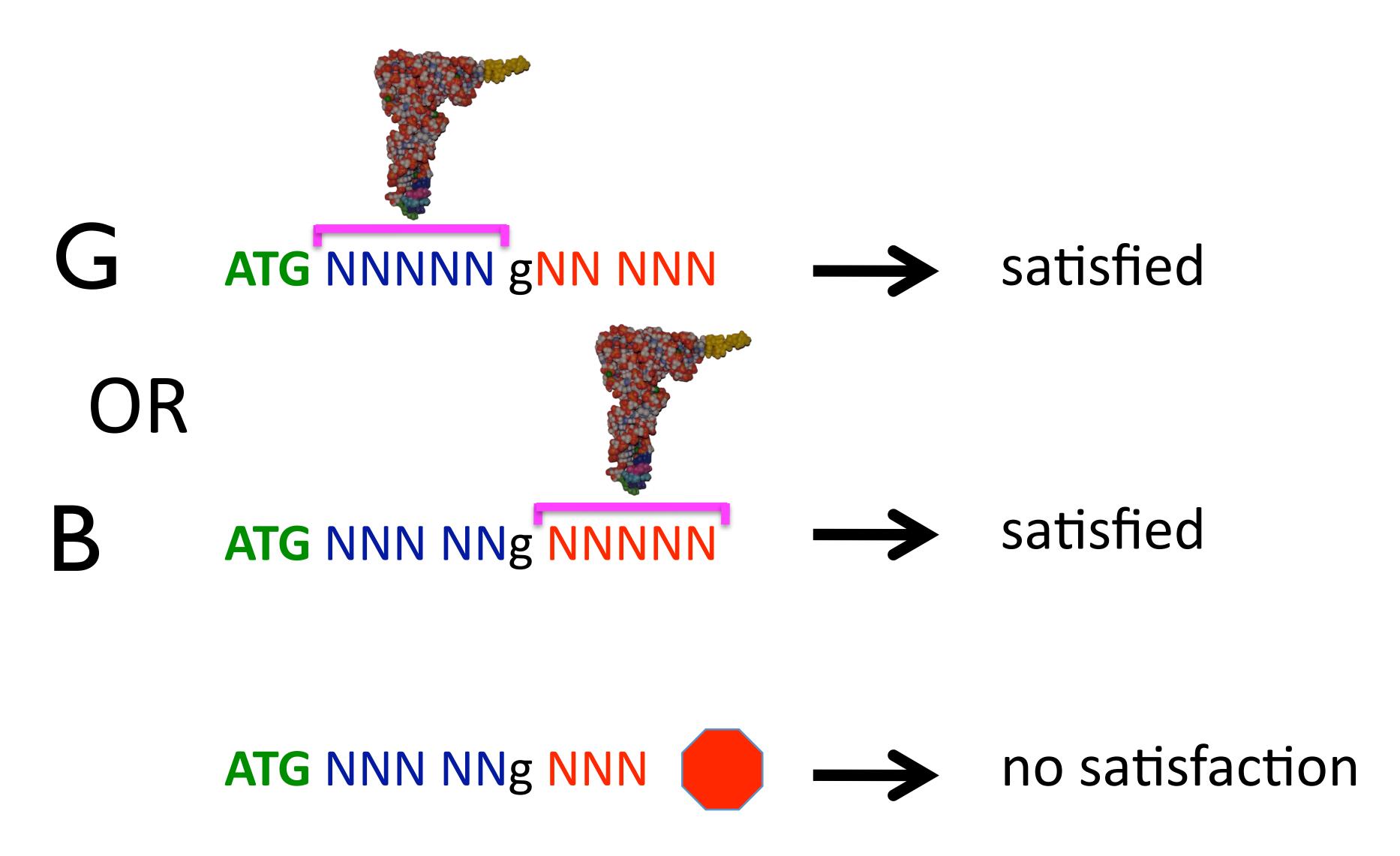
### Coding 2-SAT Clause

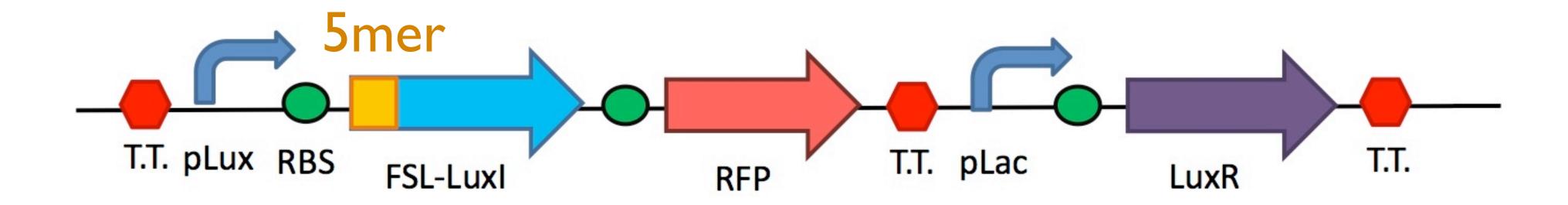


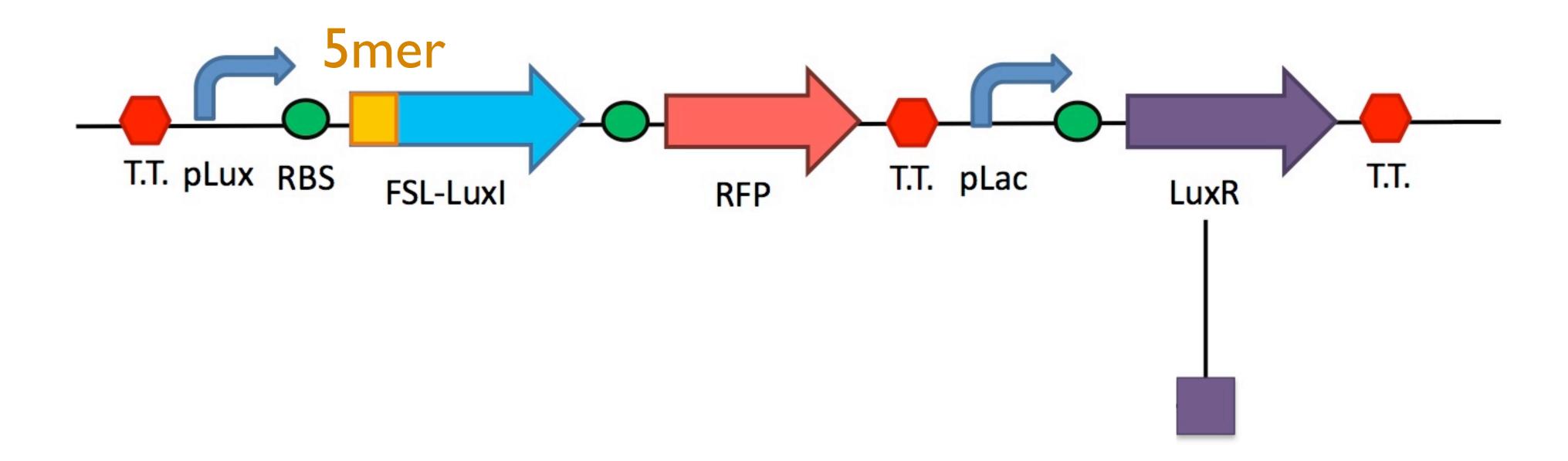
## Coding 2-SAT Clause

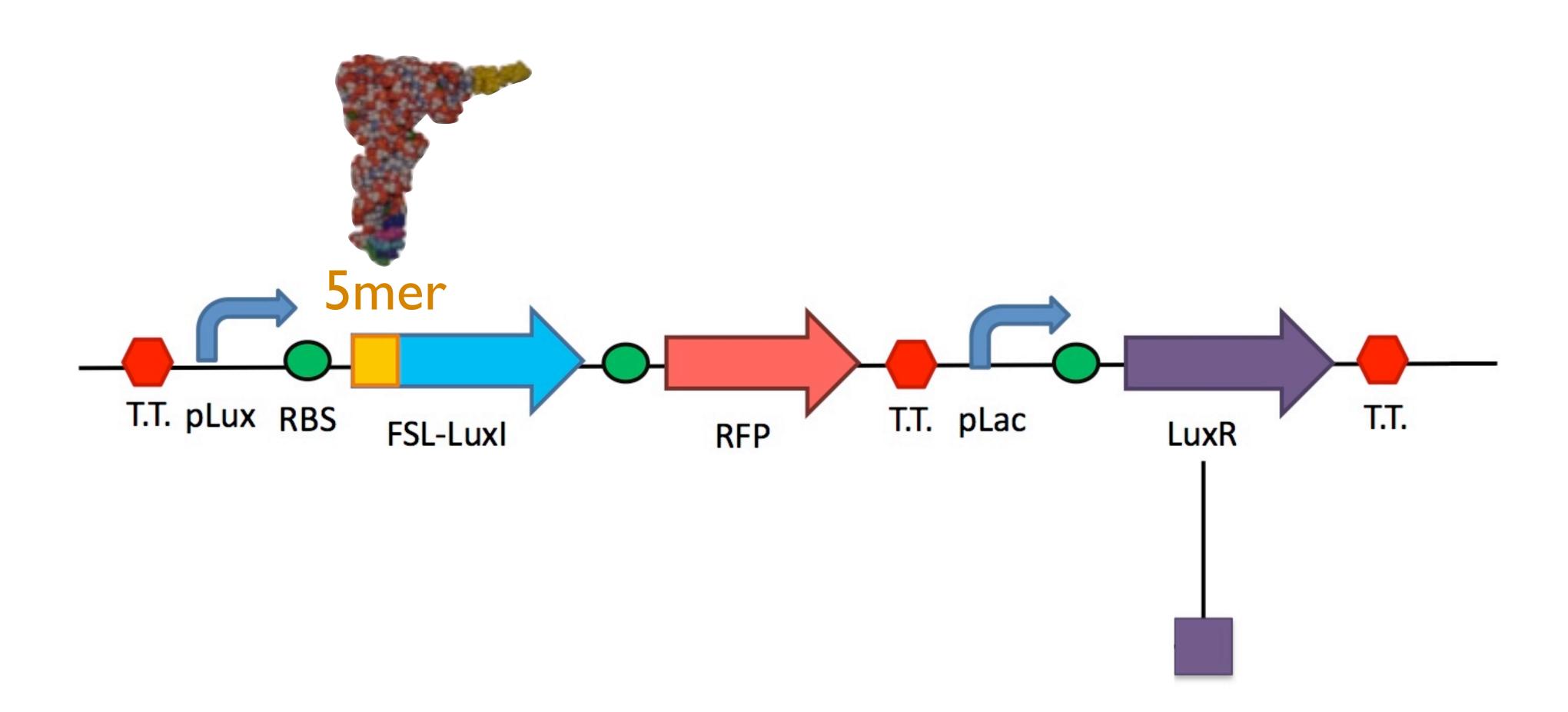


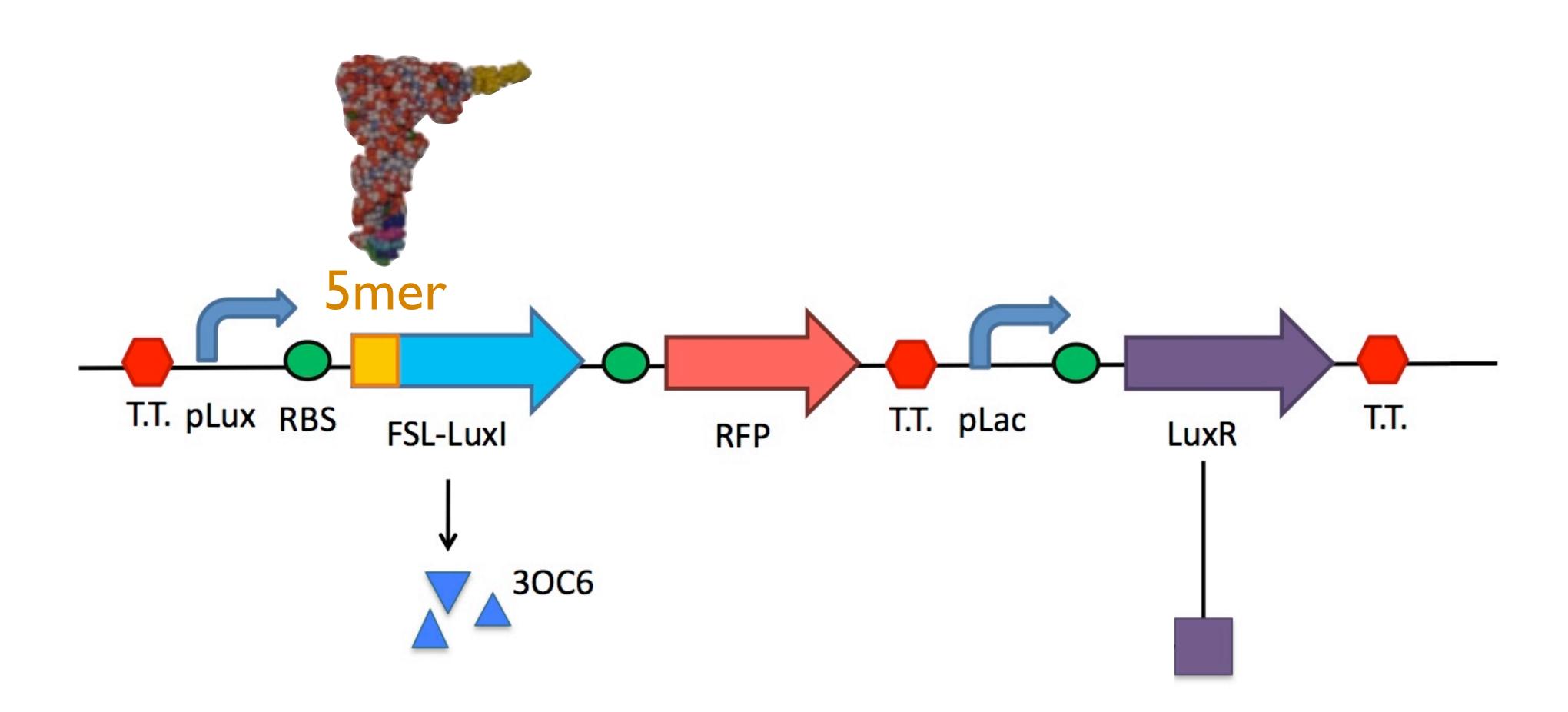
### Coding 2-SAT Clause

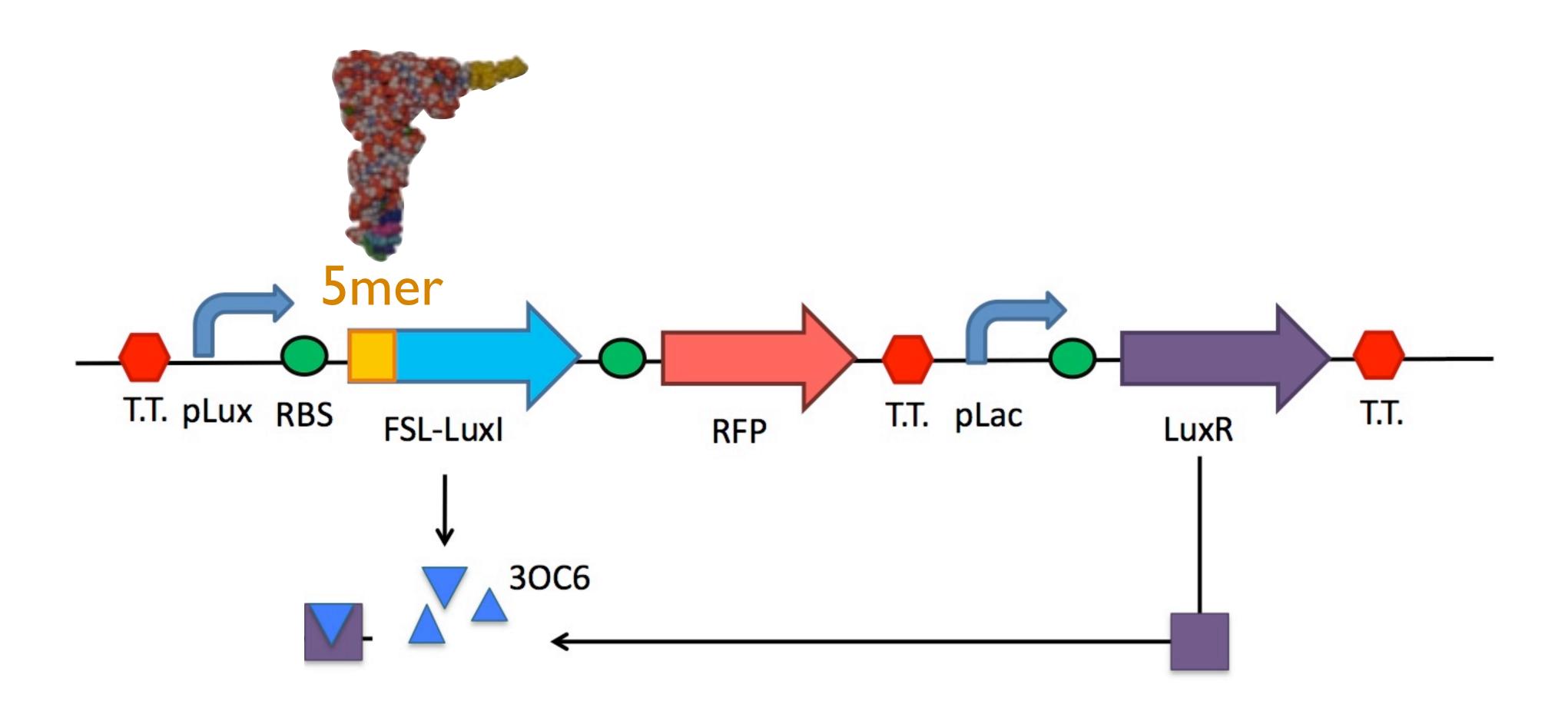


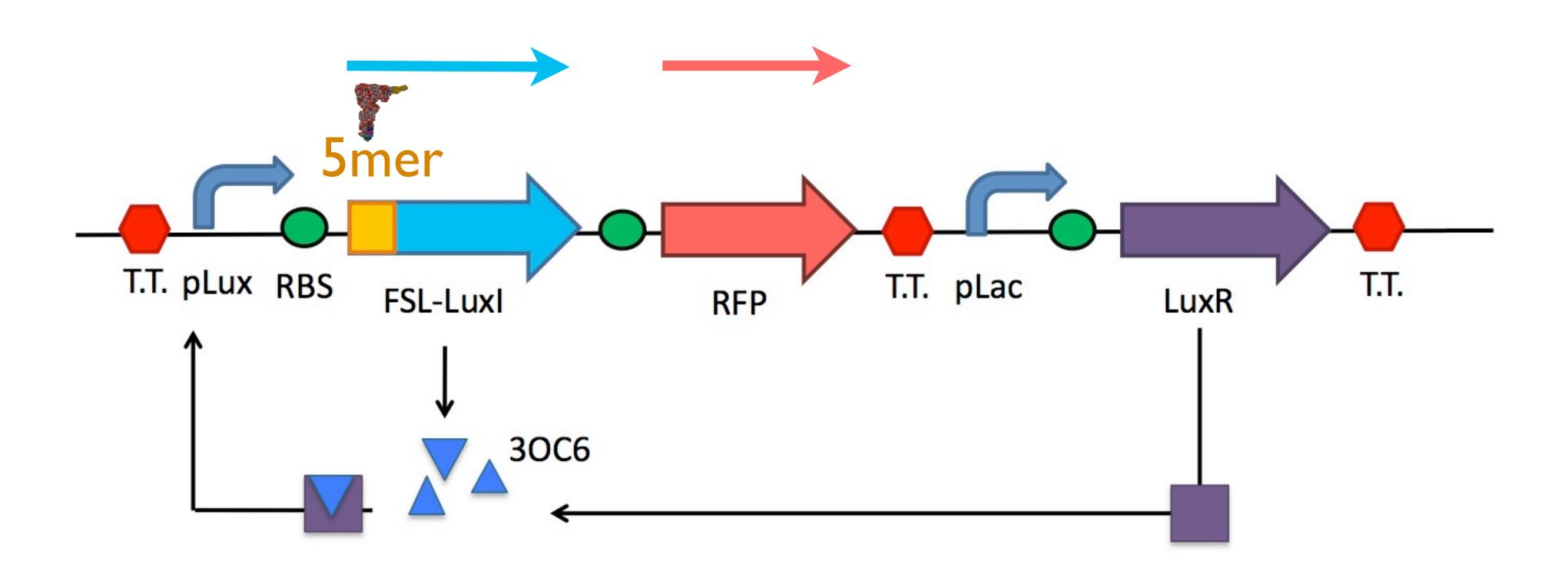


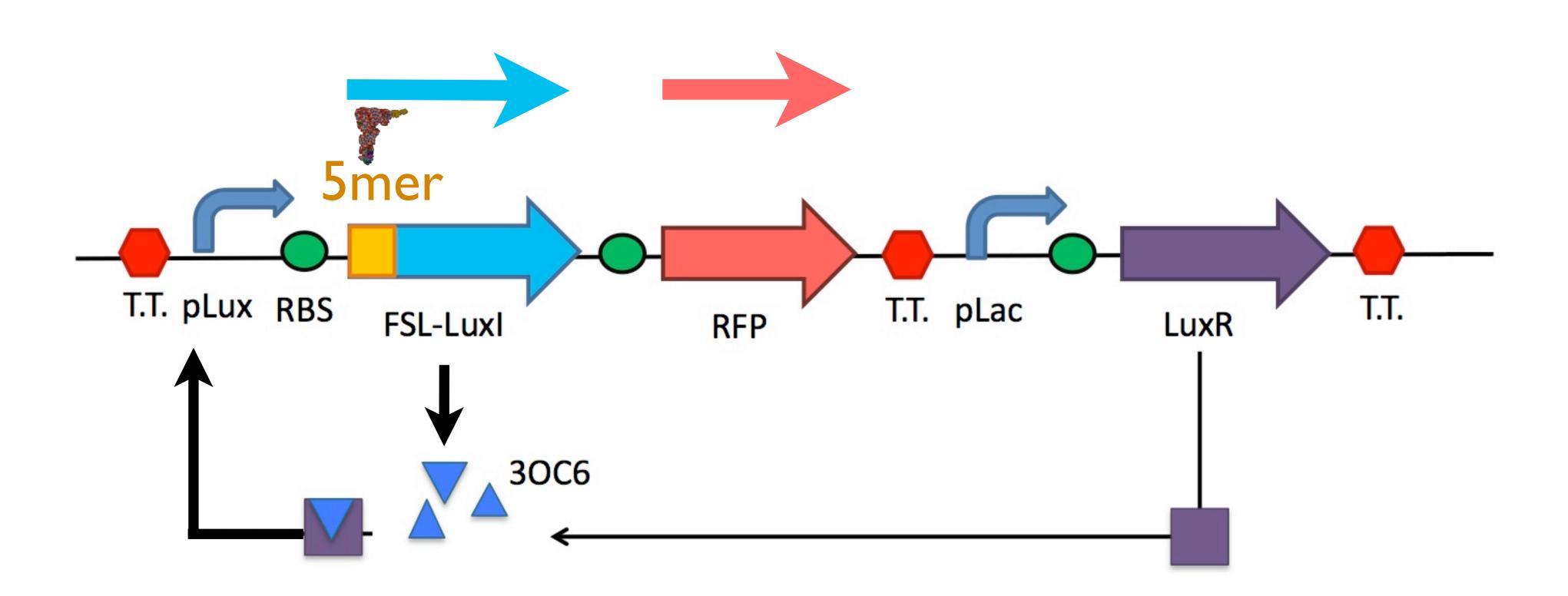










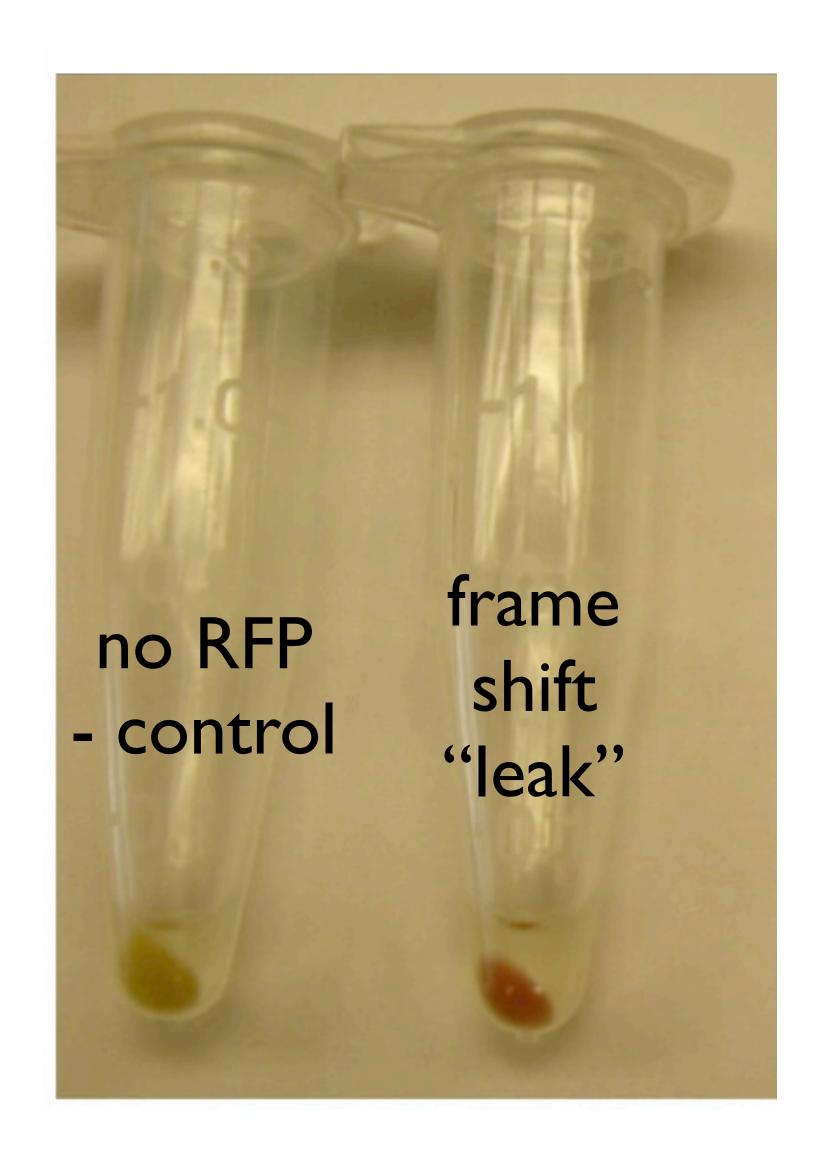


### Results



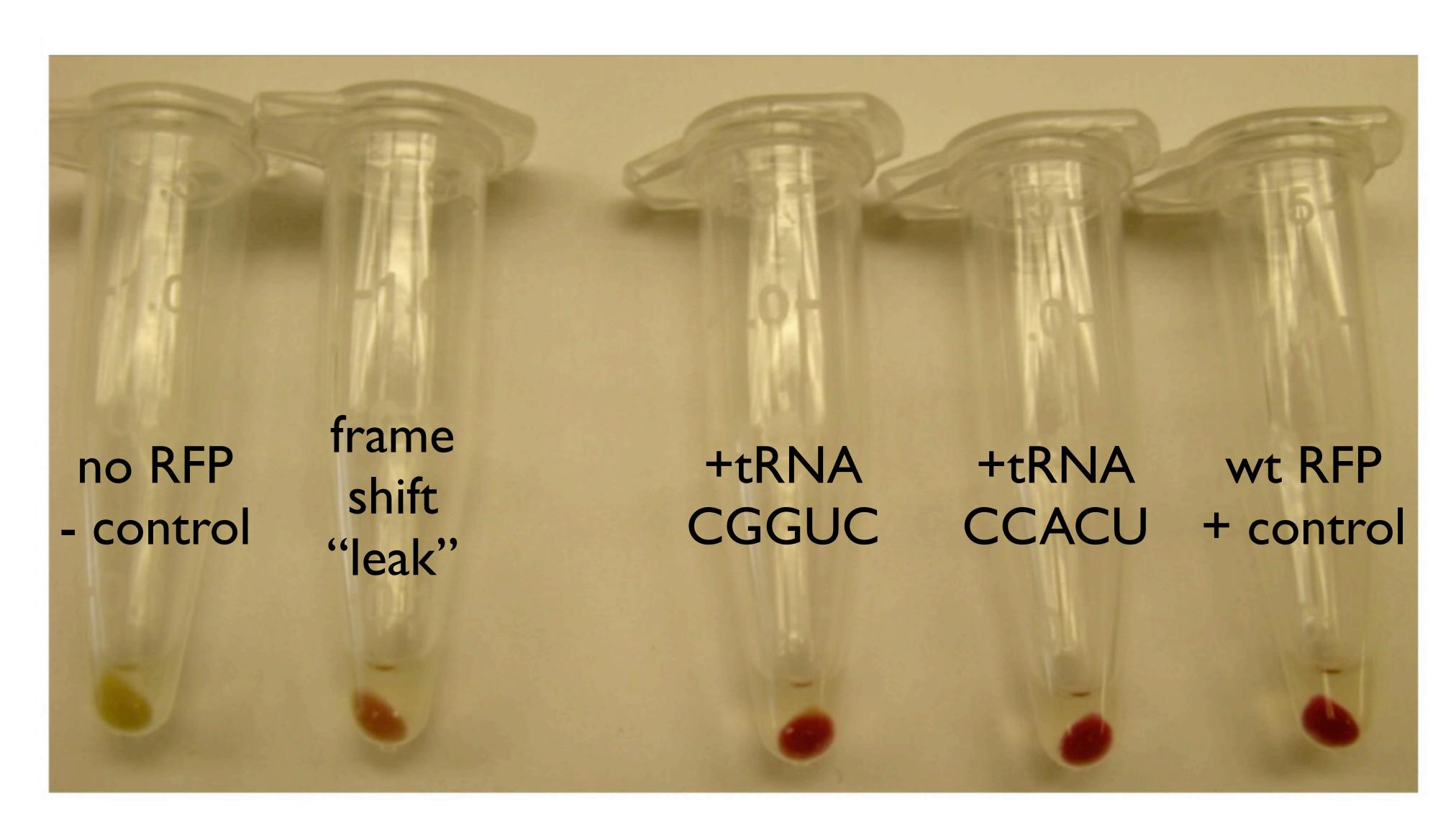


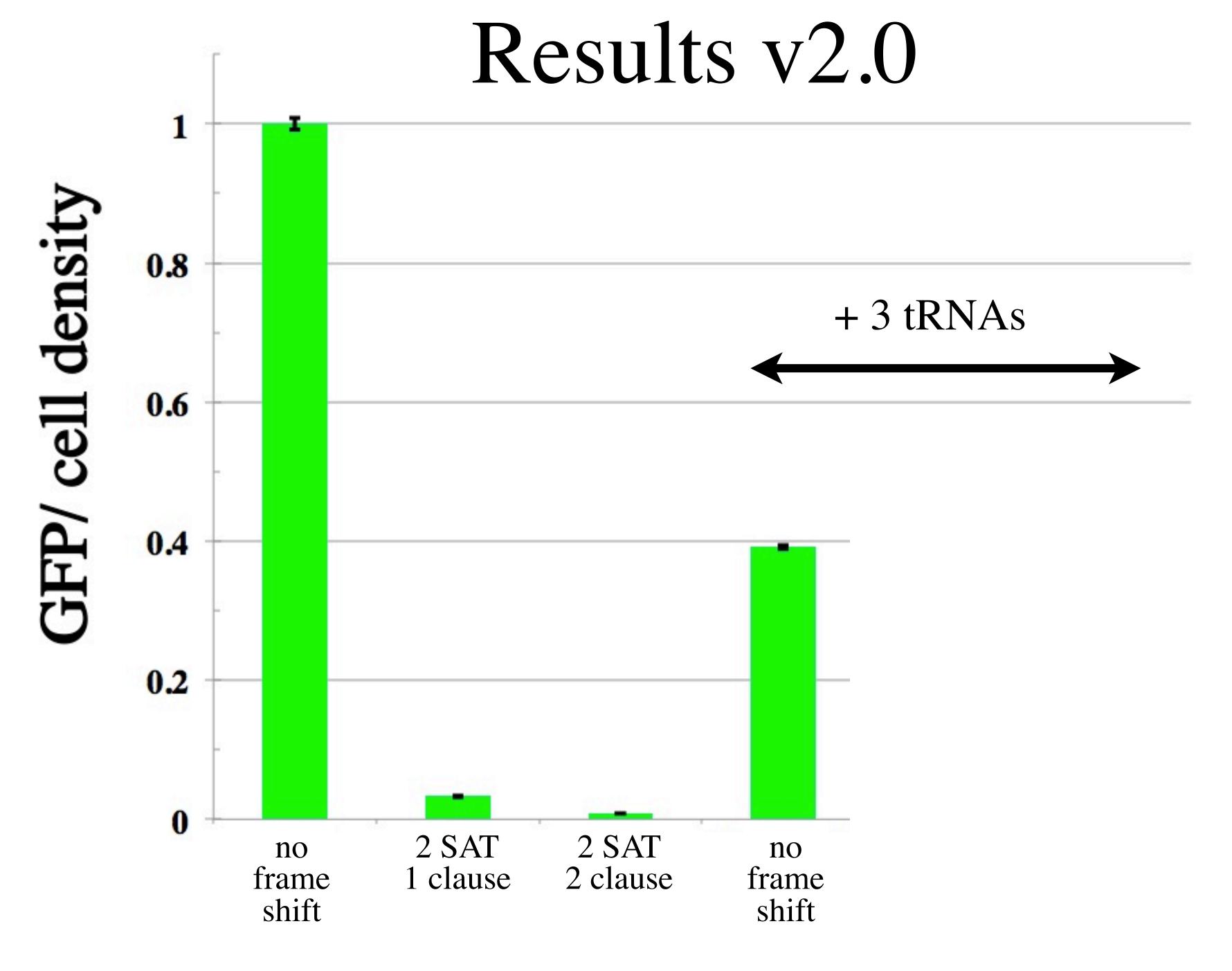
## Results

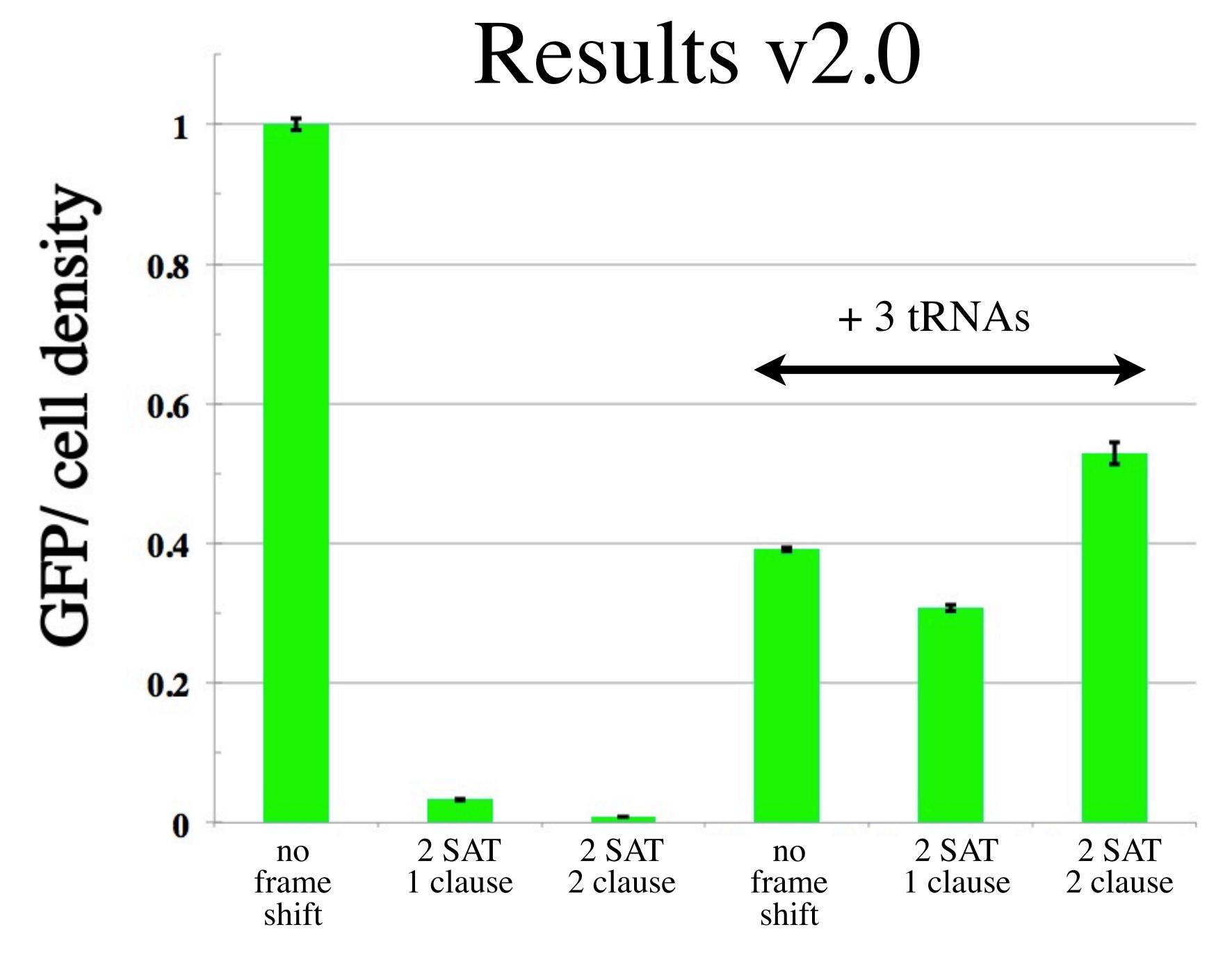




#### Results







# Can we build a bacterial cryptographic hash function?

#### What is a hash function?



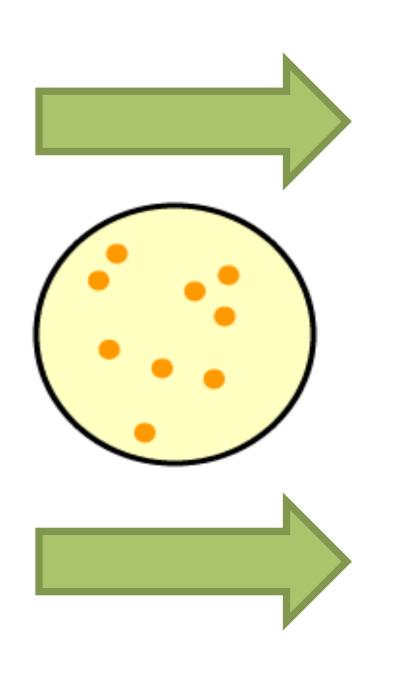
HGTf34\$2





#### Can Bacteria Perform a Hash Function?

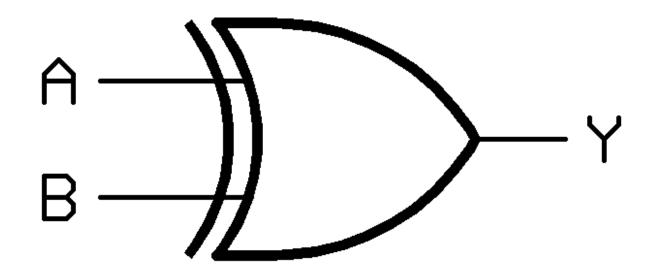




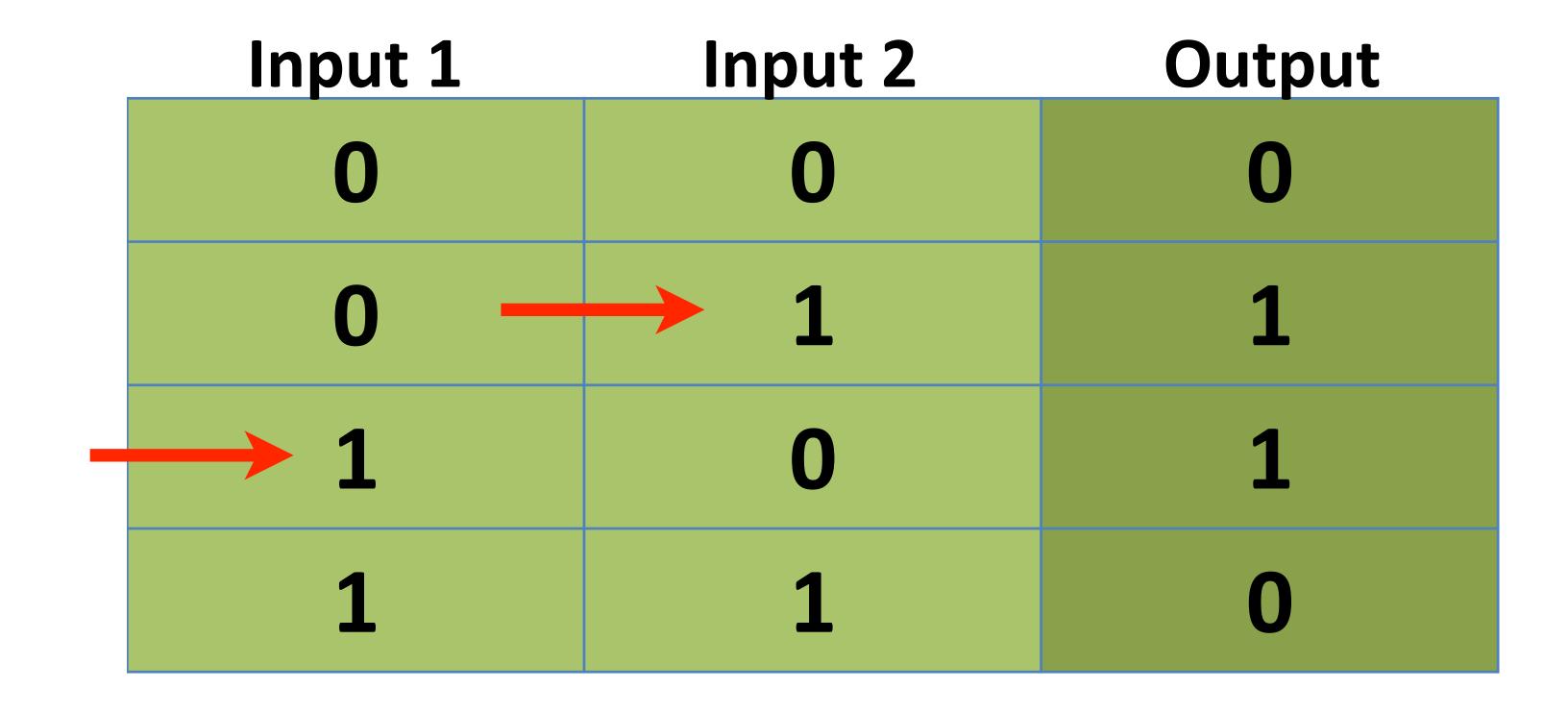
HGTf34\$2

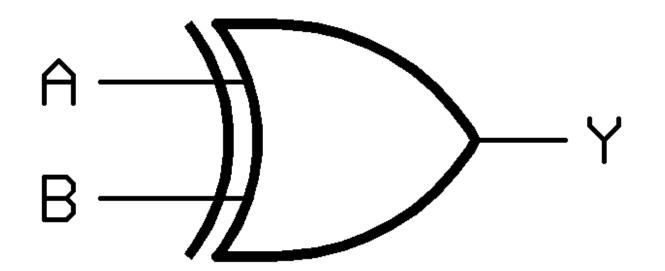
## Use XOR Logic Gate for Hash Function

| Input 1 | Input 2 | Output |
|---------|---------|--------|
| 0       | 0       | 0      |
| 0       | 1       | 1      |
| 1       | 0       | 1      |
| 1       | 1       | 0      |

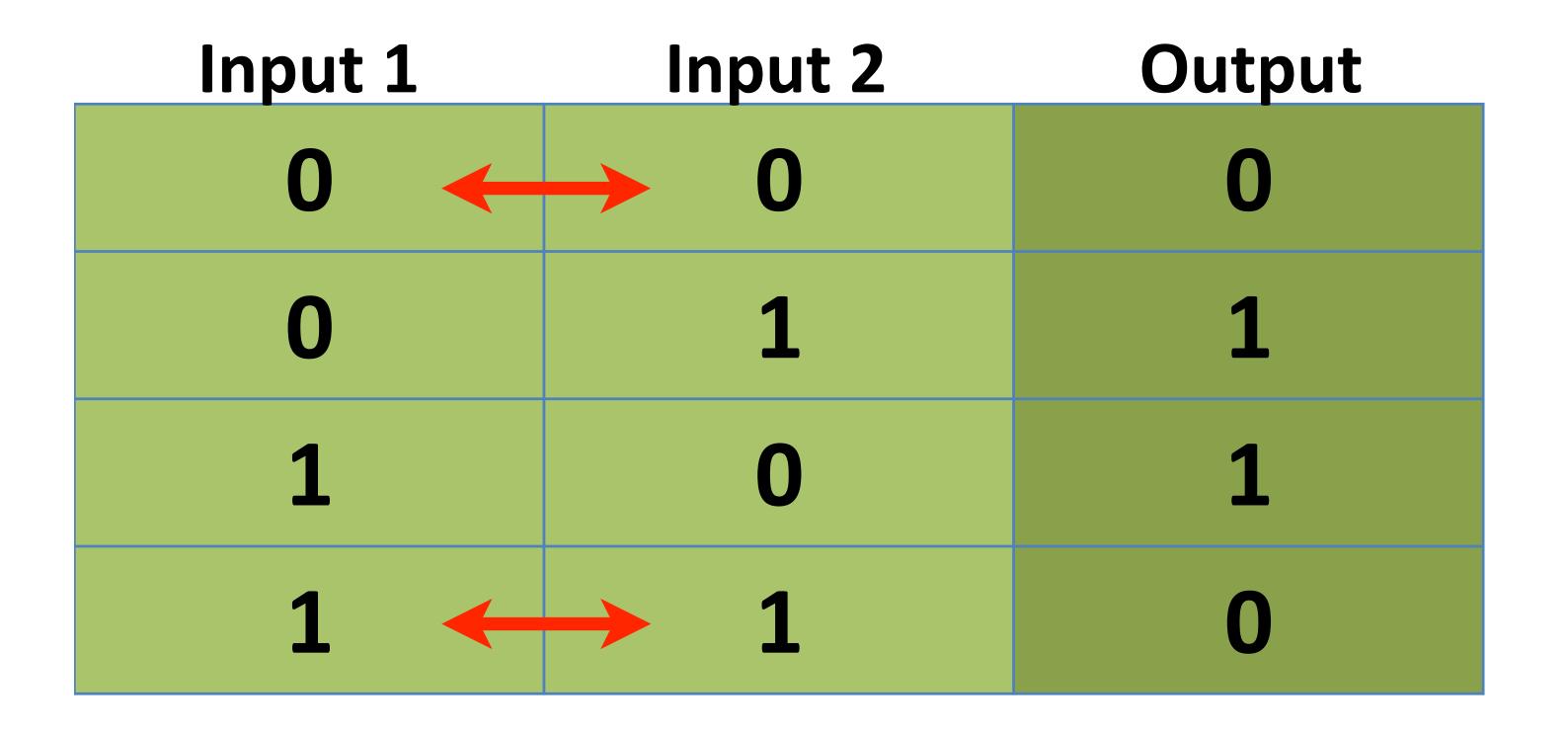


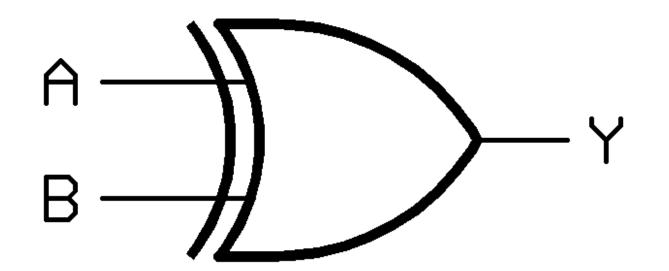
## Use XOR Logic Gate for Hash Function





## Use XOR Logic Gate for Hash Function

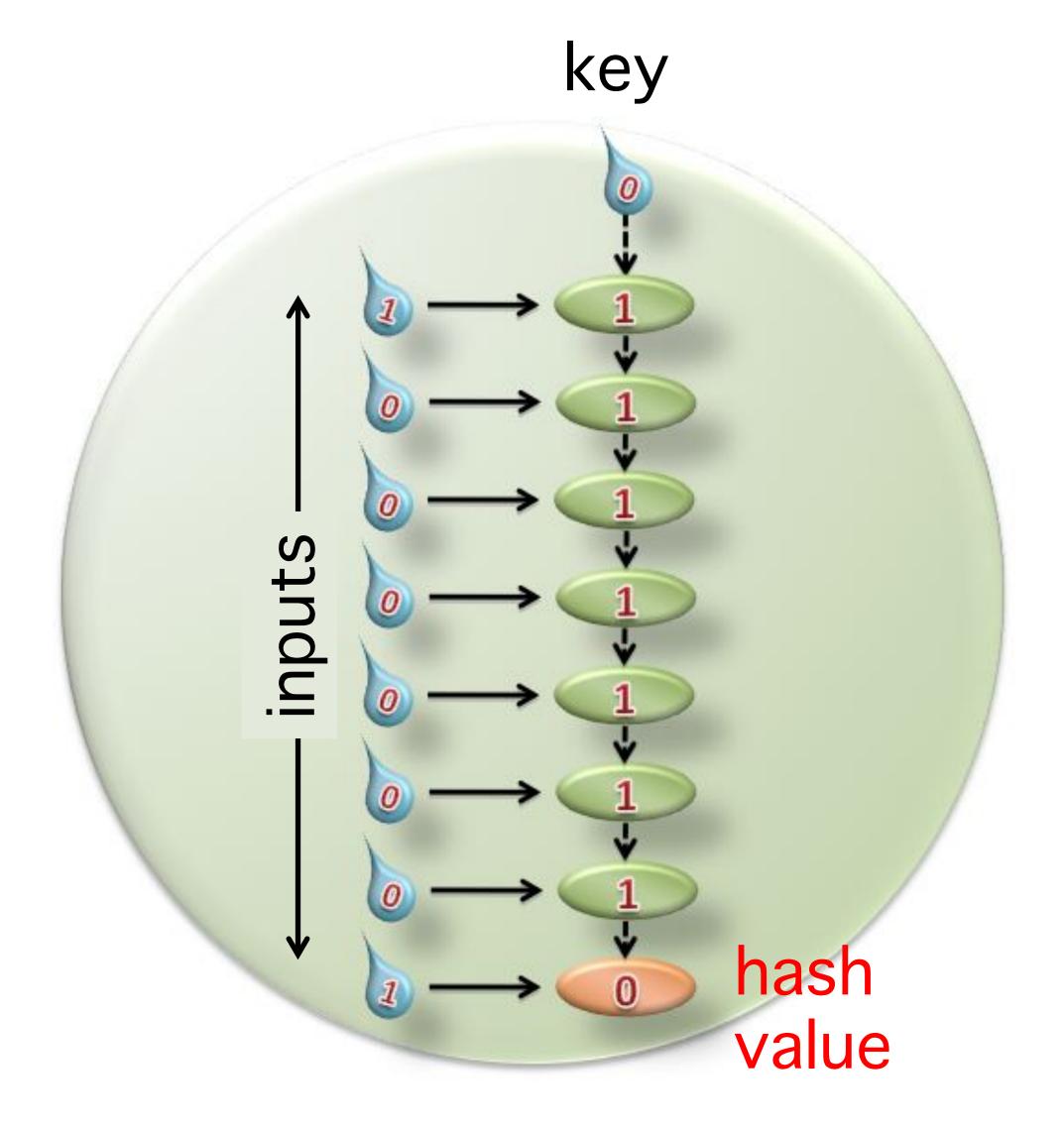




## Design Linear Bacterial Hash Function

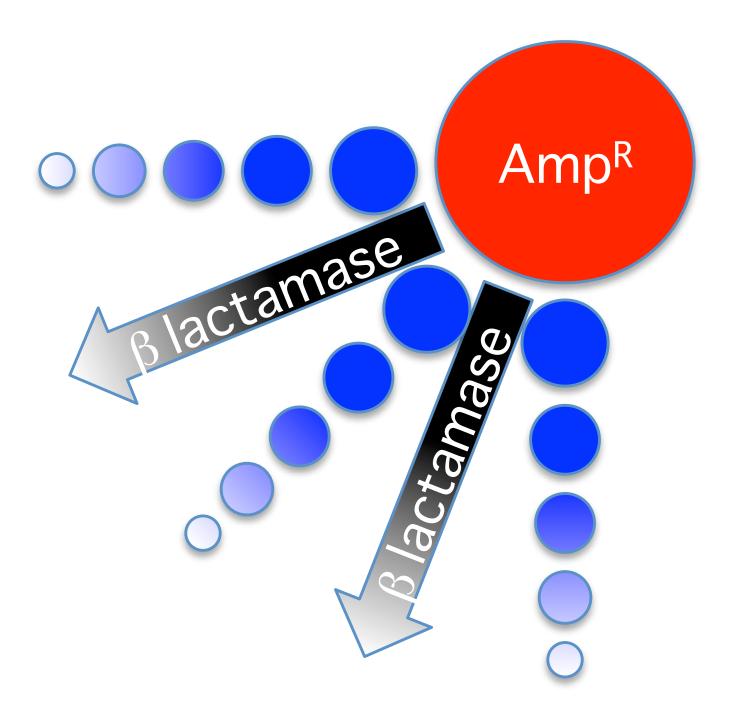
CAB = 010000001

**HASH VALUE = 0** 

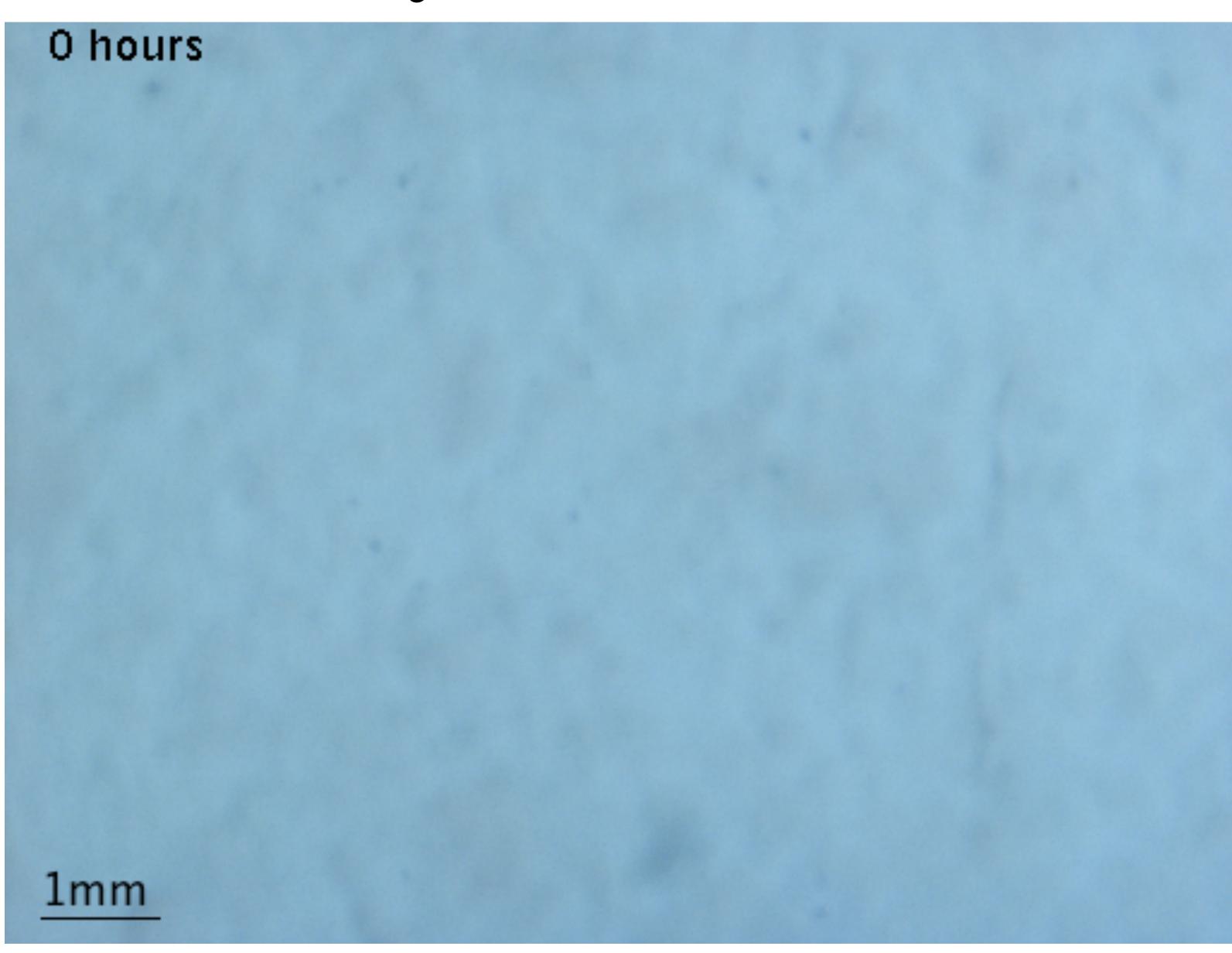


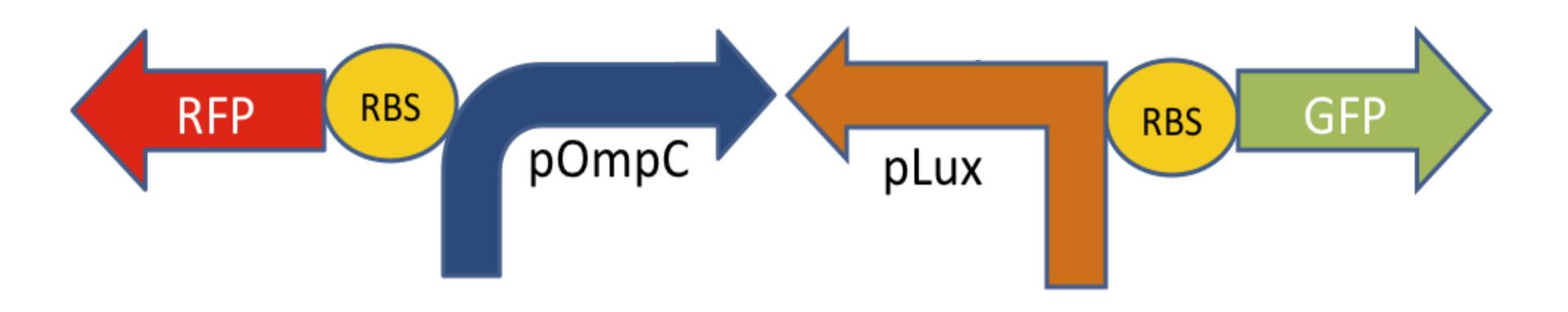
# Time-Delayed Bacterial Growth

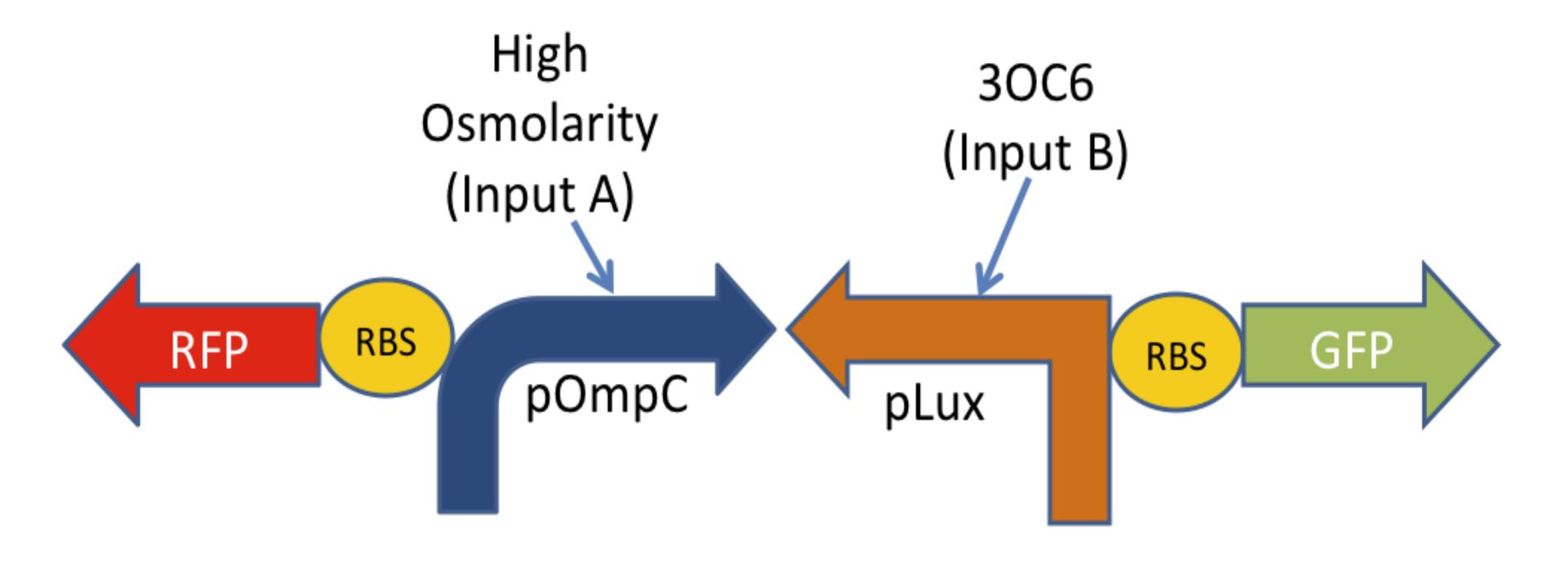


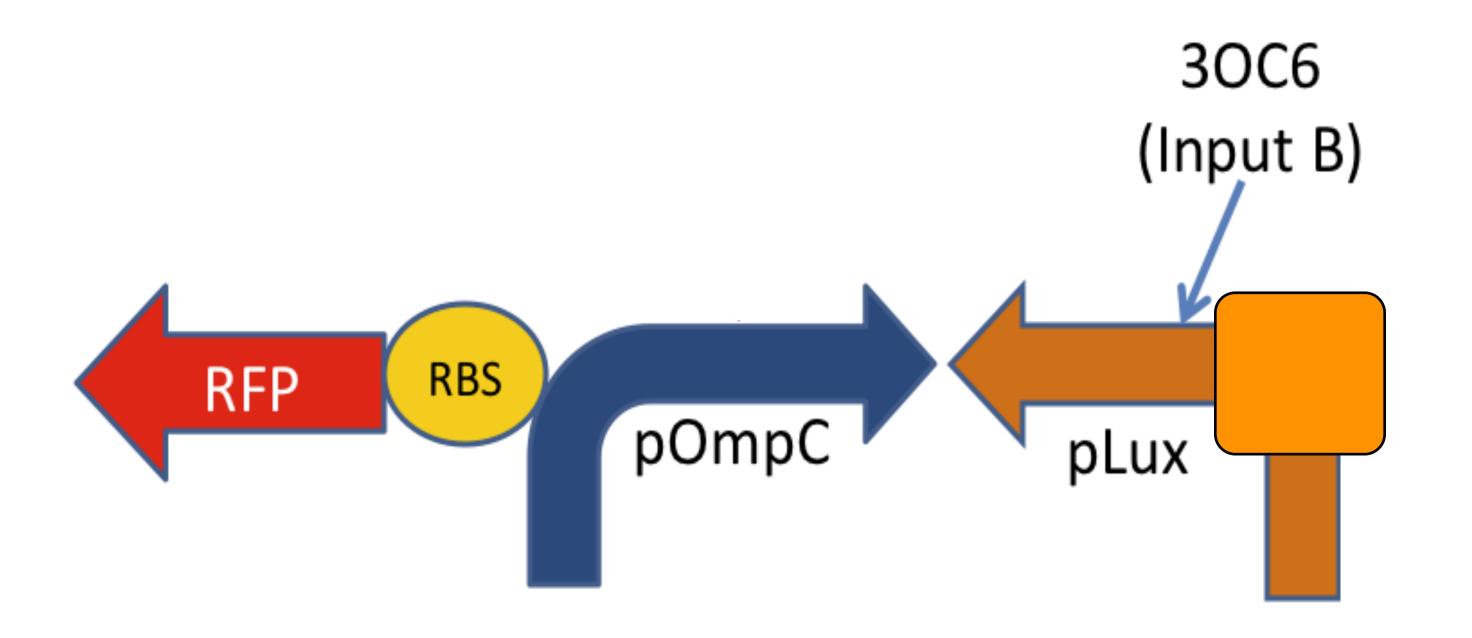


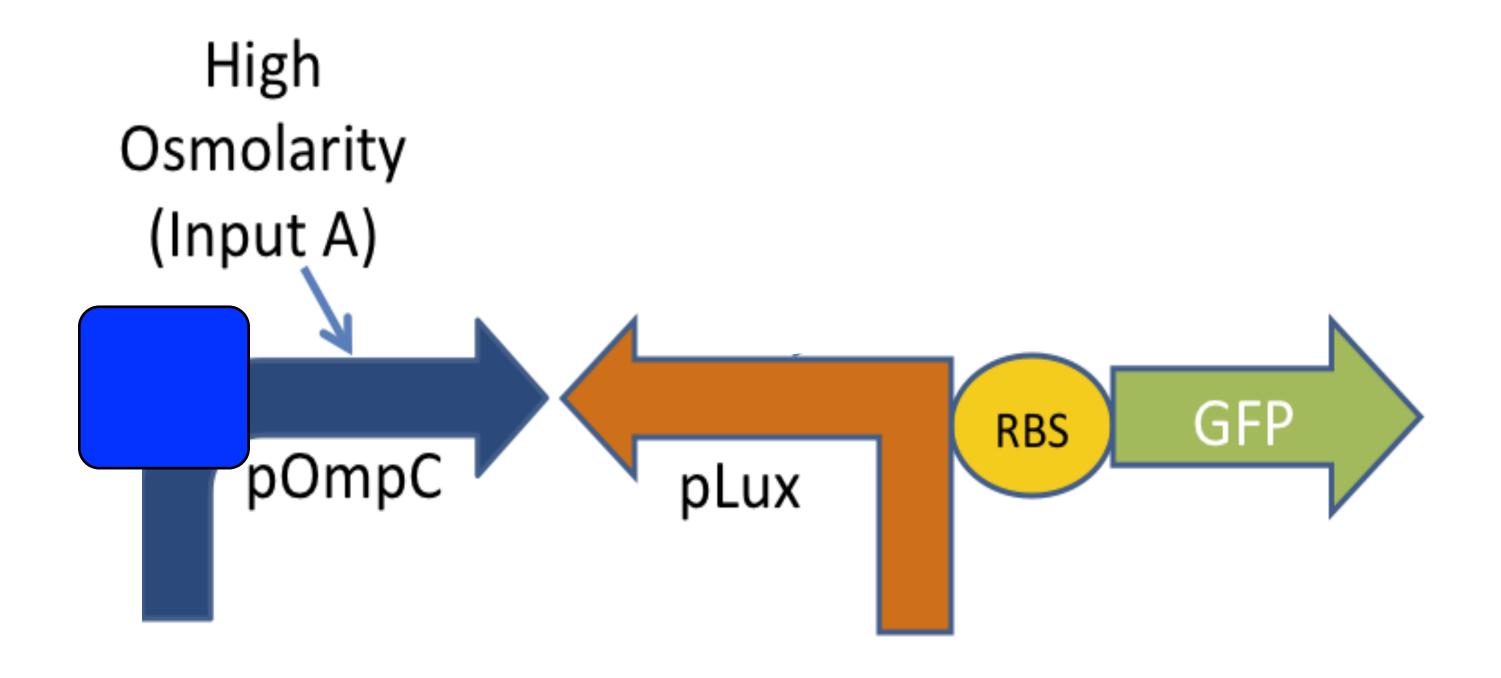
# Time-Delayed Bacterial Growth

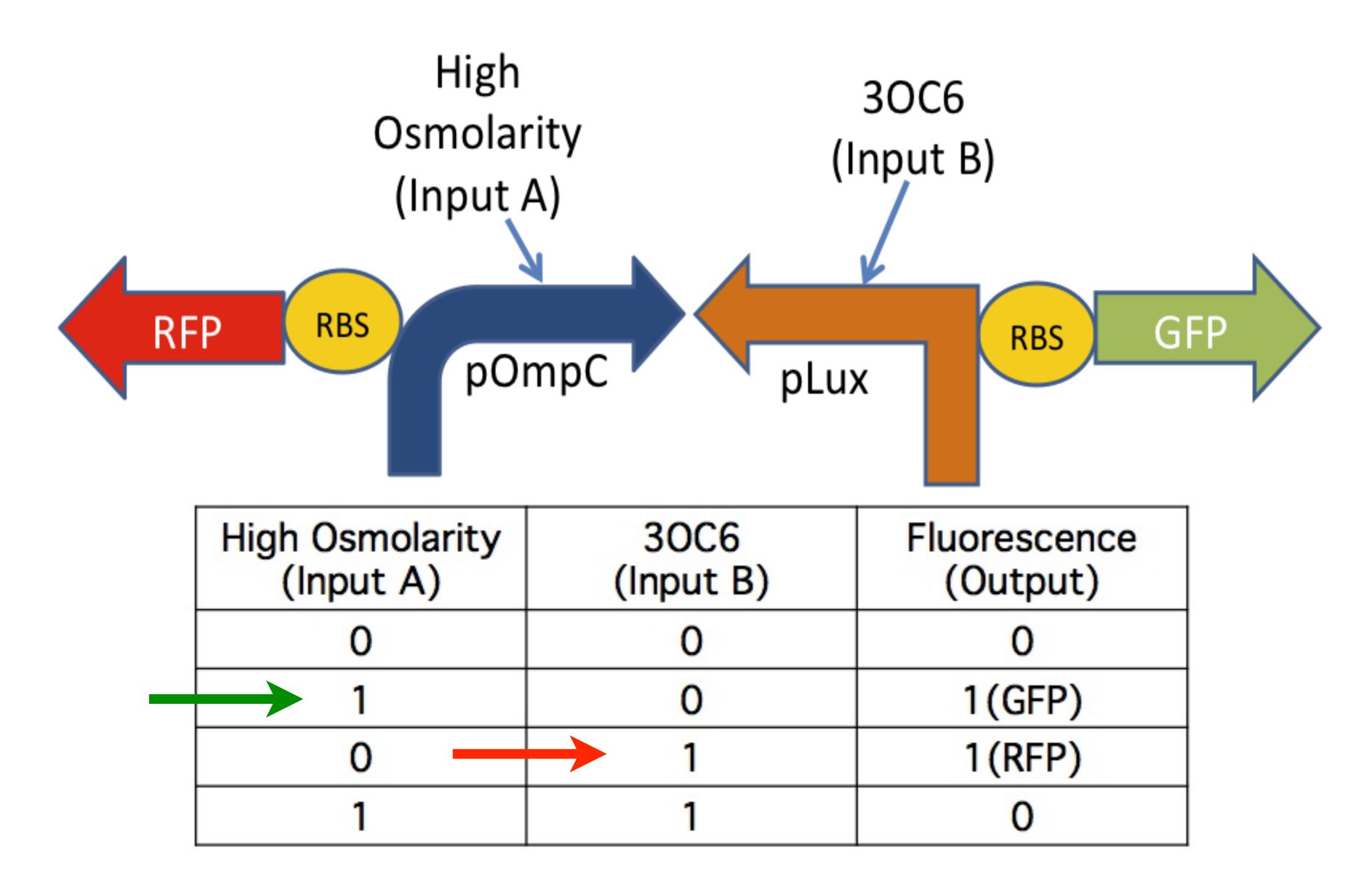


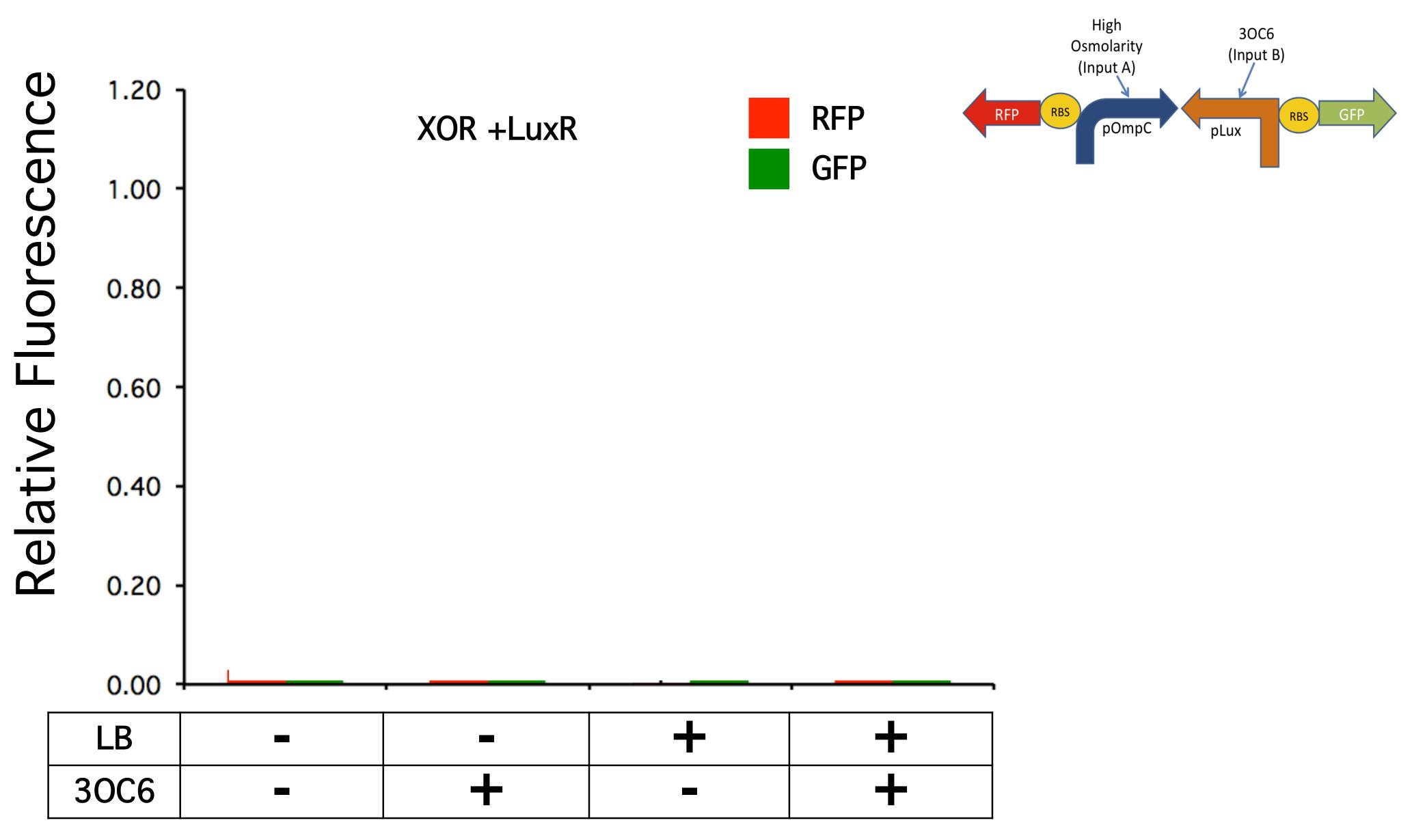


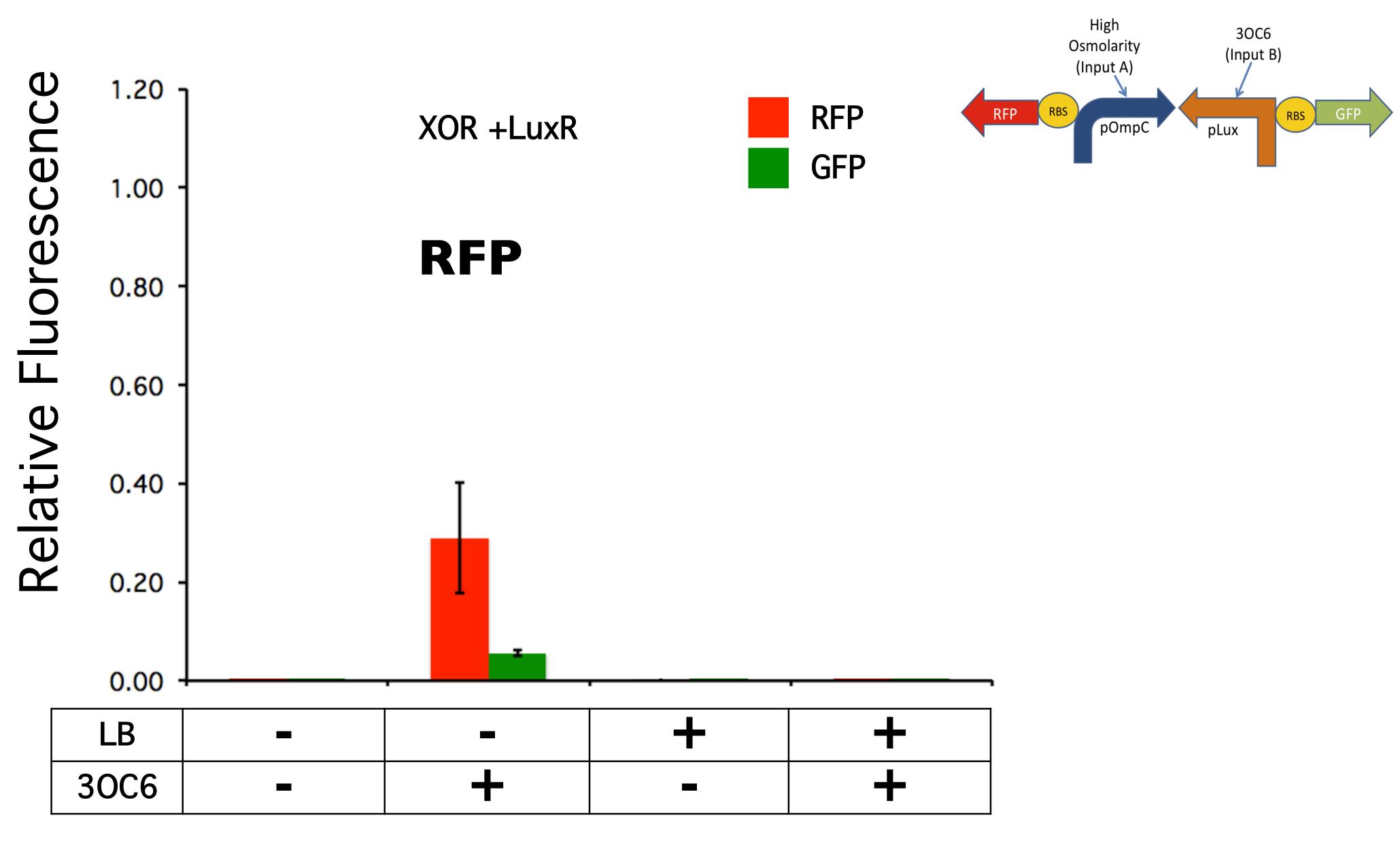


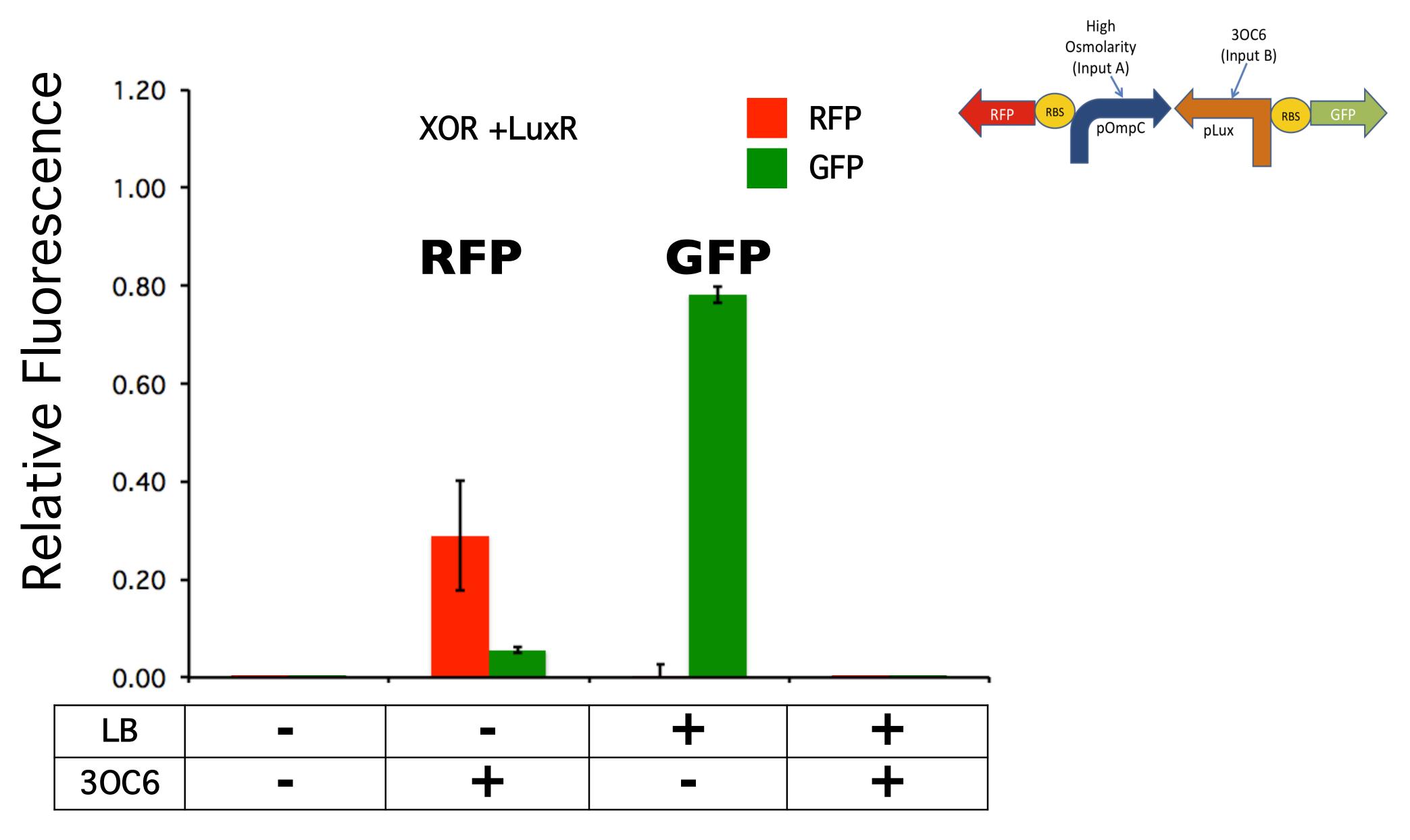


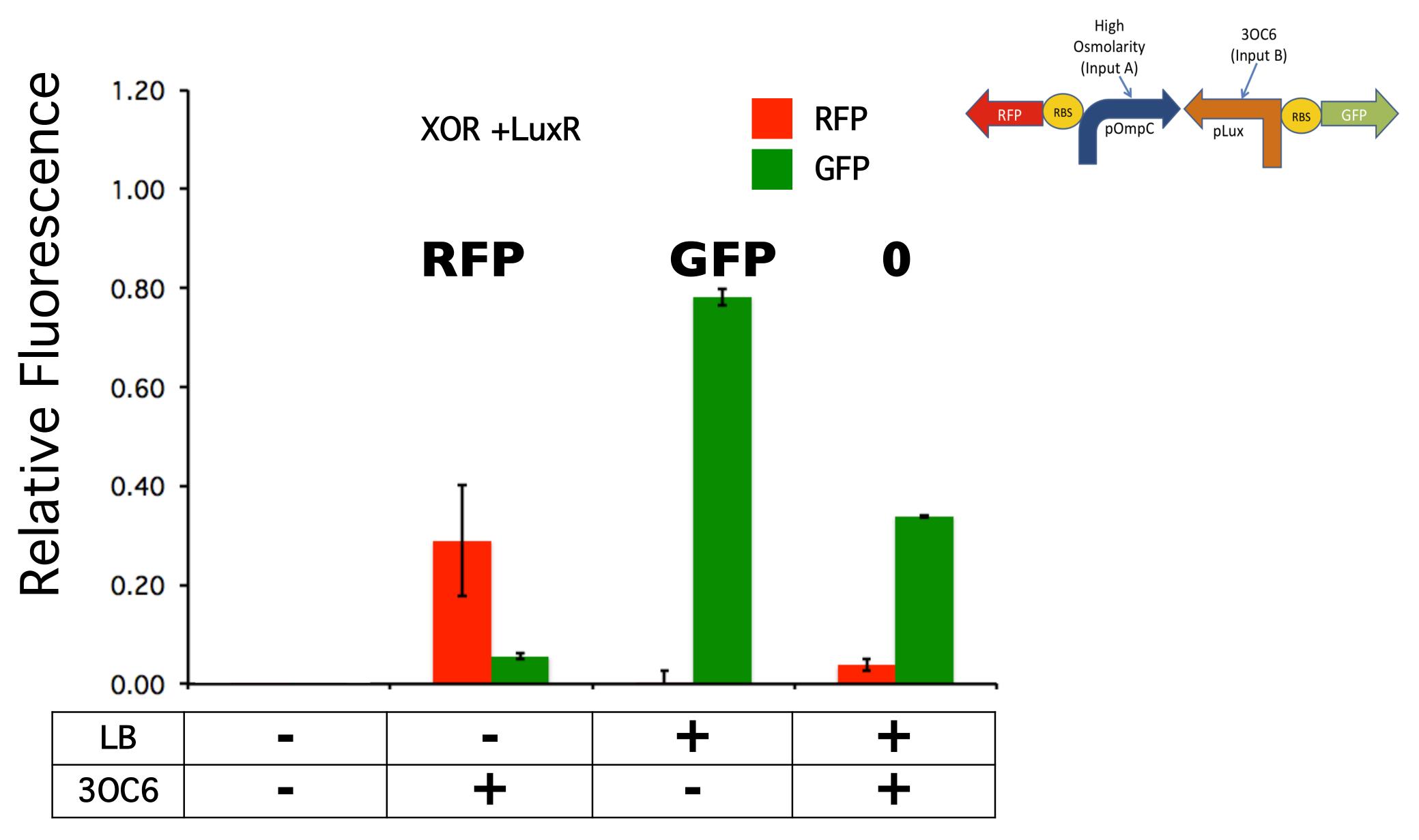


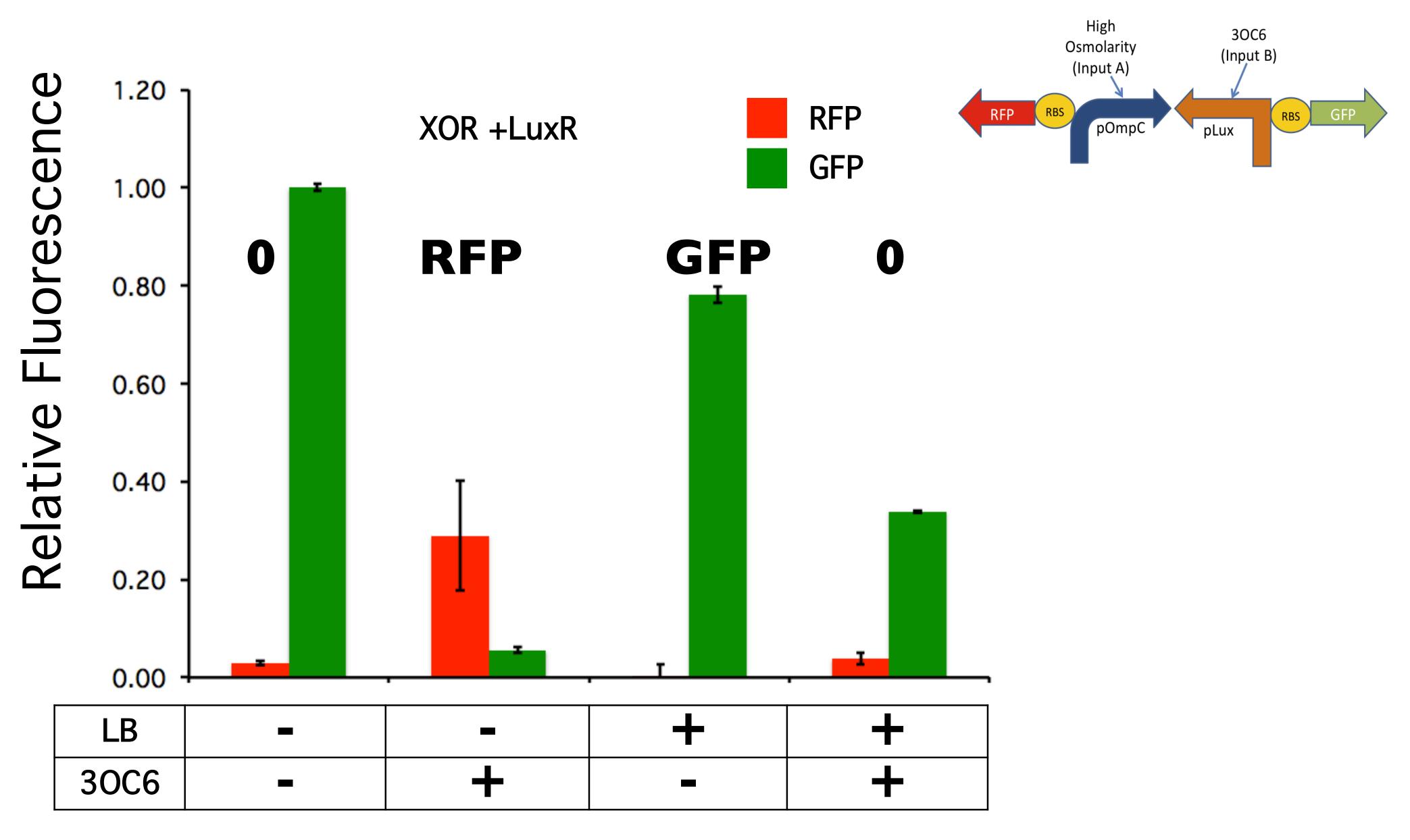


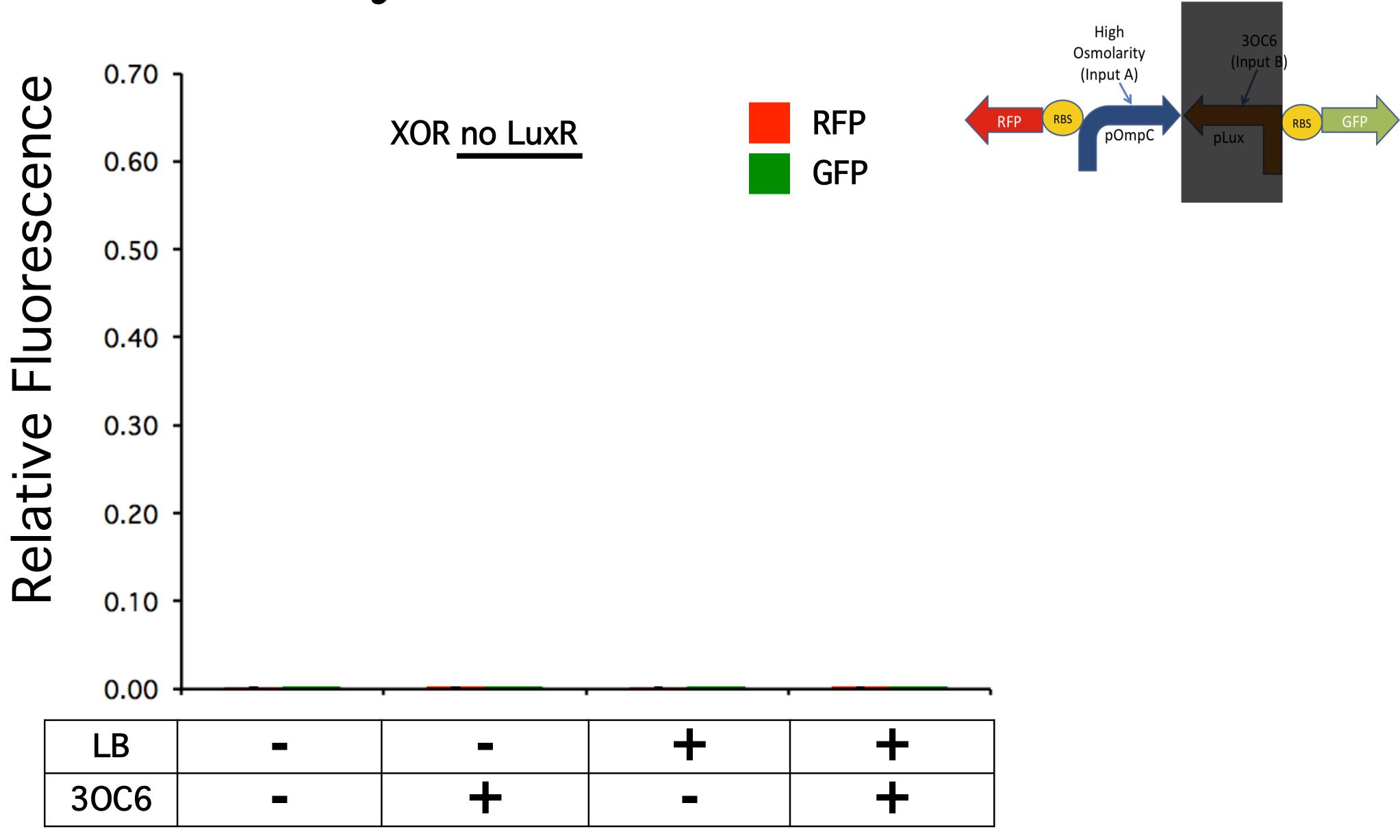


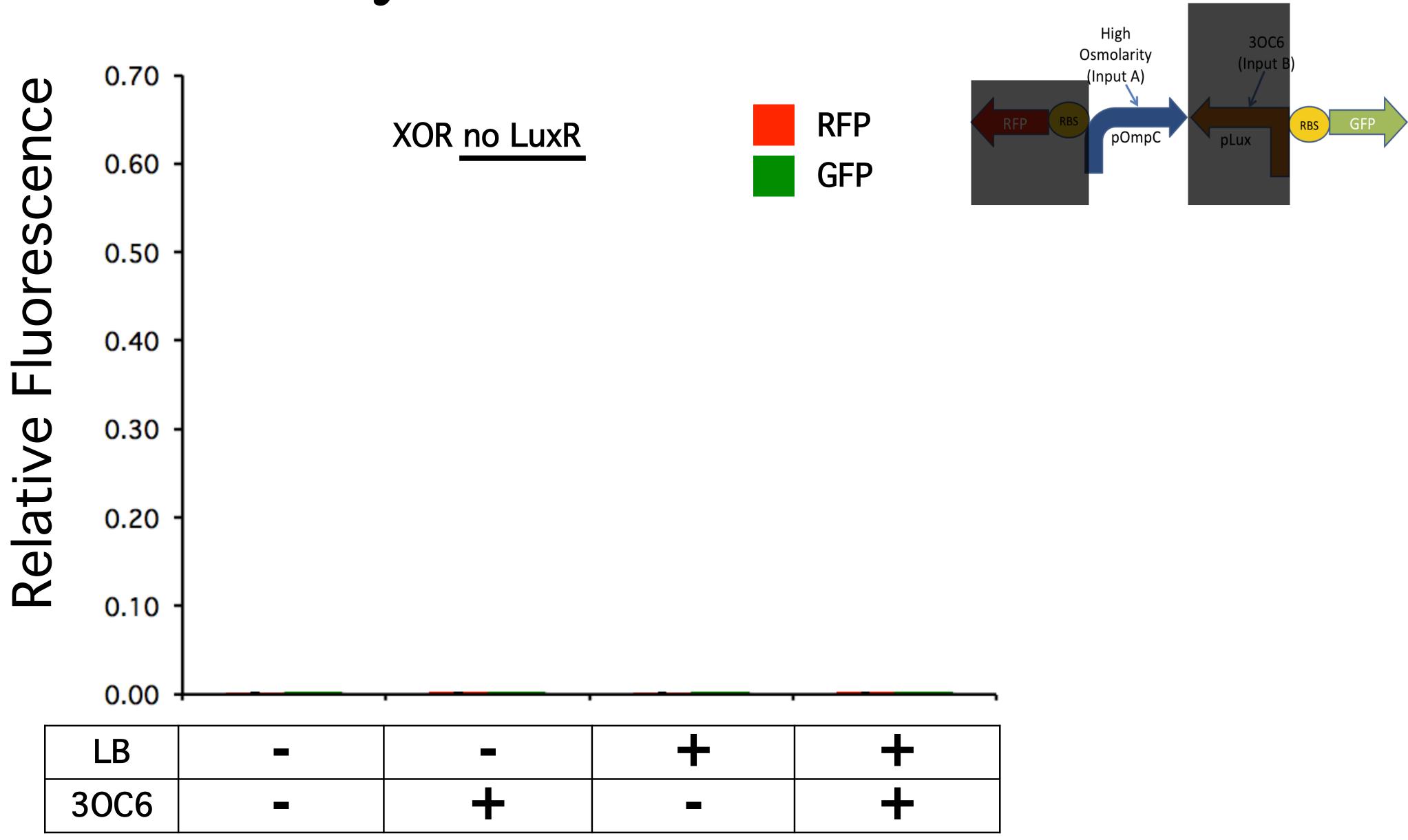


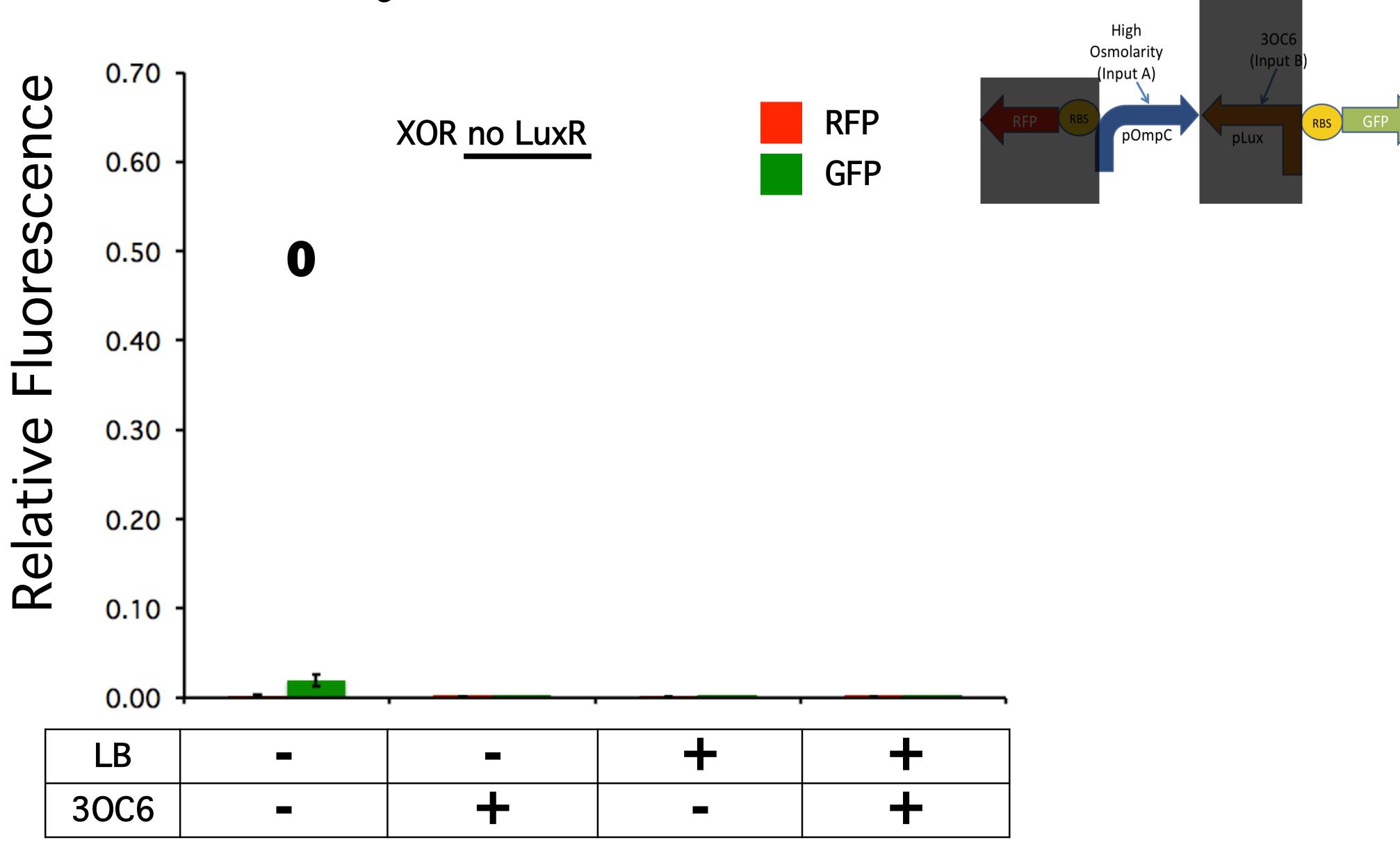


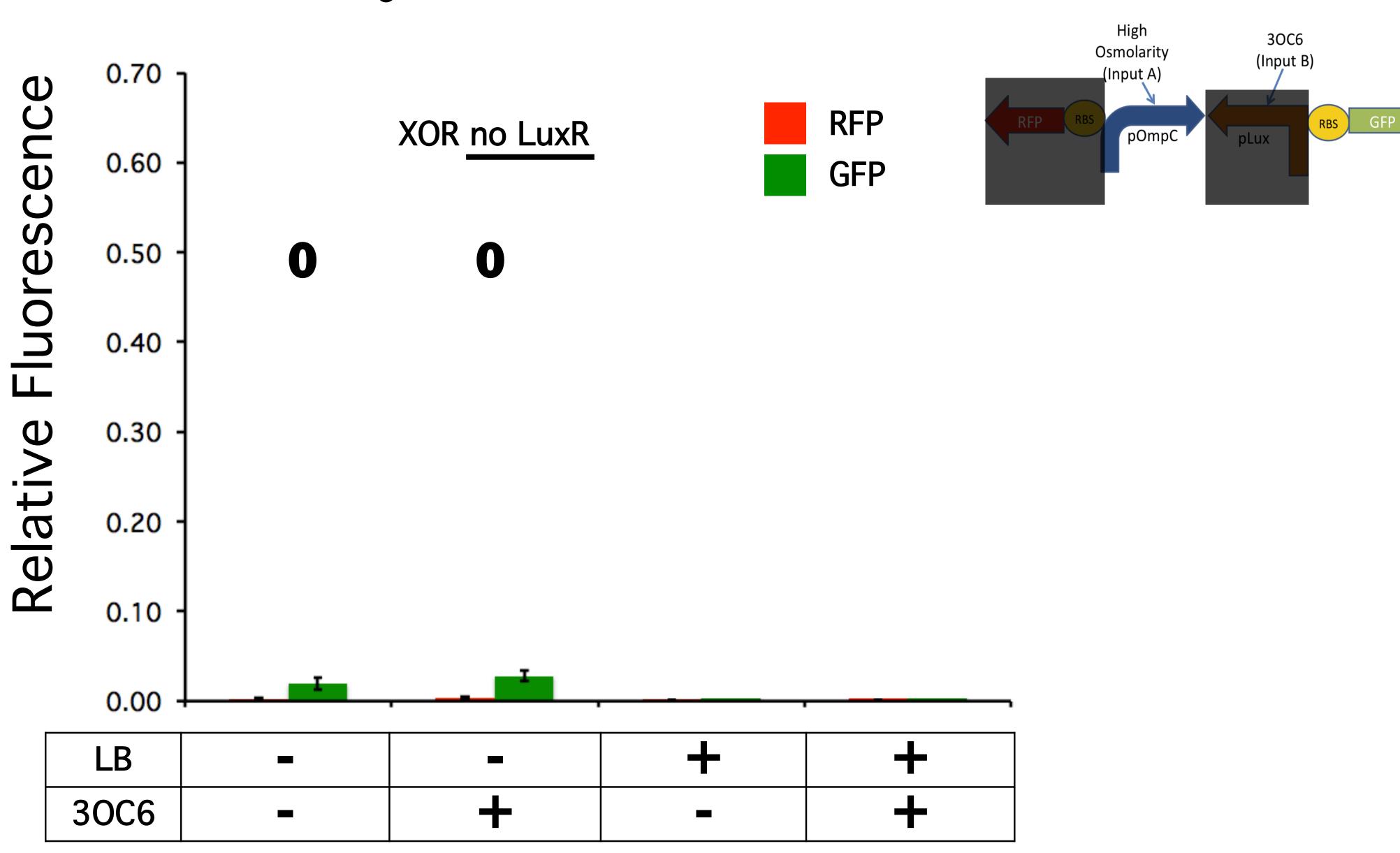


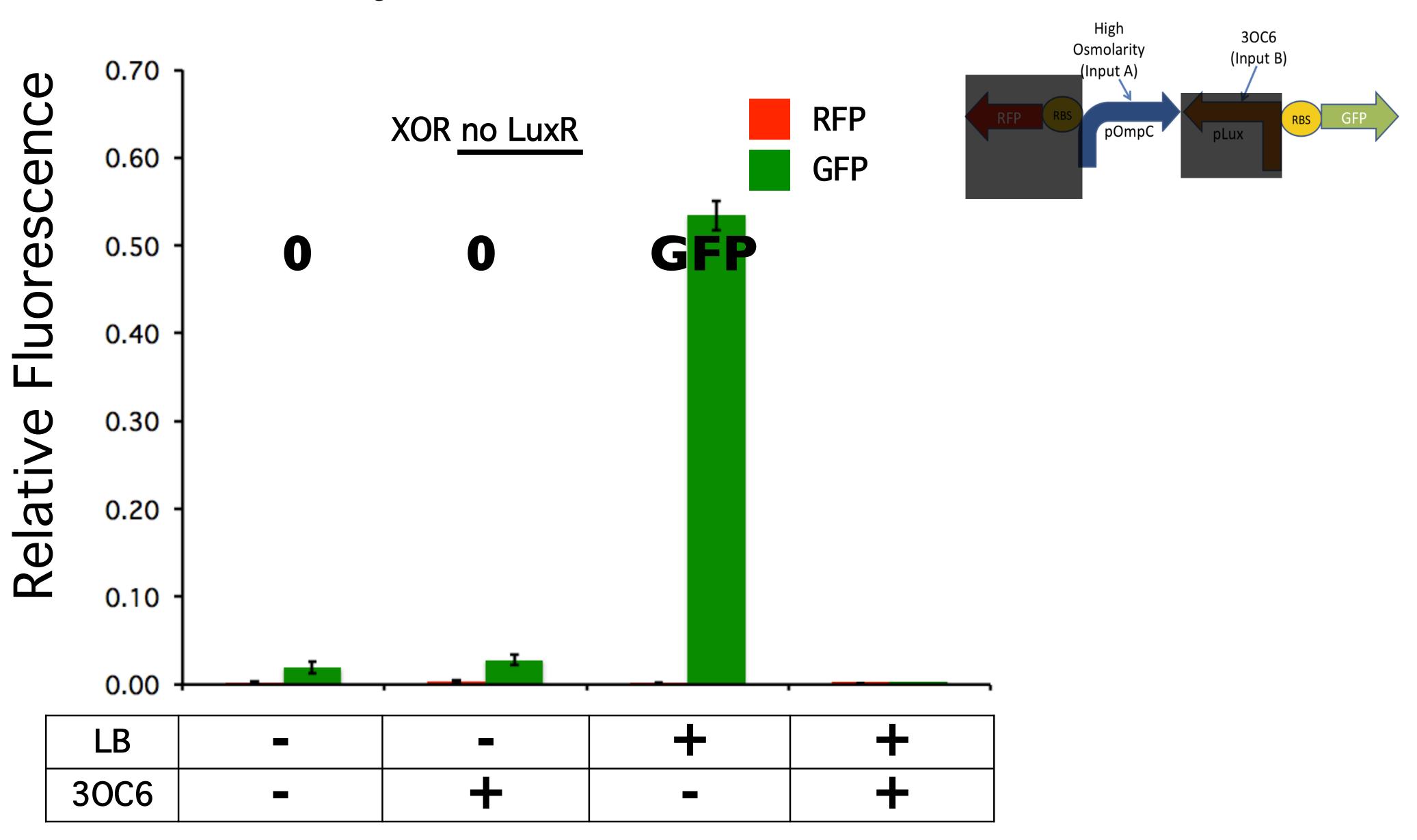


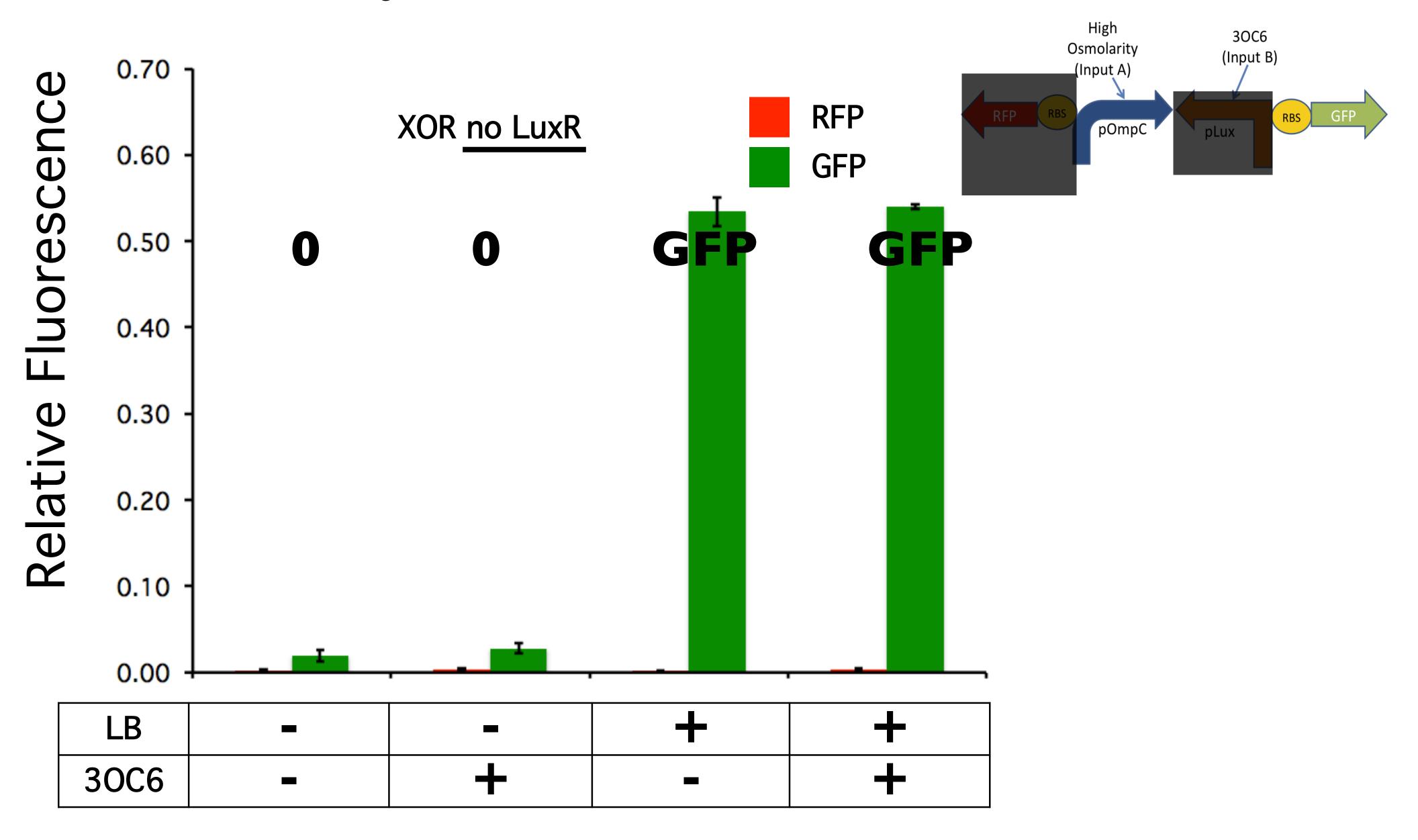


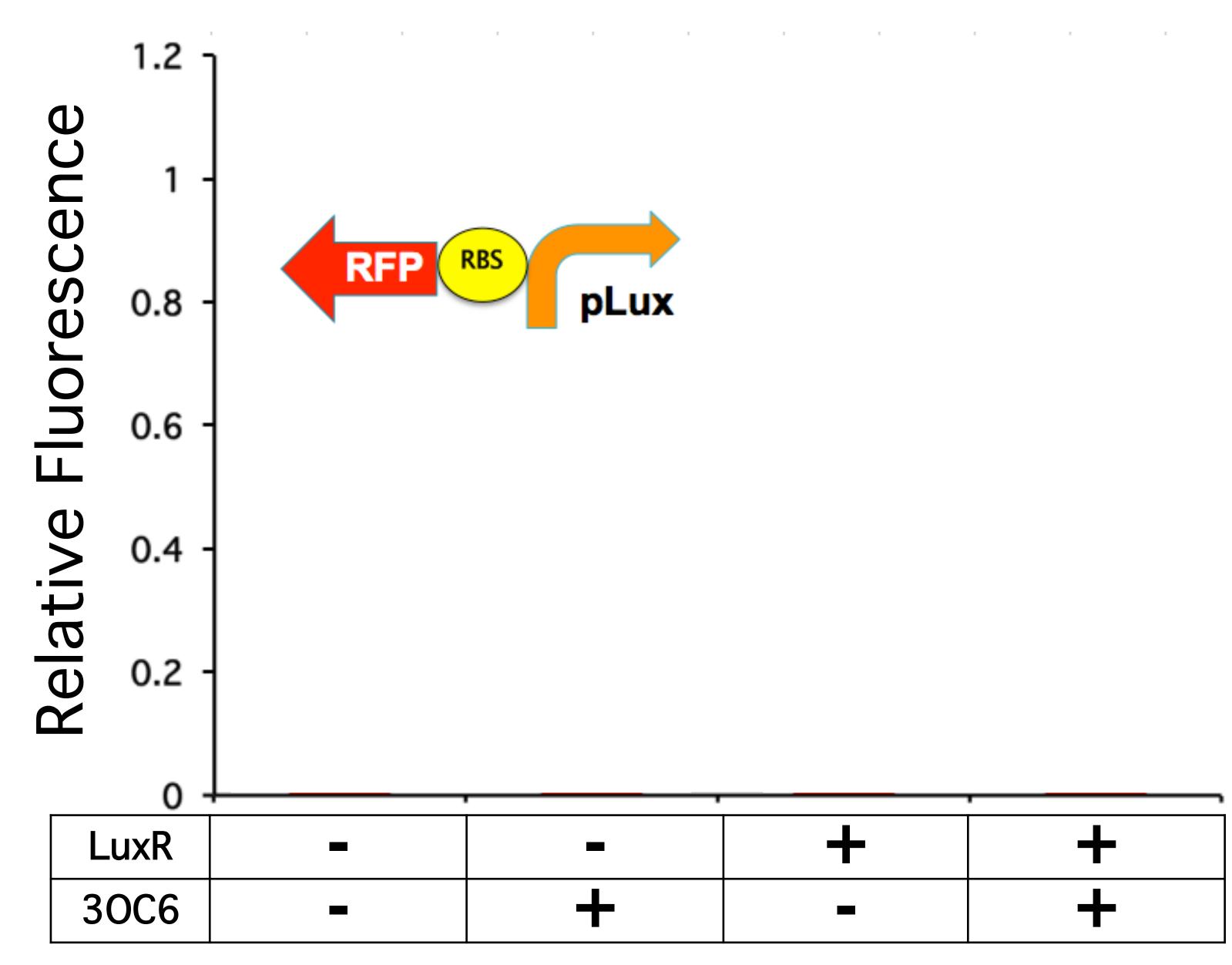


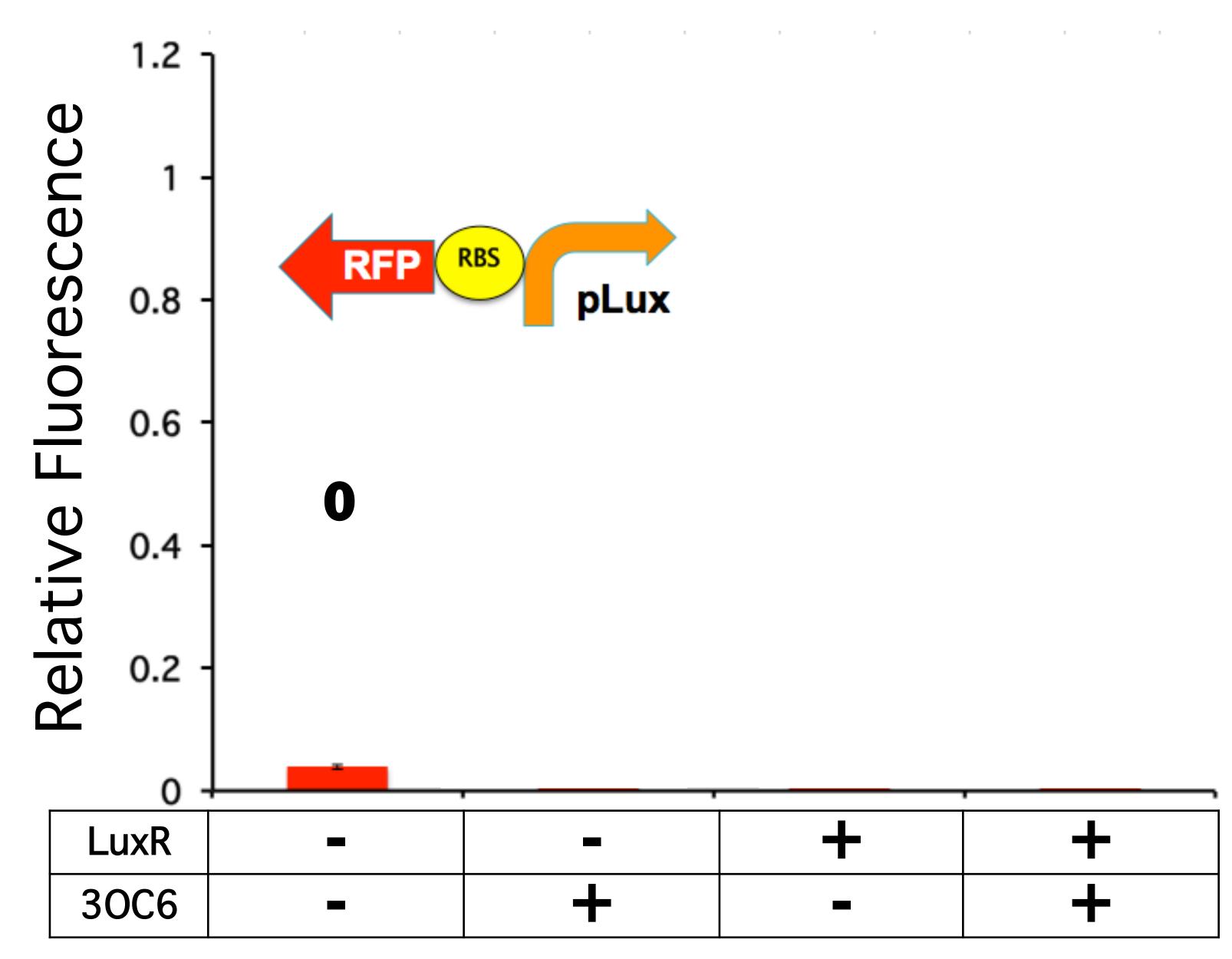


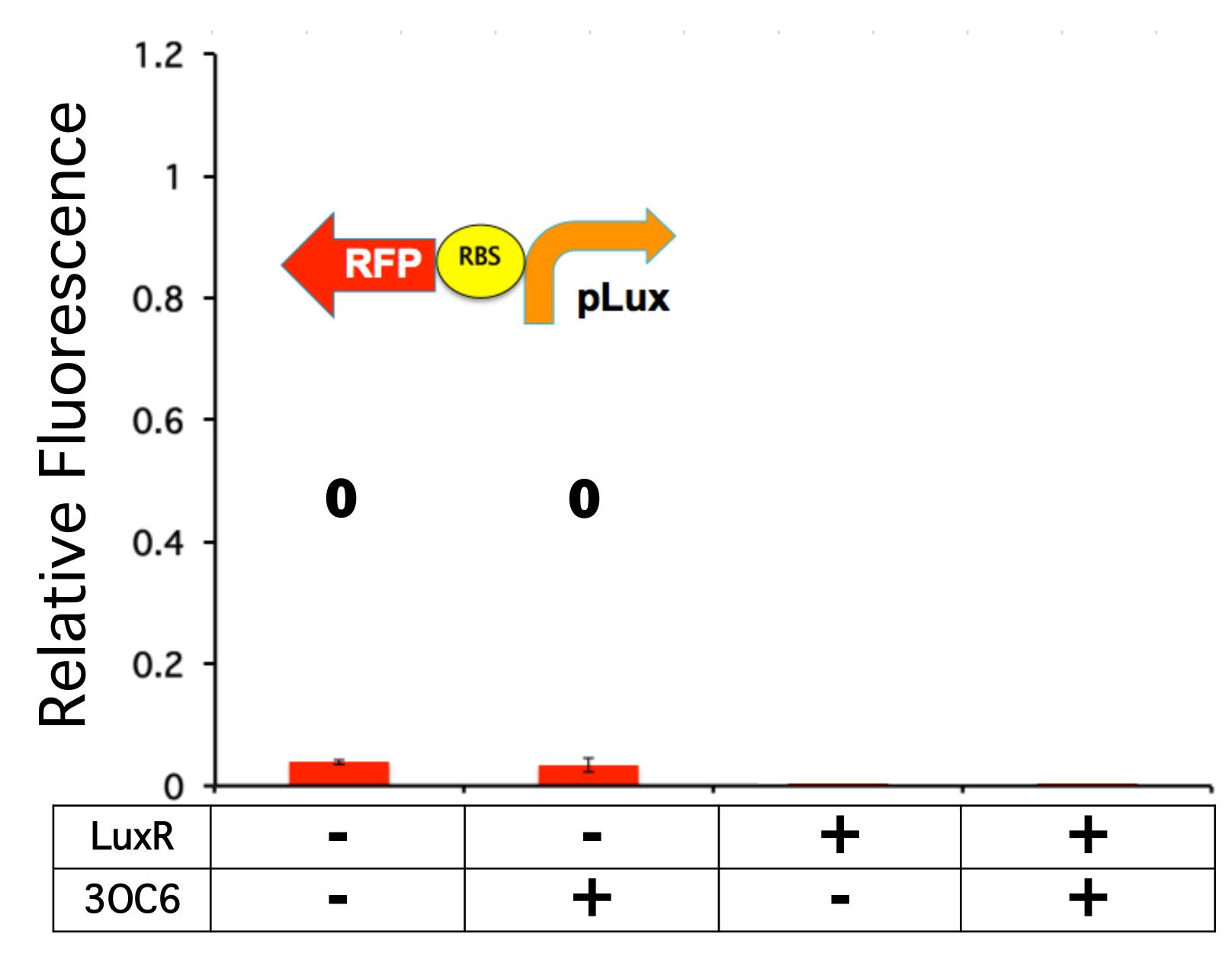


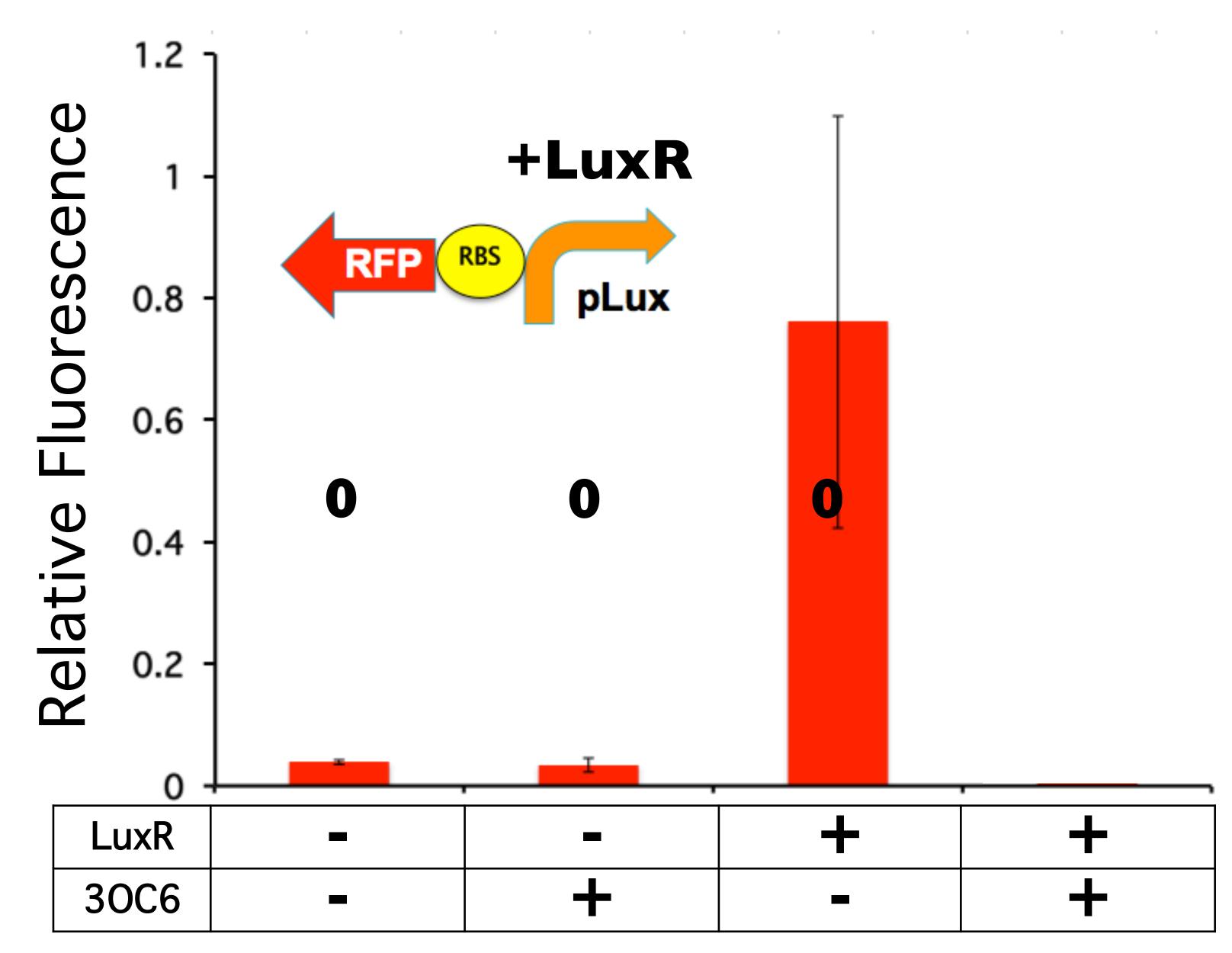


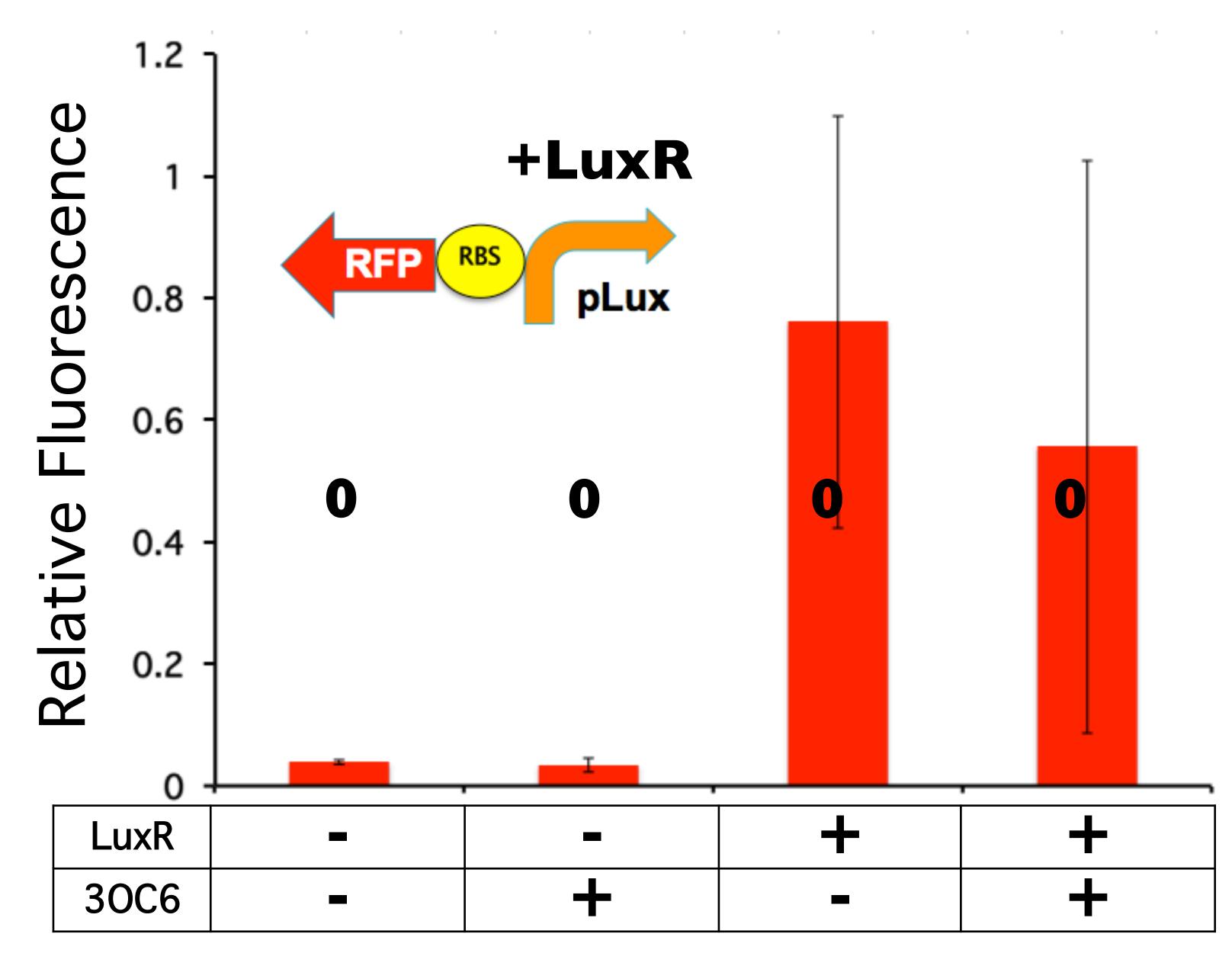






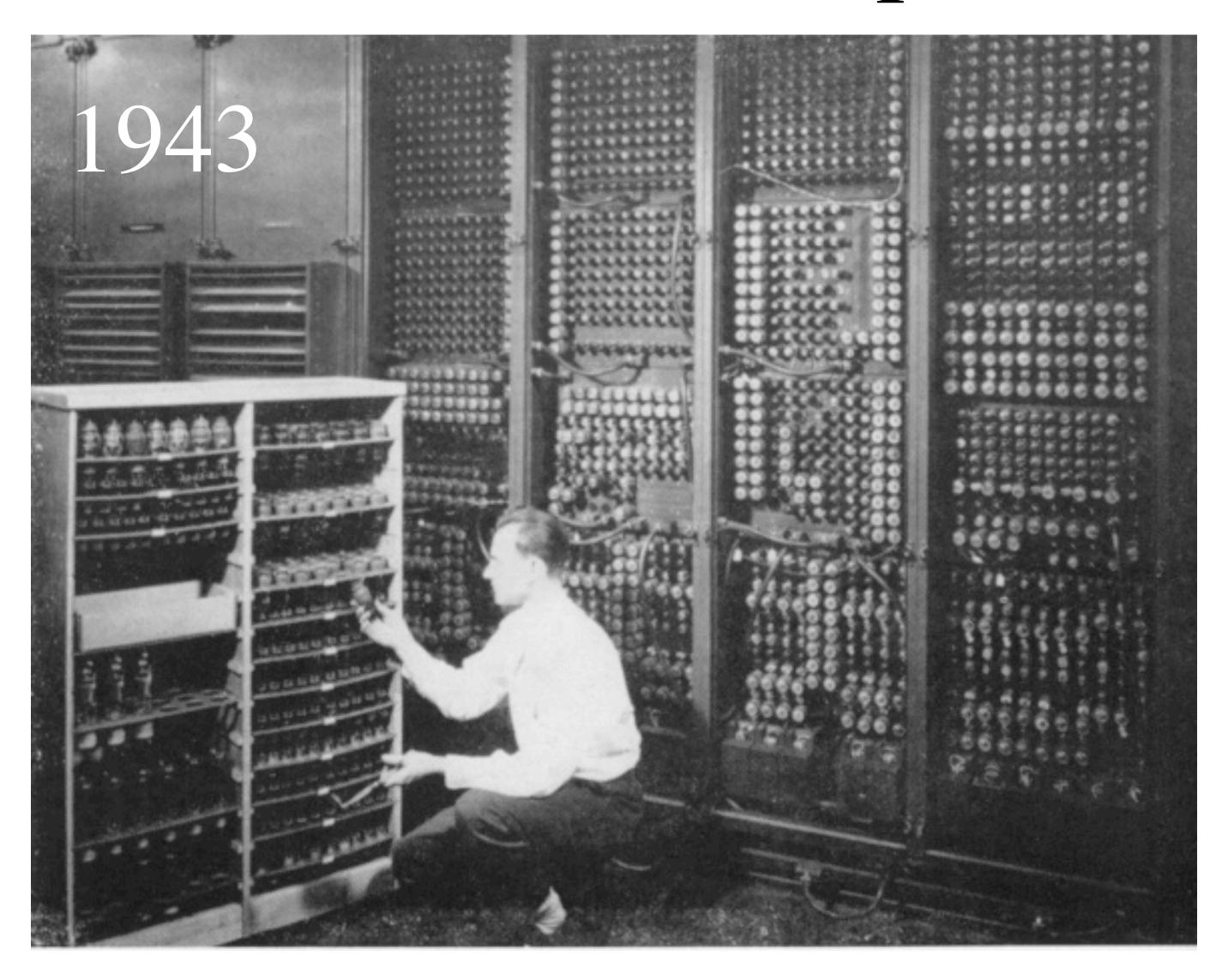






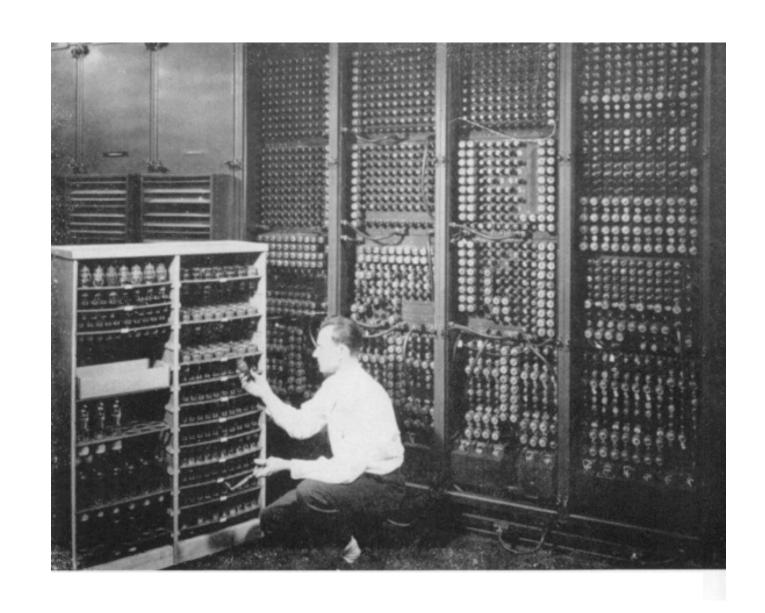
# Why build bacterial computers?

## Evolution of Computers



## Evolution of Computers

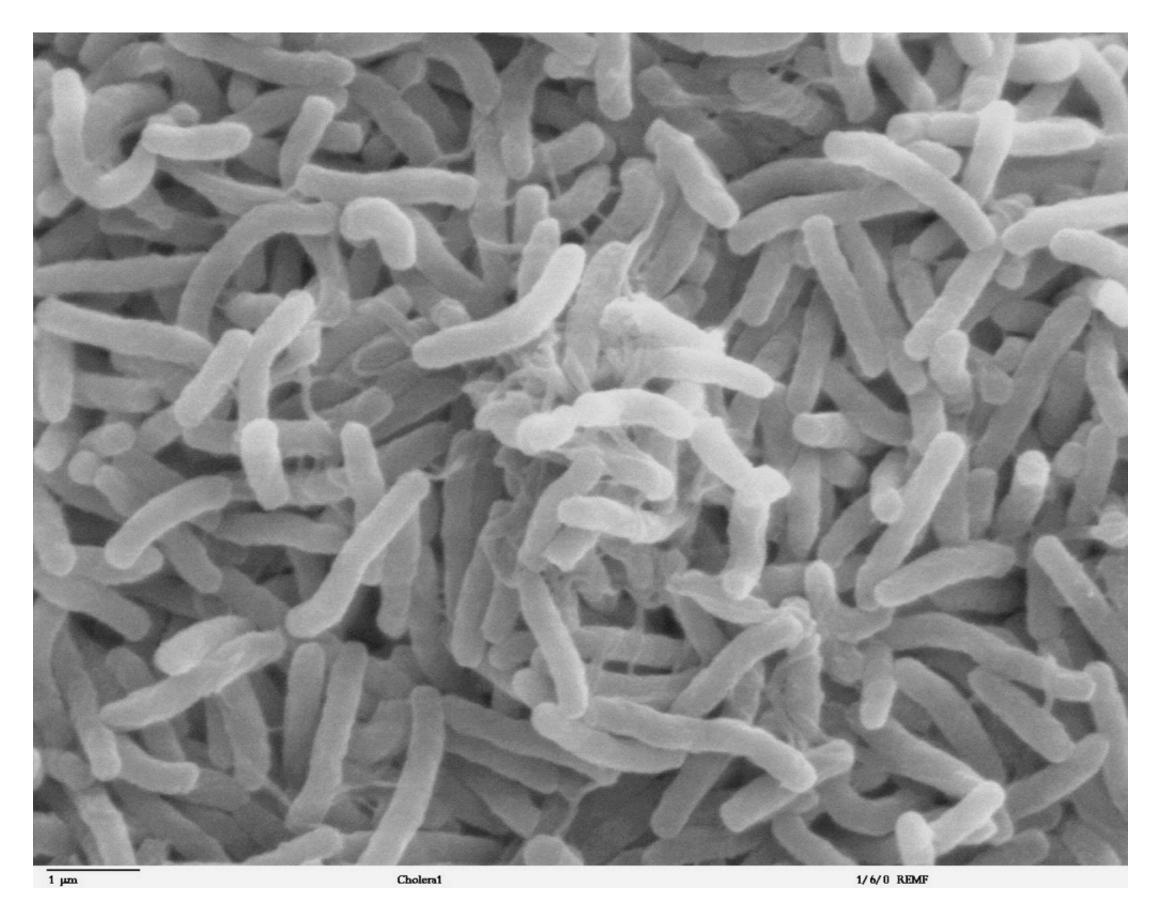
#### iPhone in 2012

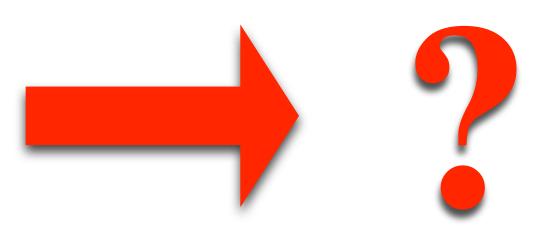




### Evolution of Bacterial Computers

#### E. coli in 2012

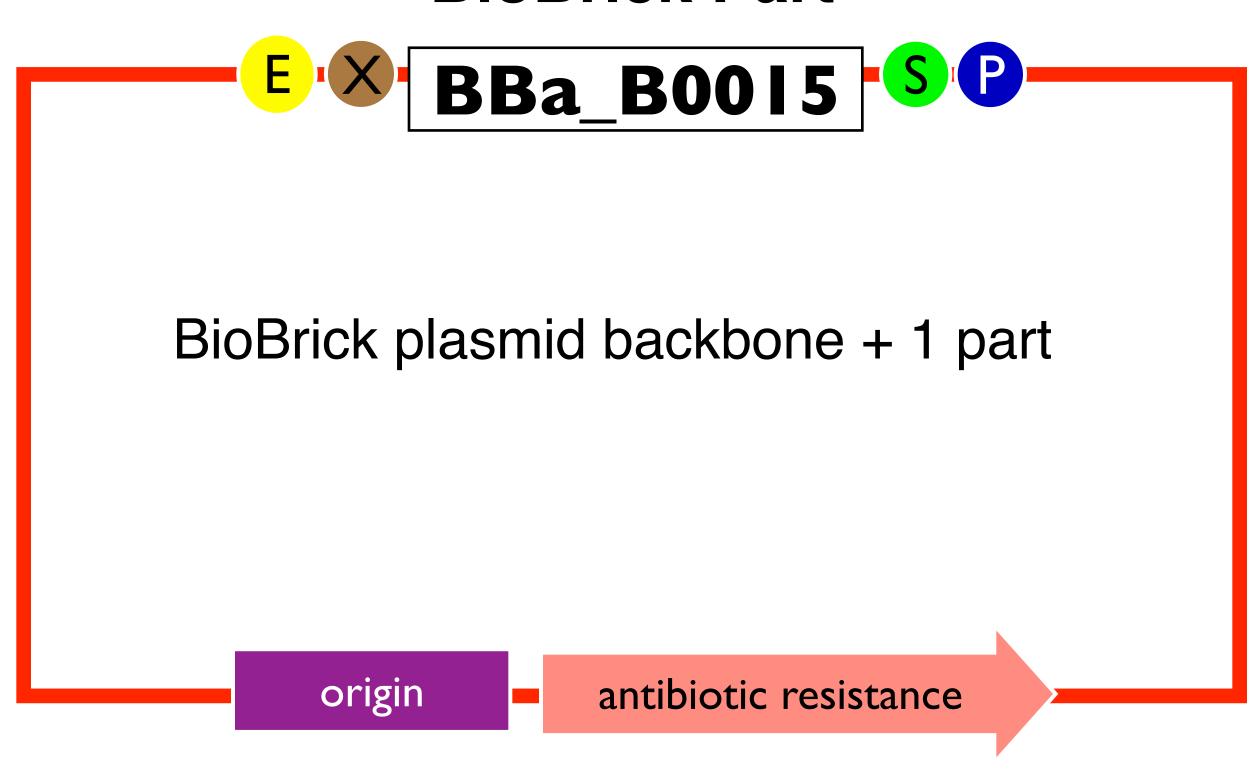




Living Hardware in 2022

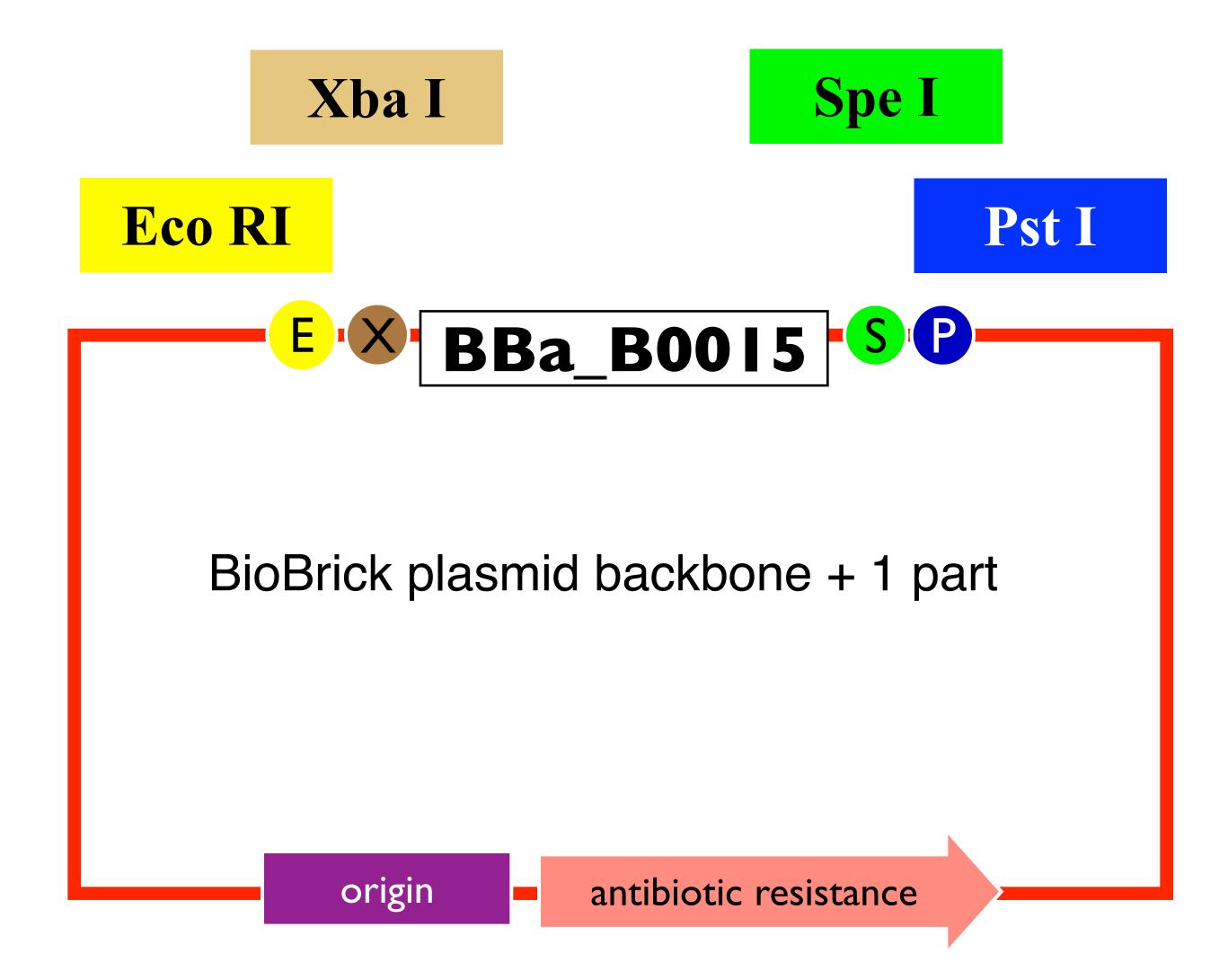
## How do we clone DNA? Can we do this for intro bio??

**BioBrick Part** 

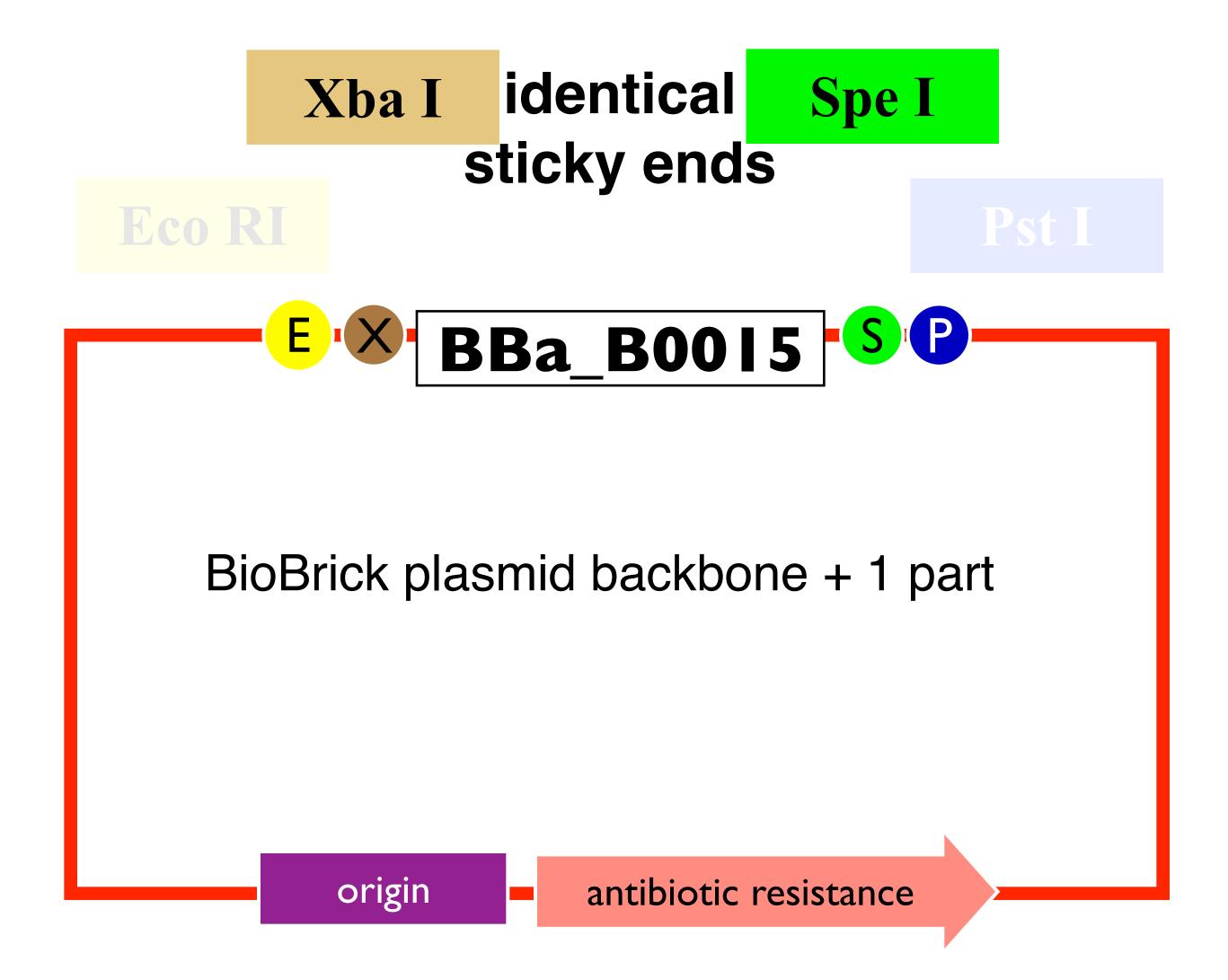


BioBrick plasmid backbone

(http://partsregistry.org/Plasmids)

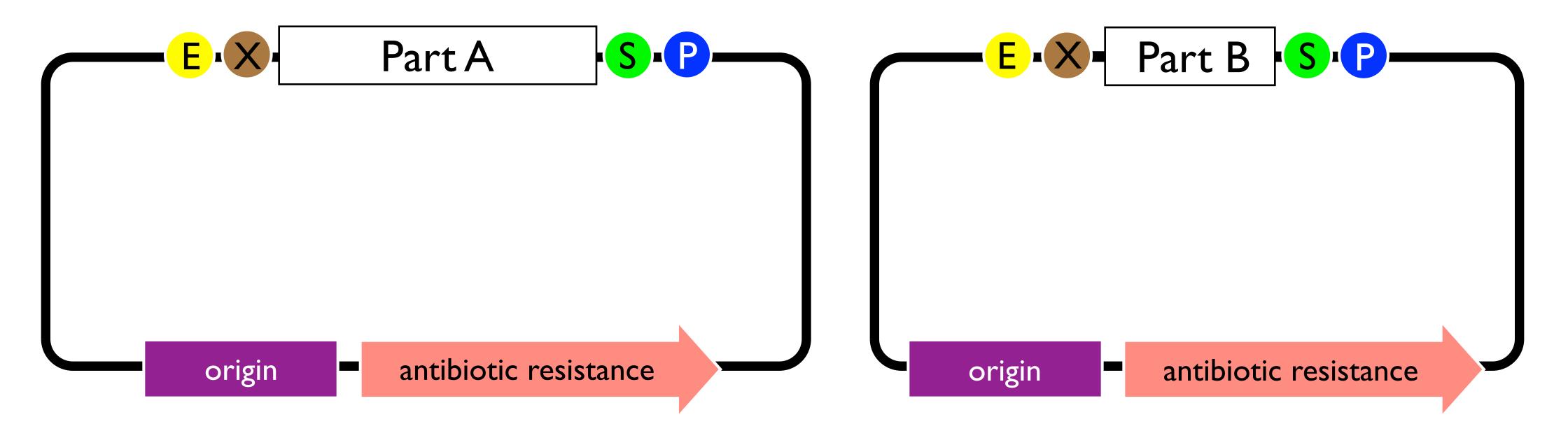


(http://partsregistry.org/Plasmids)



(http://partsregistry.org/Plasmids)

#### put B downstream of A



cut with Spe and Pst

E-X-Part A

Part B

Part B

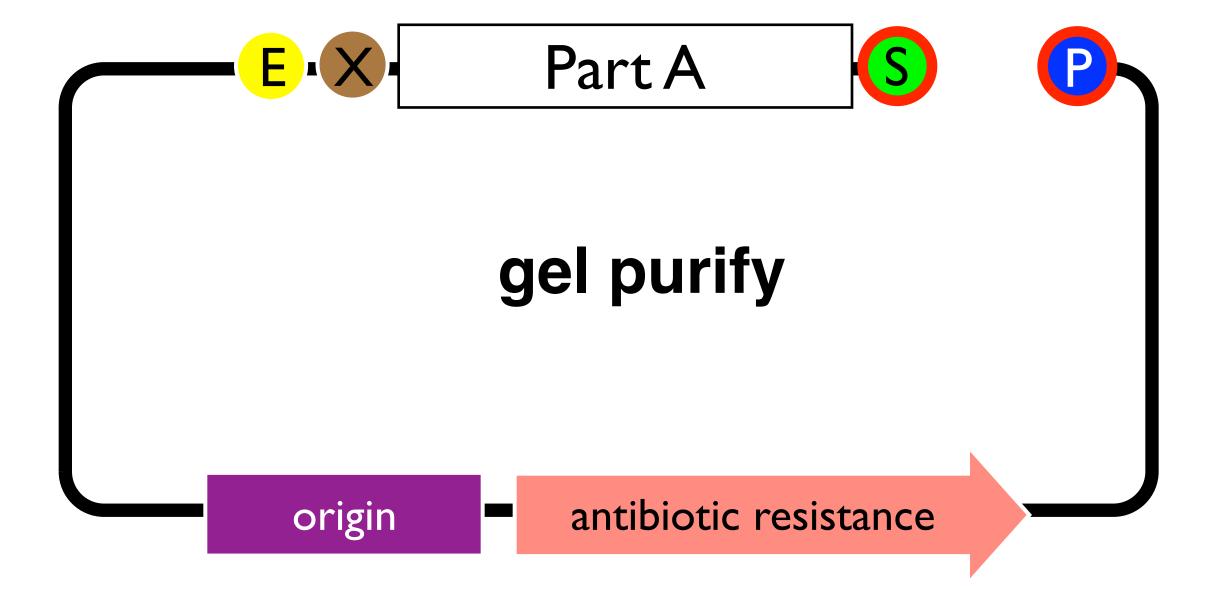
Part B

origin

antibiotic resistance

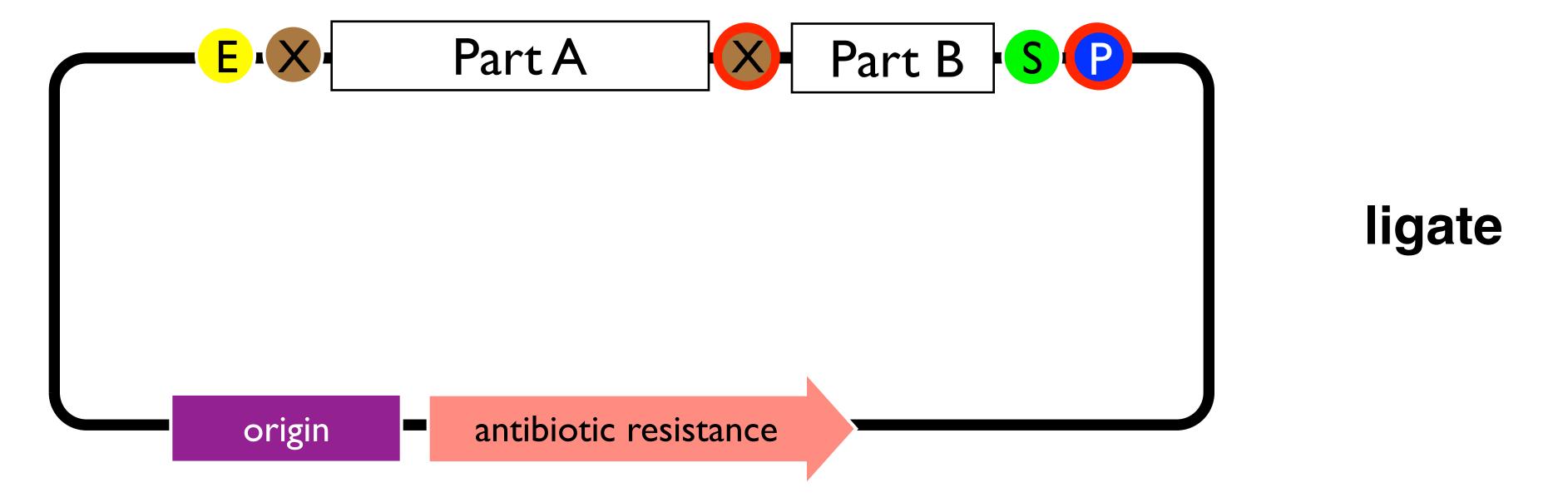
origin

antibiotic resistance

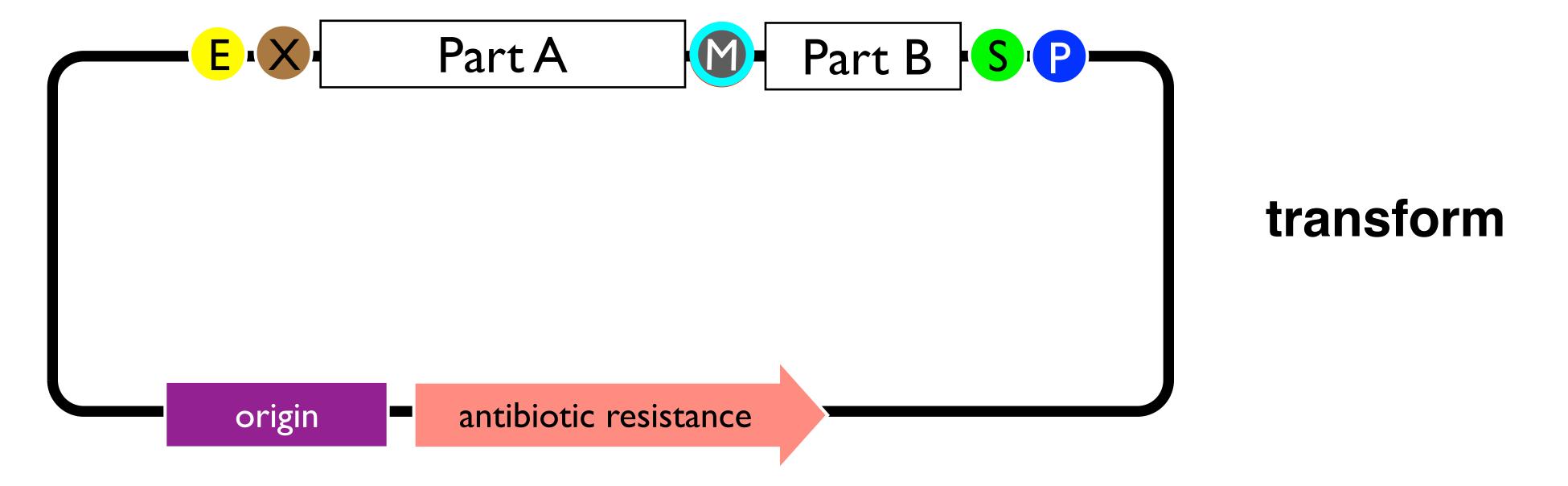


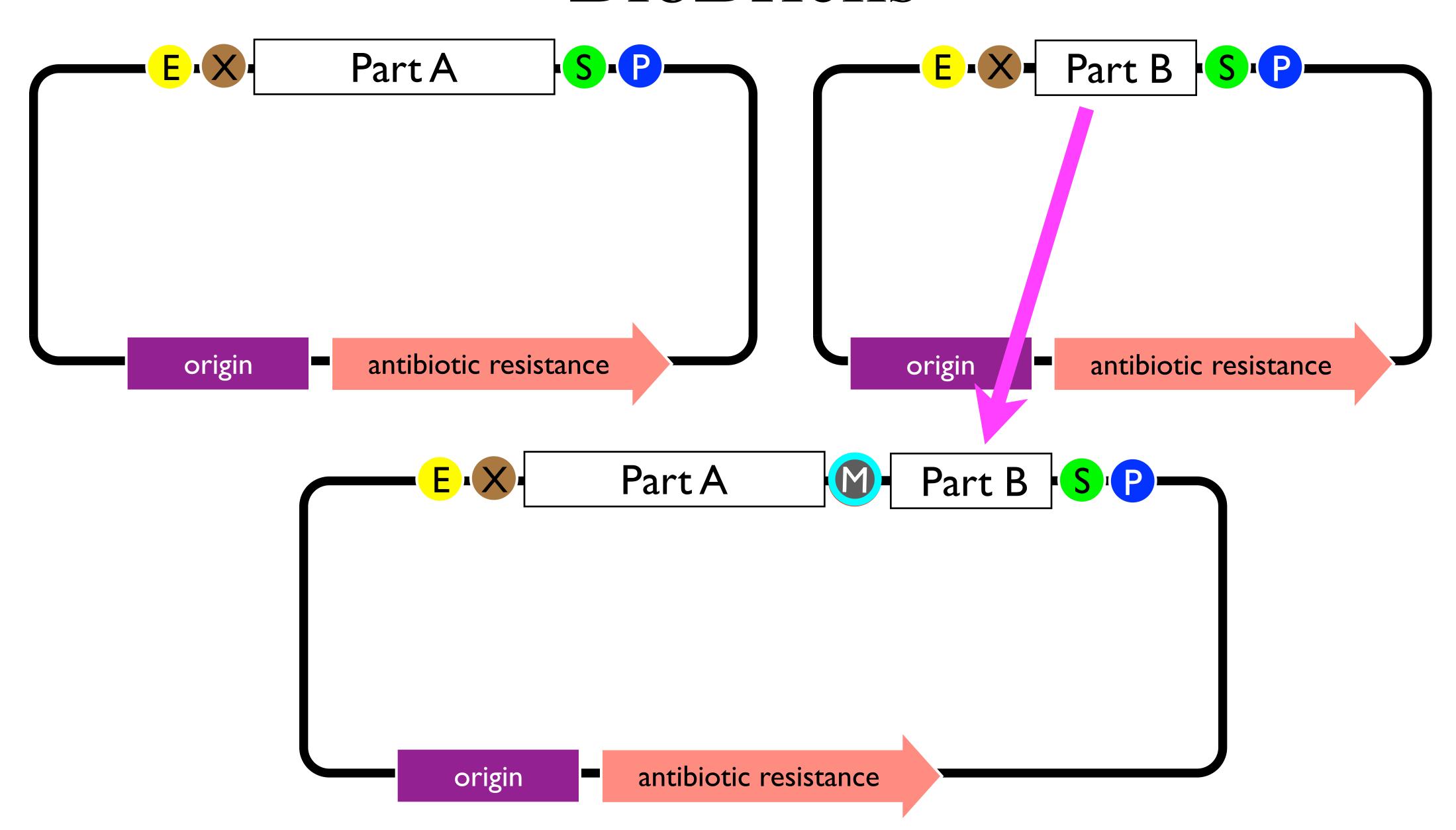


gel purify



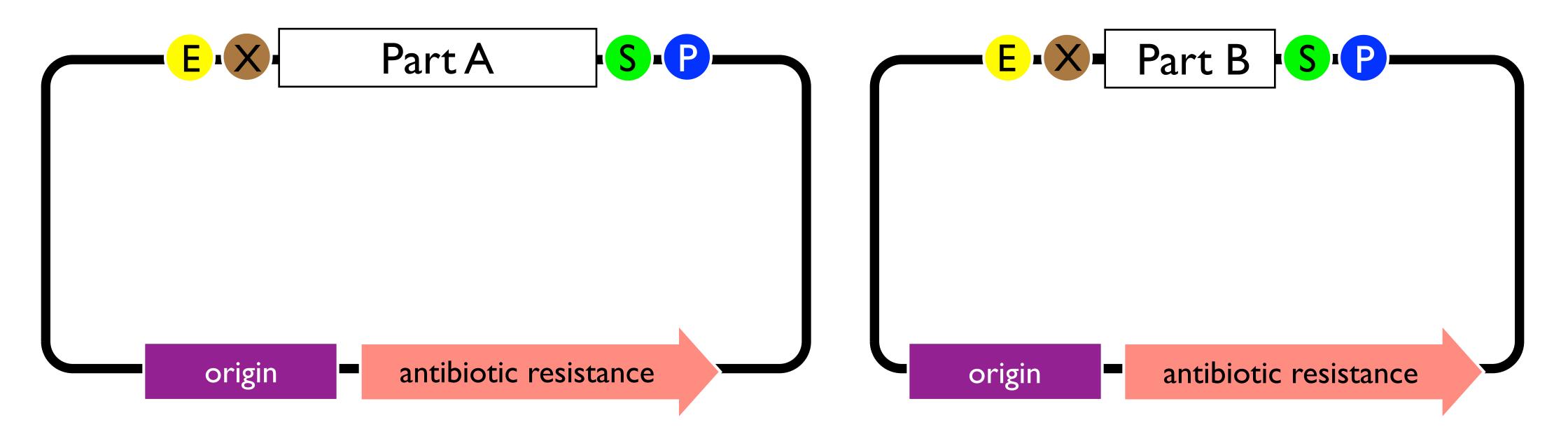
#### mixed site = scar



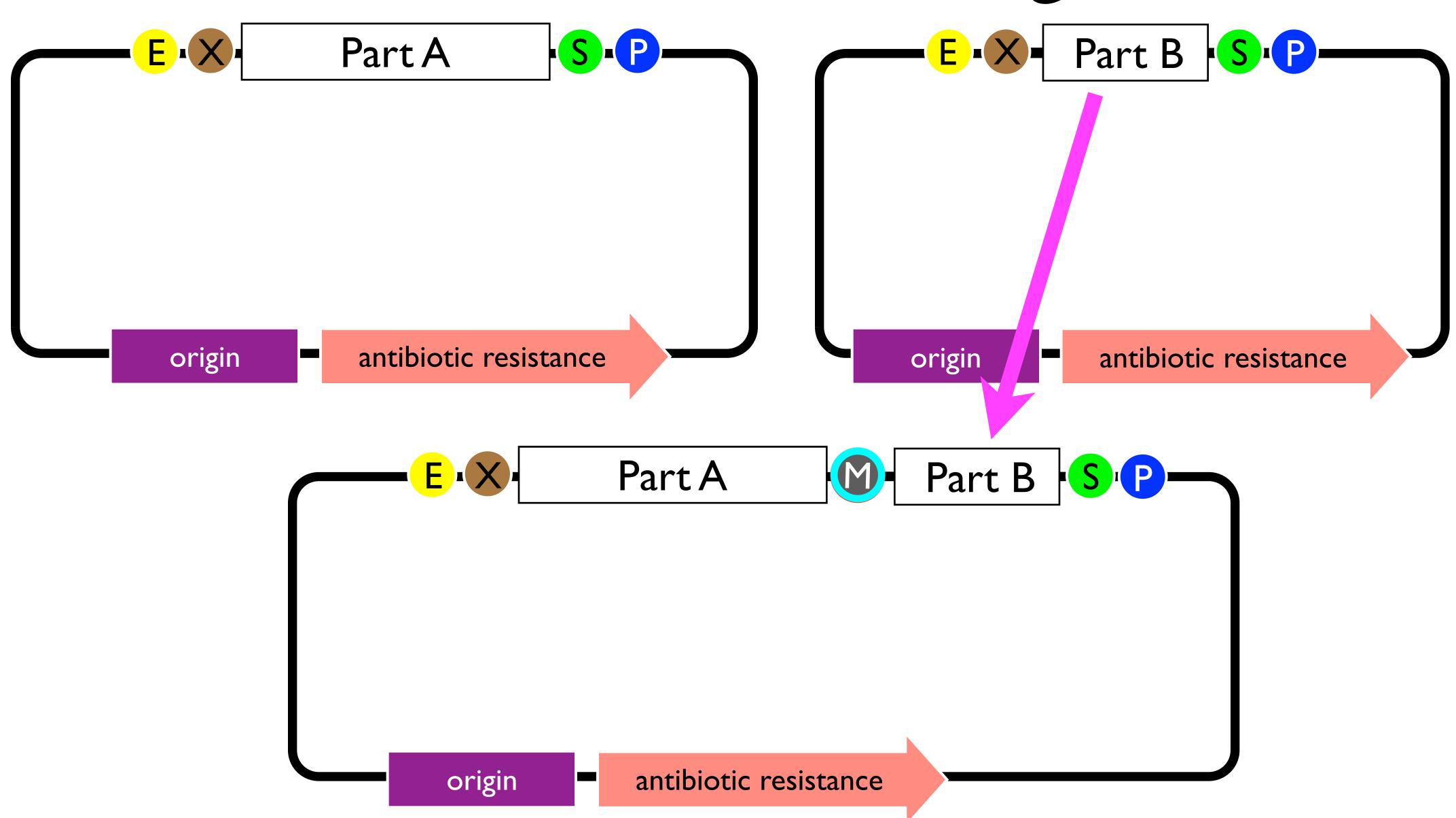


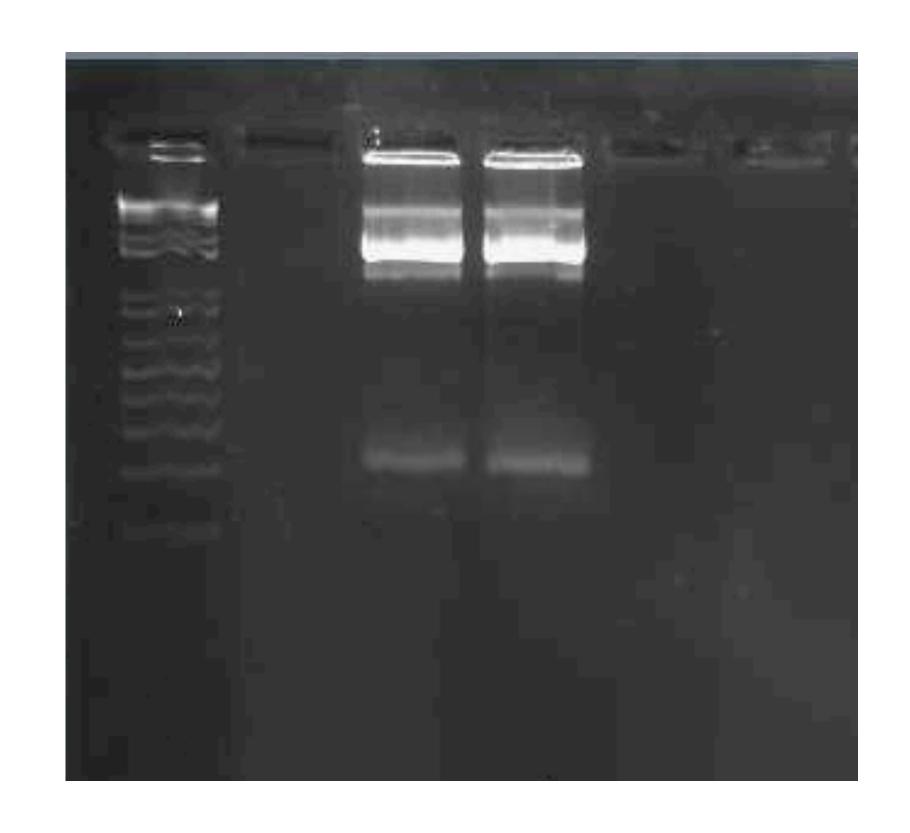
### Challenge:

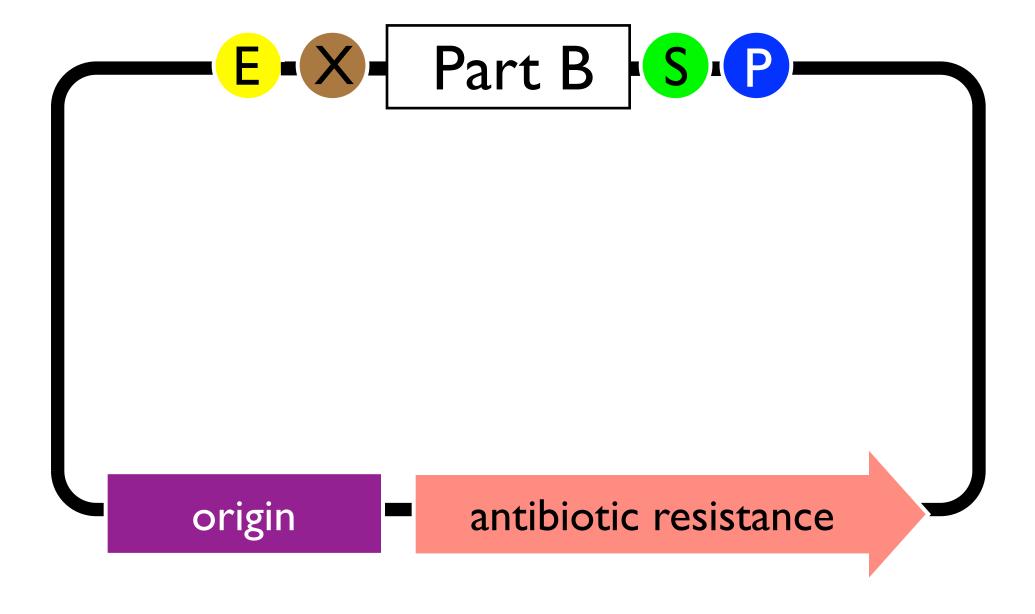
#### put A upstream of B

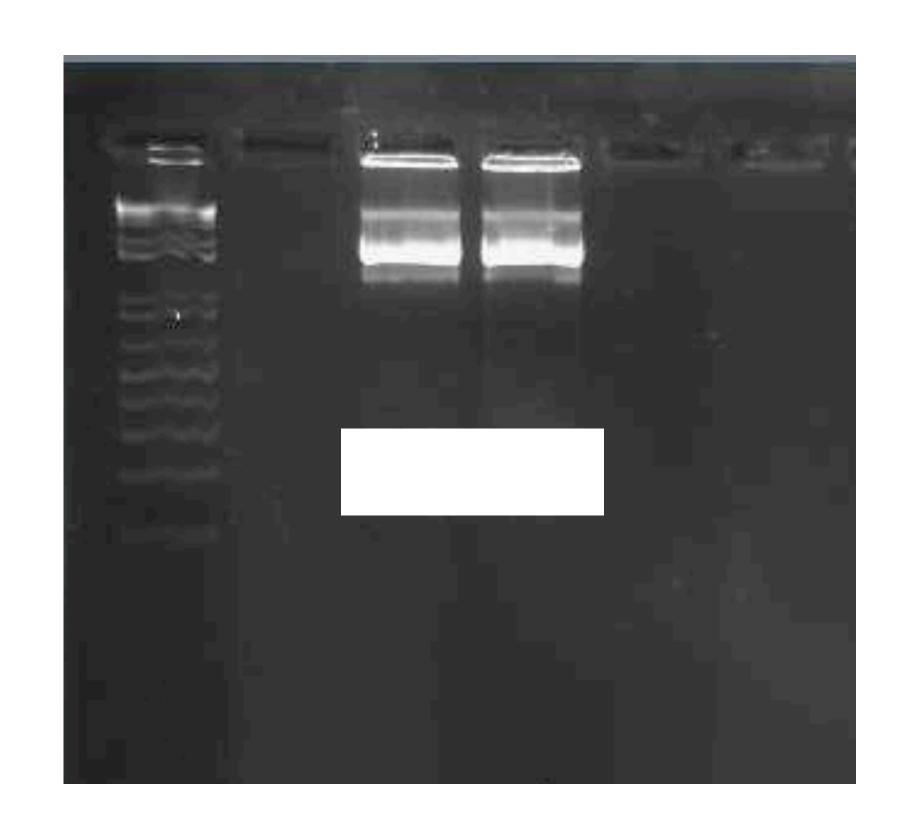


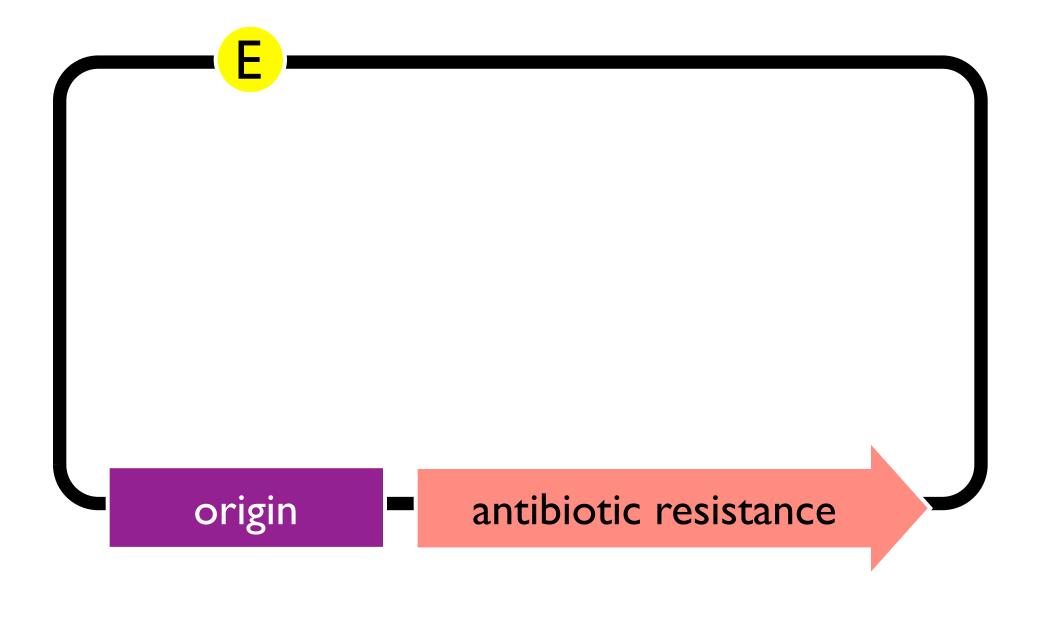
## Details of Cloning

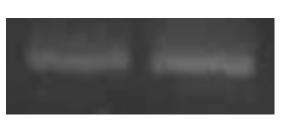








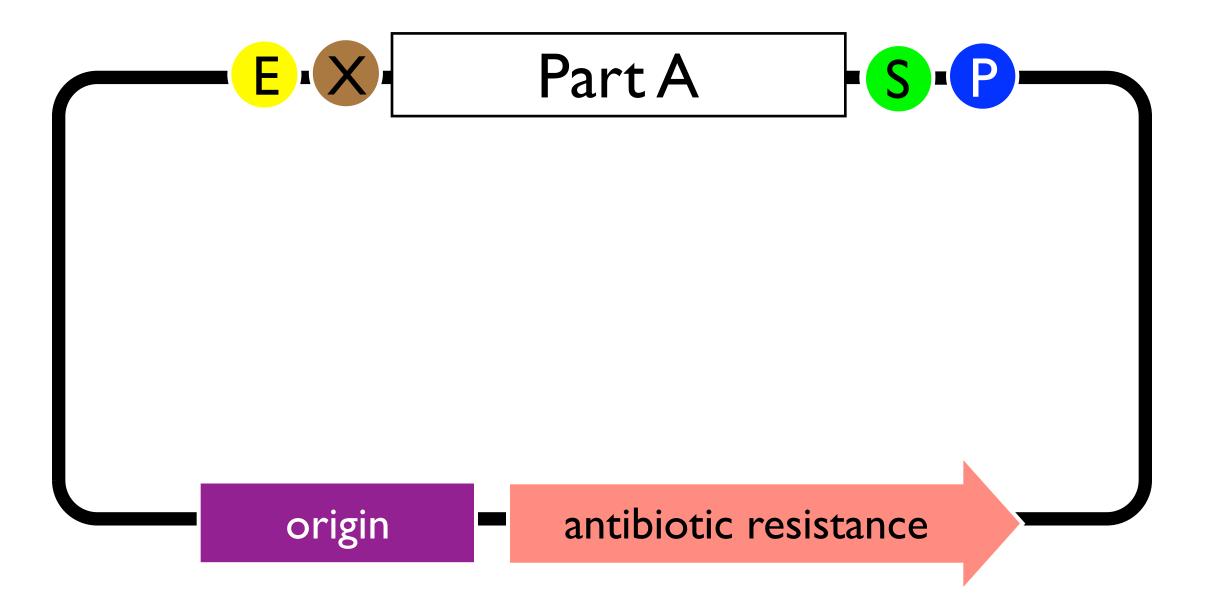


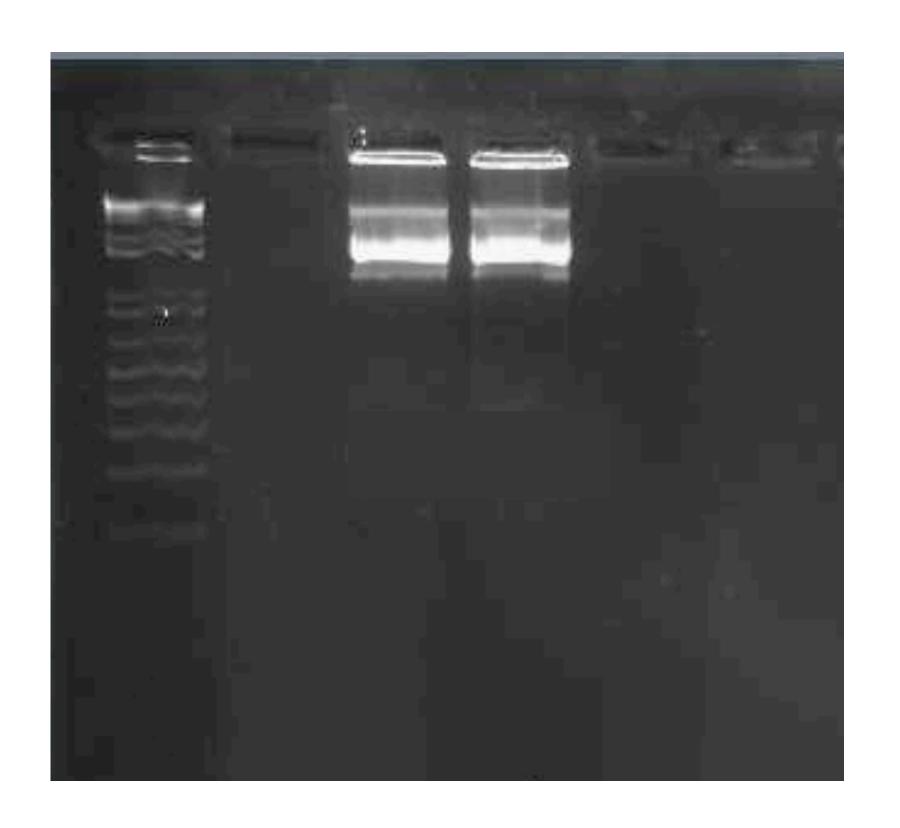






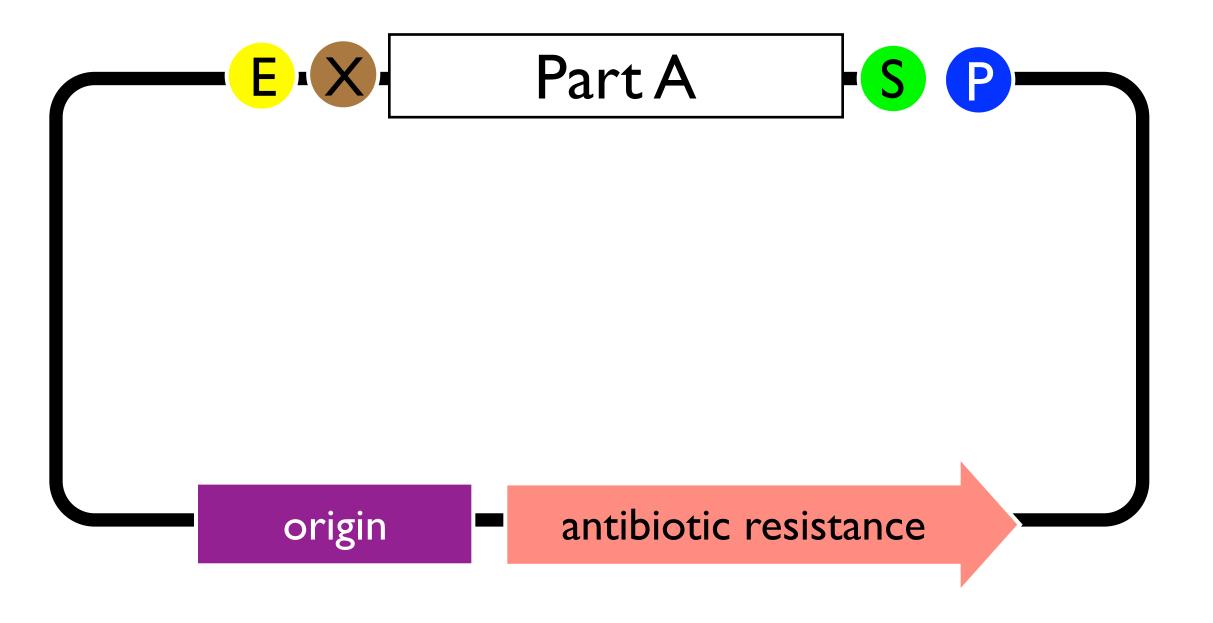






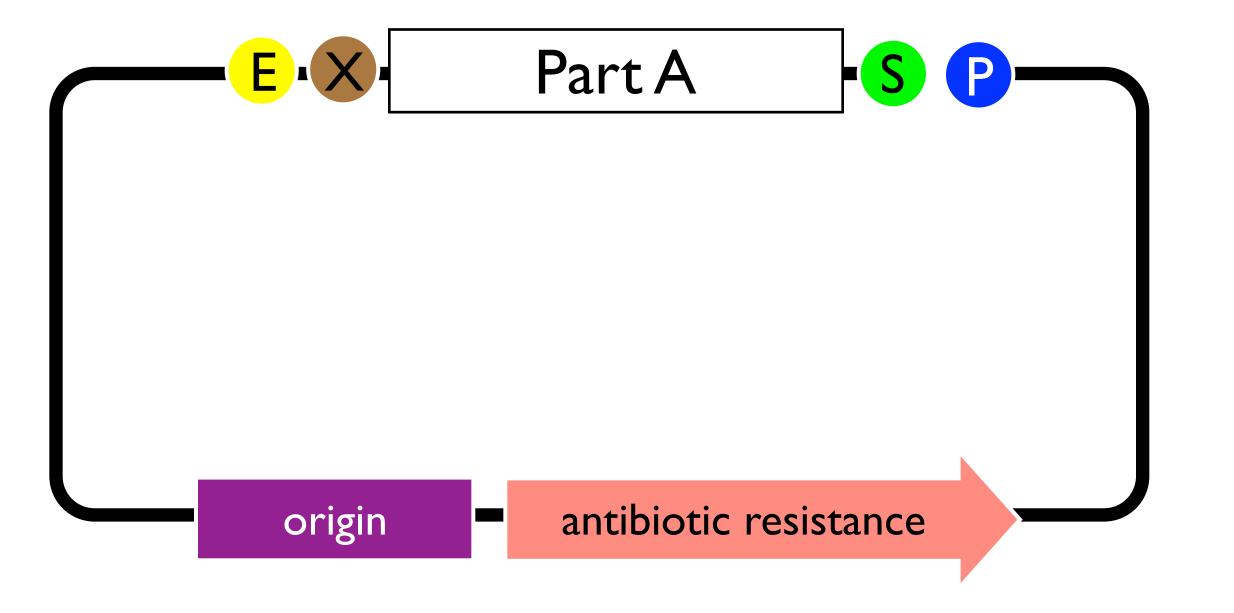


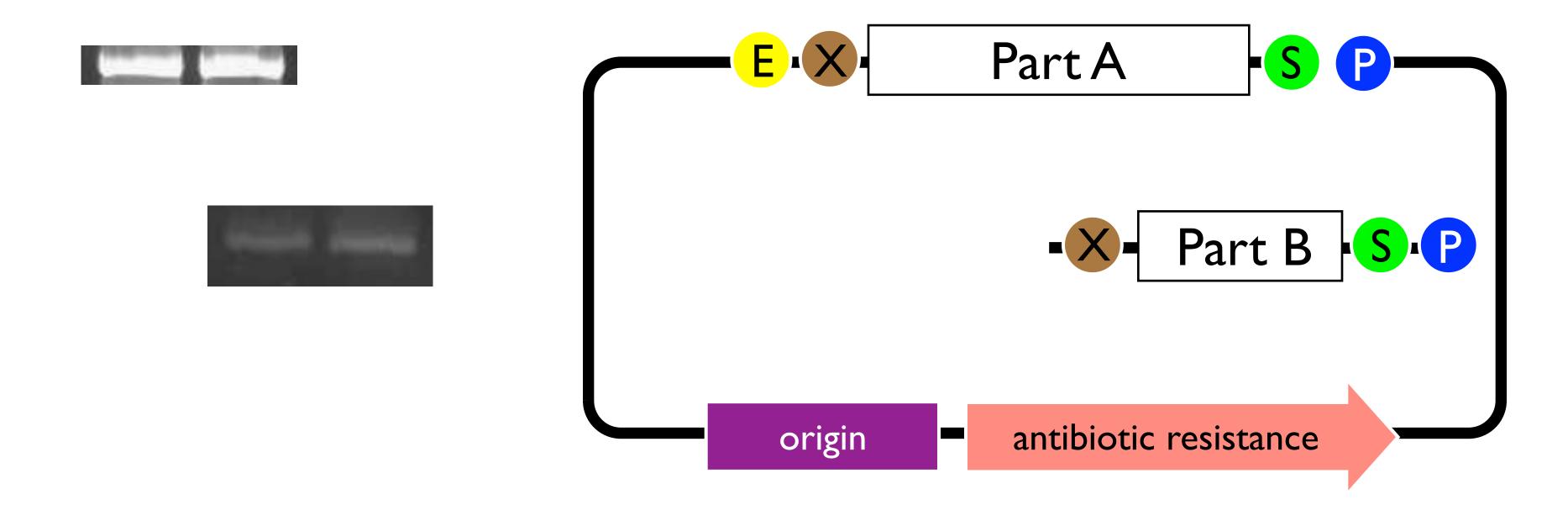




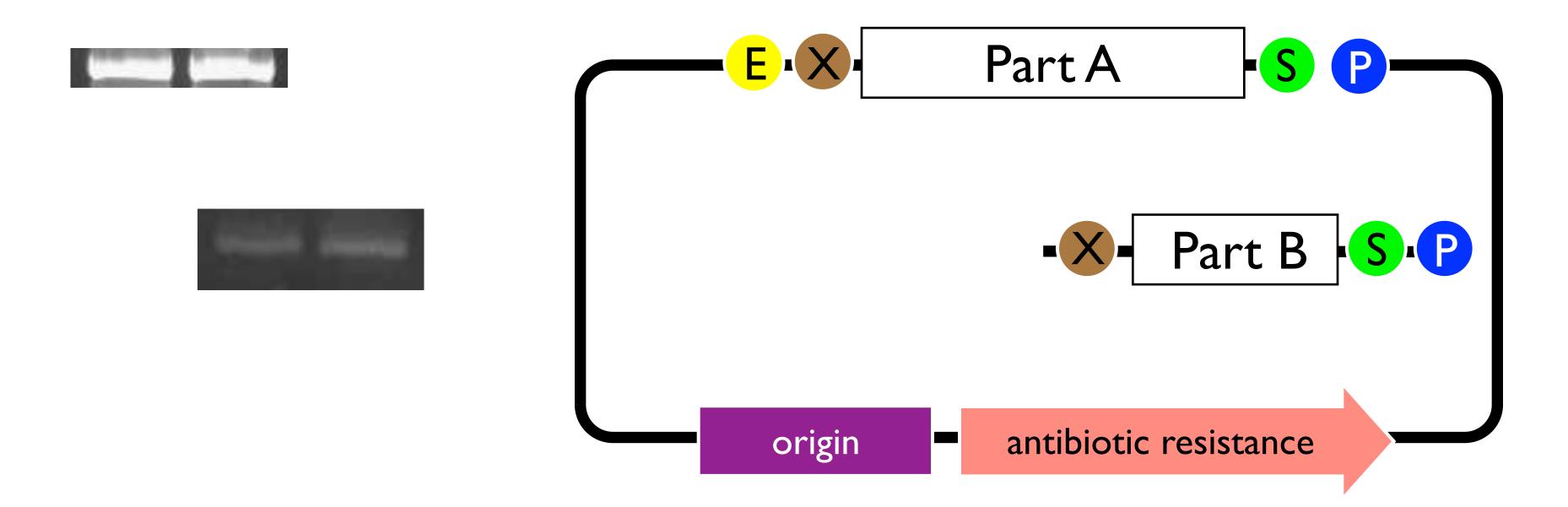




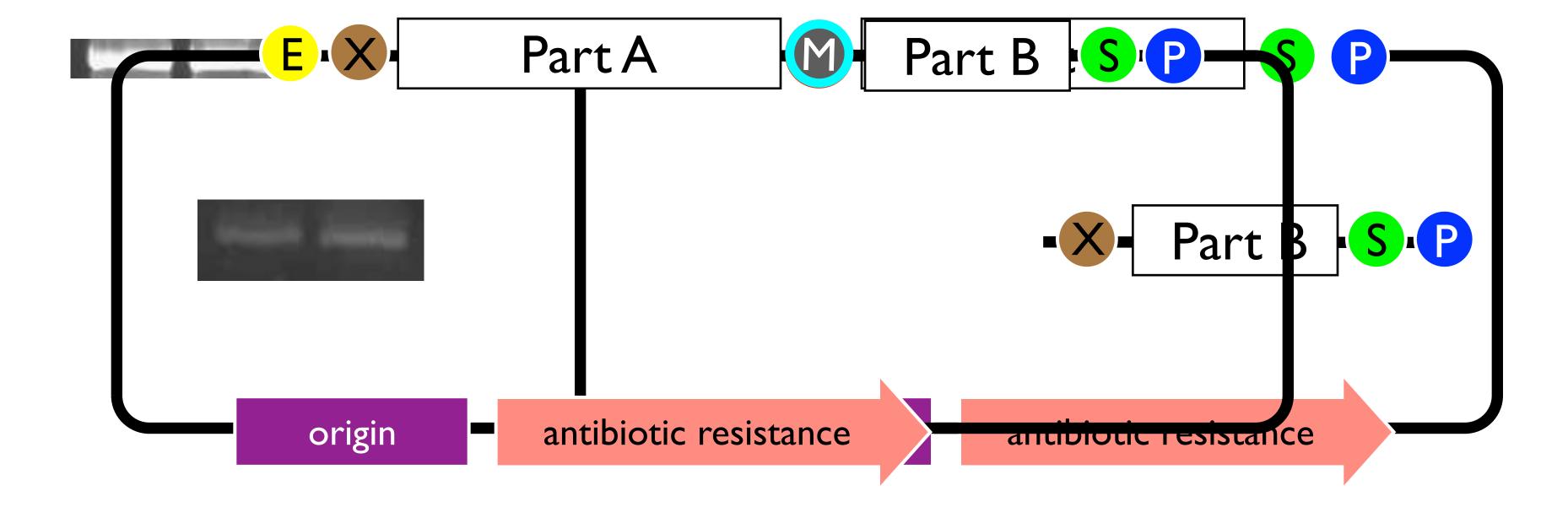




## Ligation

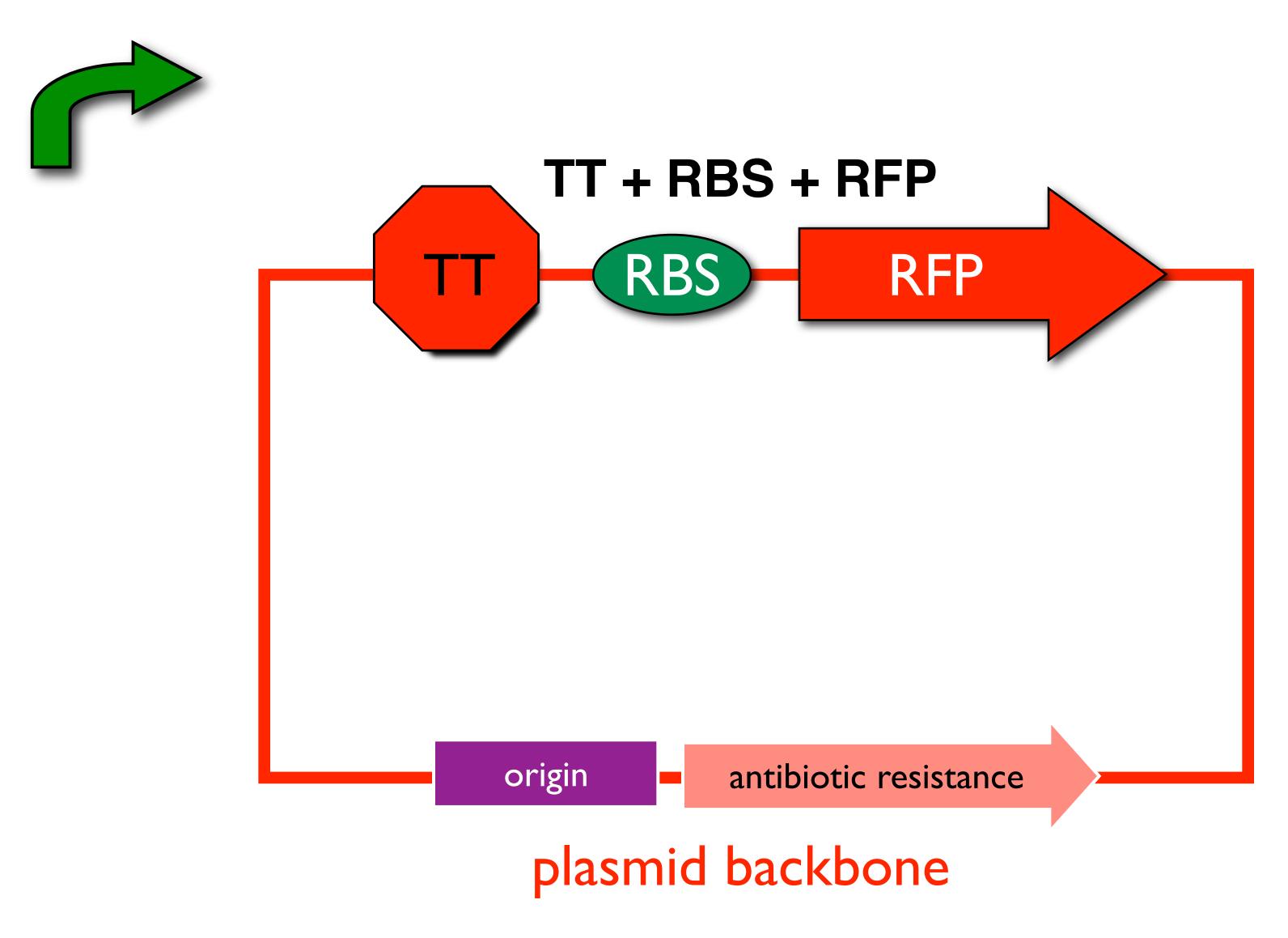


## Ligation

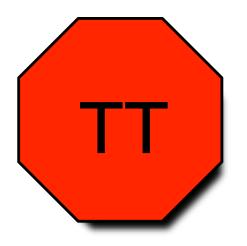


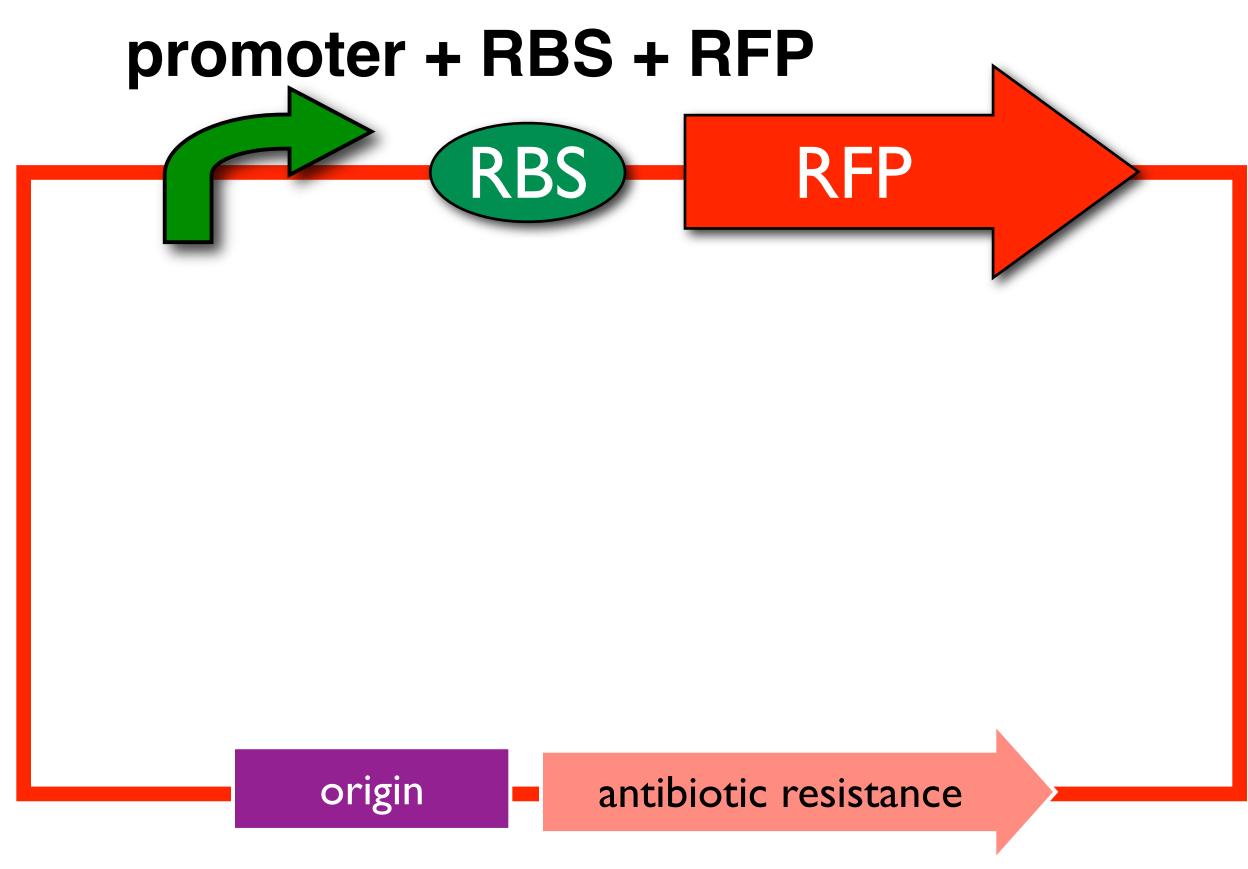
# Can intro bio students do real synthetic biology research in 3 hour labs?

## Golden Gate Assembly Method



## Golden Gate Assembly Method





plasmid backbone

GAATTC
CTTAAG

palindrome

GAATTC
CTTAAG

palindrome

GAATTC
CTTAAG

G AATTC CTTAA G

GAGACC
CTCTGG

not a palindrome

1234nGAGACC nCTCTGG

# BsaI

GGTCTCn
CCAGAGn1234

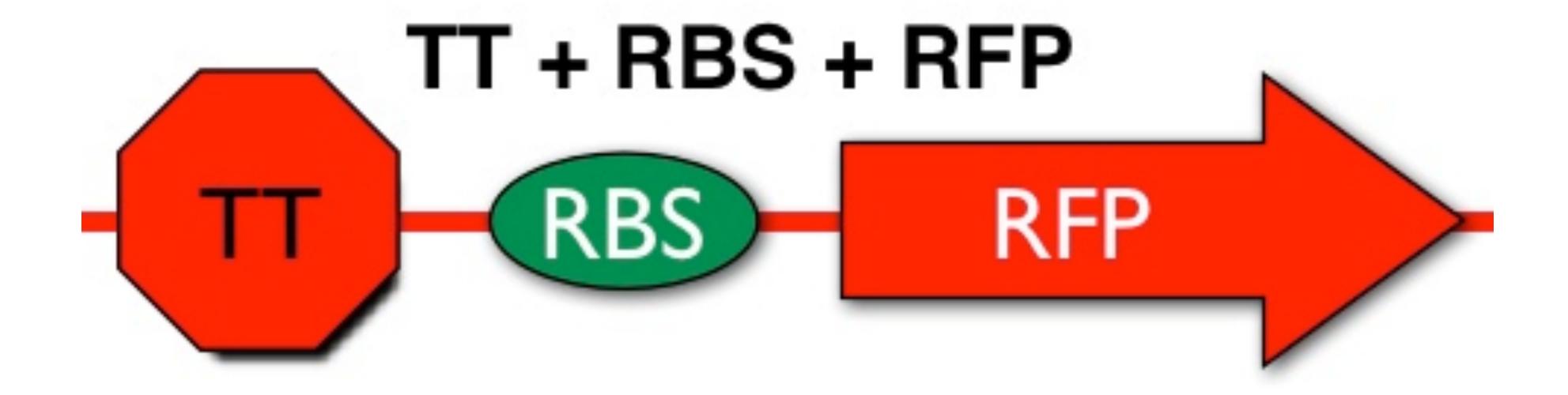
type IIs

# BsaI

1234nGAGACC

left ---nCTCTGG

GGTCTCn--- cuts right

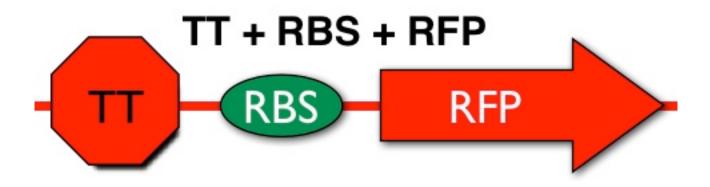


#### Bsa I

I CGACtGAGACC (TT) GGTCTCaGCGG

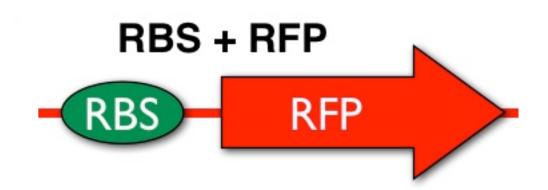
GCTGaCTCTGG (TT) CCAGAGtCGCCI

Bsa I



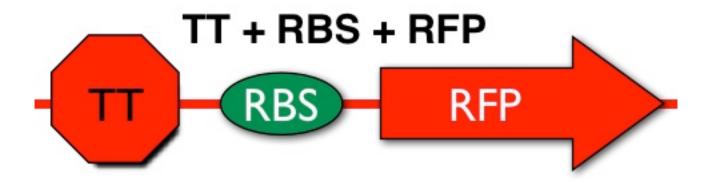
#### CGACtGAGACC (TT) GGTCTCa aCTCTGG (TT) CCAGAGtCGCC





CGACtGAGACC (TT)GGTCTCaGCGG

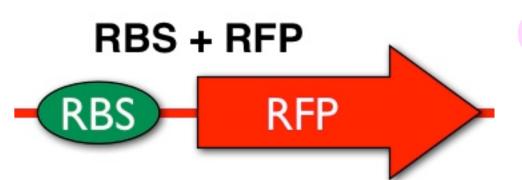
GCTGaCTCTGG (TT)CCAGAGtCGCC



#### CGACtGAGACC (TT) GGTCTCa aCTCTGG (TT) CCAGAGtCGCC

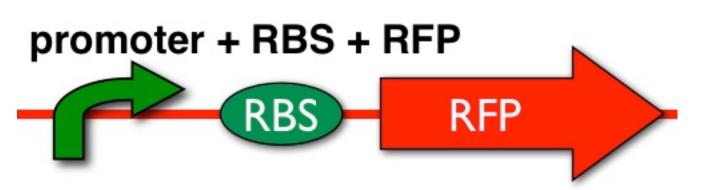


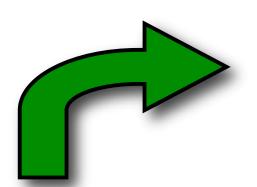


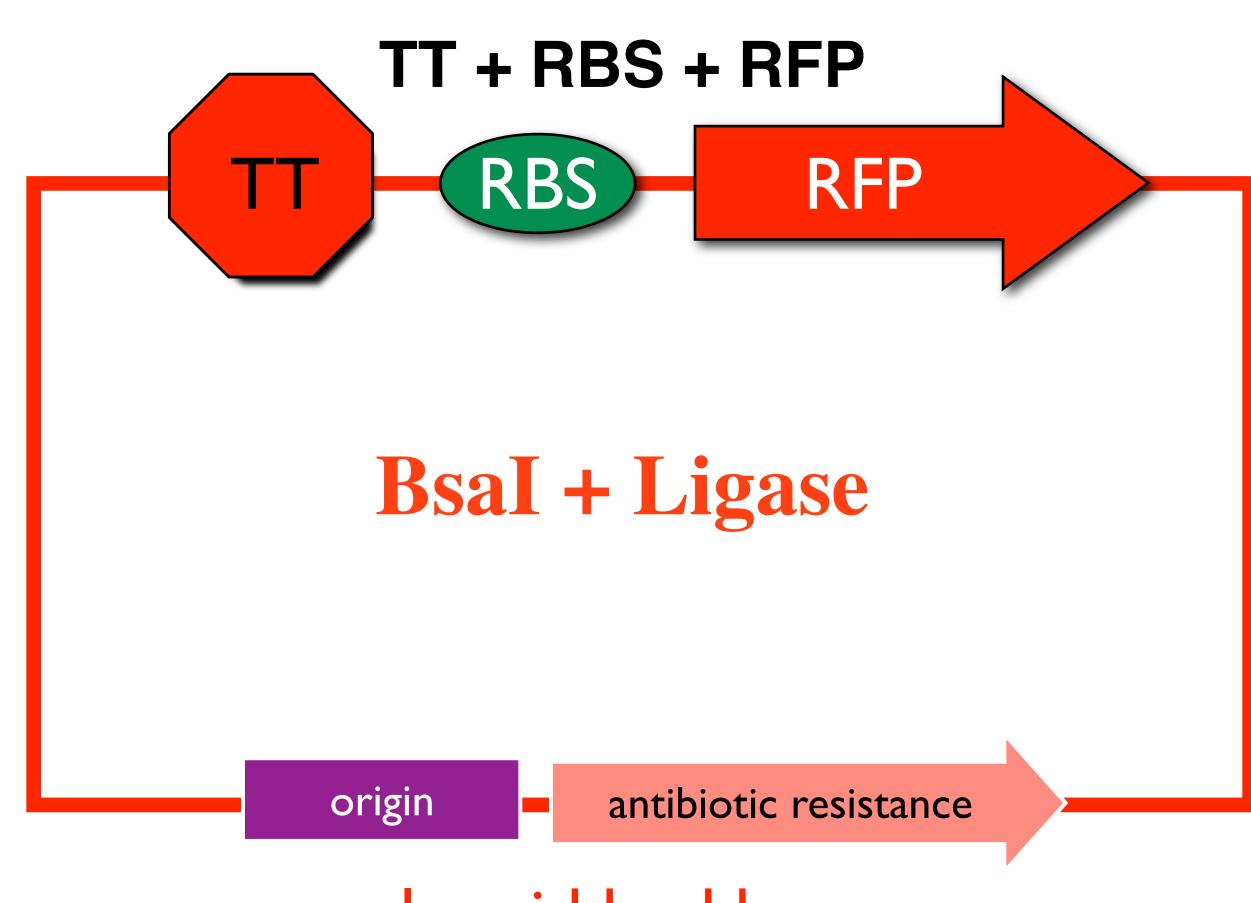


CGAC (promoter)
(promoter) CGCC

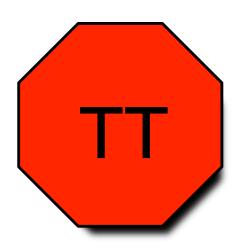


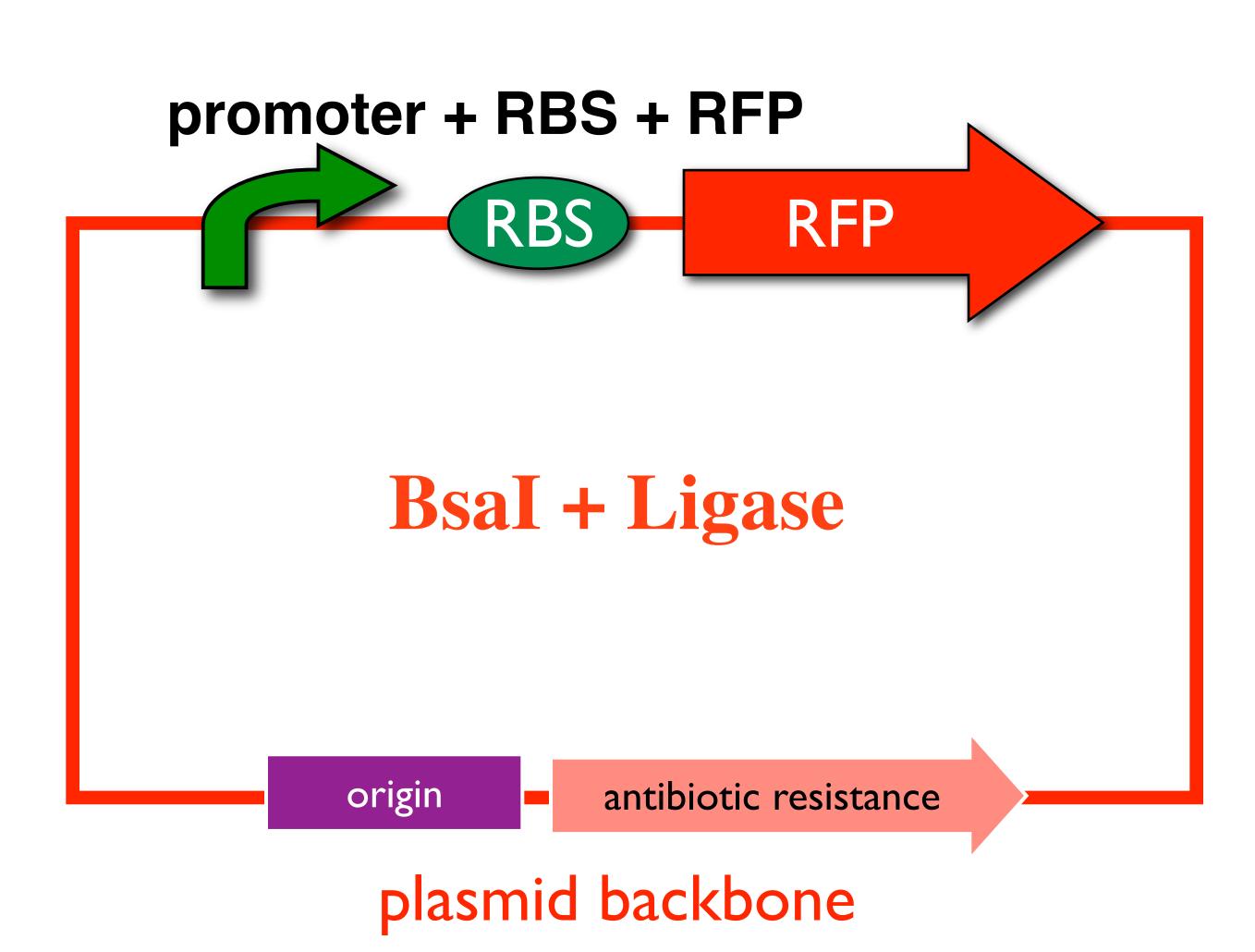


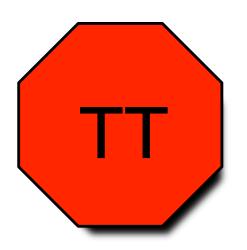


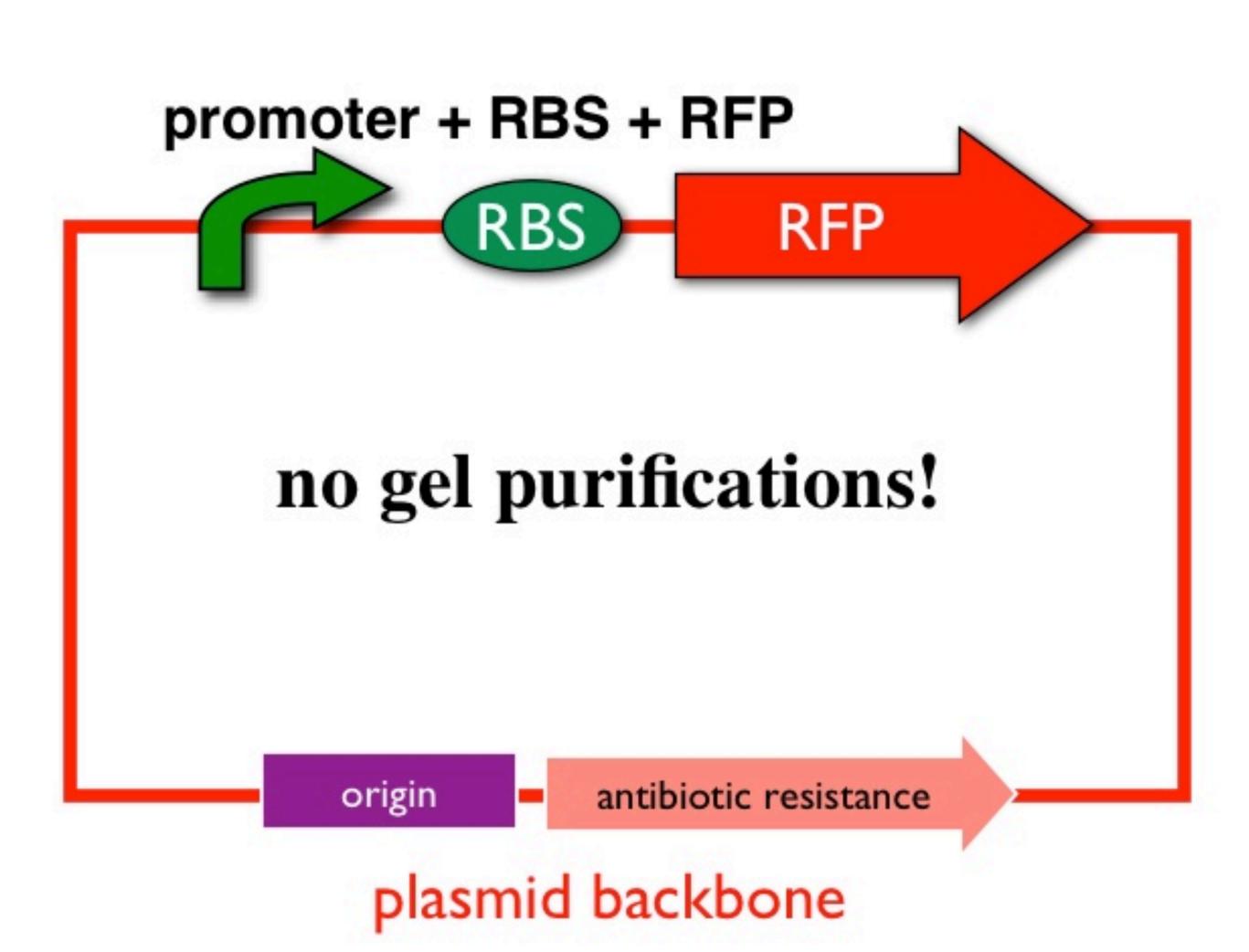


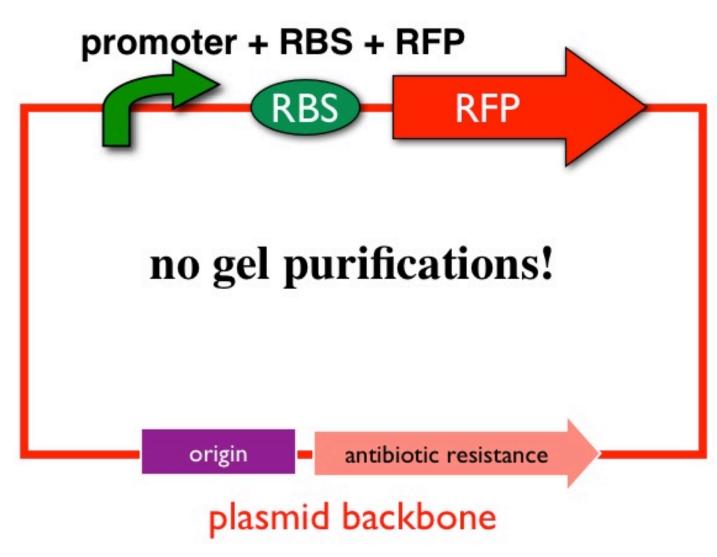
plasmid backbone







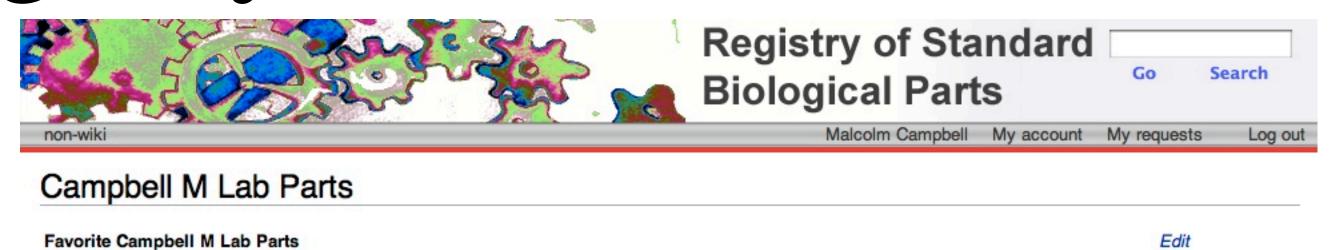






## Registry of Functional Promoters

Designer



Description

#### Campbell M Lab Parts Sandbox

Name

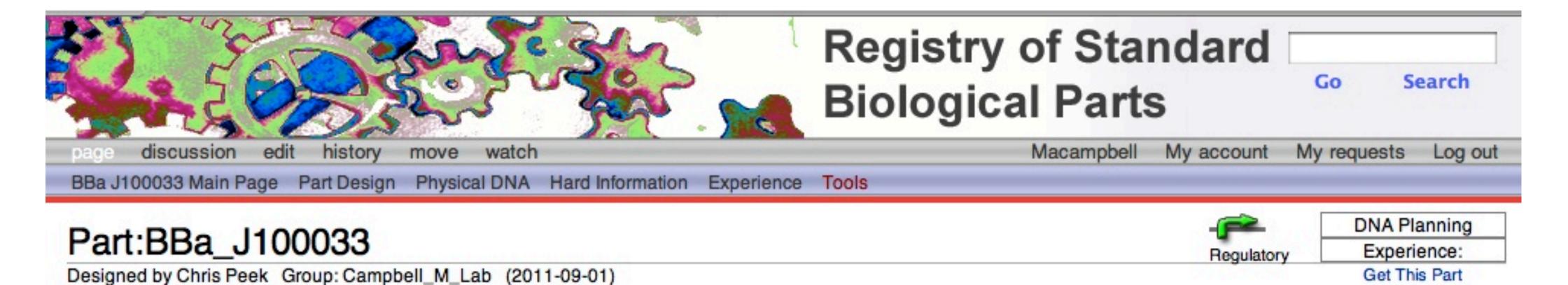
Type

Edit

Length

| -?-           |   | Name        | Type           | Description  | Designer          | Length |
|---------------|---|-------------|----------------|--|-------------------|--------|
| $\rightarrow$ |   | BBa_J100000 | Coding         | Cre with 8bp restriction sites and 1-Clause 2-SAT Problem Inserted | Eric Sawyer       | 1069   |
| - 33          |   | BBa_J100001 | Composite      | pTet+RBS+Cre2SAT1Clause+pLpp+tRNA CCACU                            | Eric Sawyer       | 1357   |
| 5.01          |   | BBa_J100002 | Composite      | pTet+RBS+Cre2SAT1Clause+pLpp+tRNA CGGUC                            | Eric Sawyer       | 1357   |
| - 1           |   | BBa_J100003 | Generator      | pTet+RBS+Cre2SAT1Clause  | Eric Sawyer       | 1149   |
| - 33          |   | BBa_J100004 | Reporter       | pTet+LoxP+RBS+RFP+LoxP   | Eric Sawyer       | 870    |
|               |   | BBa_J100005 | Other          | Palindromic Stop Sequence  | Eric Sawyer       | 221    |
| 100           |   | BBa_J100006 | Intermediate   | LoxP+Stop Sequence+LoxP  | Eric Sawyer       | 305    |
| - 39          |   | BBa_J100007 | Intermediate   | pLac+RBS+LoxP+Stop Sequence+LoxP                                   | Eric Sawyer       | 533    |
|               |   | BBa_J100008 | Composite      | pLpp-tRNA CCACU-pLpp-tRNA CUAGU                                    | Eric Sawyer       | 408    |
| - 33          |   | BBa_J100009 | Composite      | pLpp-tRNA CCACU-pLpp-tRNA CGGUC                                    | Eric Sawyer       | 408    |
| 5.61          |   | BBa_J100010 | Composite      | pLpp-tRNA CUAGU-pLpp-tRNA CGGUC                                    | Eric Sawyer       | 408    |
|               |   | BBa_J100011 | Composite      | pLpp-tRNA CCACU-pLpp-tRNA CUAGU-pLpp-tRNA CGGUC                    | Eric Sawyer       | 616    |
| - 33          |   | BBa_J100012 | Intermediate   | RBS-RFP-RBS  | Eric Sawyer       | 747    |
|               |   | BBa_J100013 | Coding         | Luxl with 1 Clause 2-SAT Problem                                   | Eric Sawyer       | 638    |
| 1.00          |   | BBa_J100014 | Coding         | Luxl with 2 Clause 2-SAT Problem                                   | Eric Sawyer       | 652    |
| - 39          |   | BBa_J100015 | Composite      | 1 Clause 2-SAT Problem with Frameshifted Luxl and a GFP Reporter   | Eric Sawyer       | 2757   |
| 200           |   | BBa_J100016 | Composite      | 2 Clause 2-SAT Problem with Frameshifted Luxl and a GFP Reporter   | Eric Sawyer       | 2771   |
| - 33          |   | BBa_J100017 | Composite      | TT+pLux+RBS+LuxI(2-SAT 2 clause)+RBS+GFP+pLac+RBS+LuxR+tRNAs       | Eric Sawyer       | 3395   |
| 5.4           |   | BBa_J100018 | Protein_Domain | First Half of AspC gene  | Catherine Doyle   | 448    |
|               |   | BBa_J100019 | Protein_Domain | First half of livE gene  | Julia Fearrington | 457    |
| - 33          |   | BBa_J100020 | Protein_Domain | Second Half of AspC  | Catherine Doyle   | 869    |
|               |   | BBa_J100021 | Protein_Domain | First Half of PyrE   | Catherine Doyle   | 488    |
| 100           |   | BBa_J100022 | Protein_Domain | Second Half of PyrE  | Catherine Doyle   | 280    |
| - 39          |   | BBa_J100025 | Protein_Domain | First half of CAT gene   | James Harden      | 434    |
|               |   | BBa_J100026 | Protein_Domain | second half ilvE gene  | Julia Fearrington | 574    |
| - 33          |   | BBa_J100027 | Protein_Domain | second half of TyrB  | James Harden      | 288    |
| 5.61          |   | BBa_J100028 | Other          | placeholder insert for Bsal Golden Gate Assembly of promoter       | Malcolm Campbell  | 877    |
|               |   | BBa_J100029 | Regulatory     | The promoter of rpoDPhs  | Maggie Baay       | 76     |
| - 33          |   | BBa_J100030 | Regulatory     | phoA is an inducible promoter induced by phosphate starvation.     | Scott Hall        | 76     |
|               |   | BBa_J100031 | Regulatory     | Constitutive promoter C on Gene 1 of T7, transcribes RNA Pol.      | Caroline Vrana    | 100    |
|               |   | BBa_J100032 | Regulatory     | proUP3 promoter  | Molly Marshall    | 90     |
|               |   | BBa_J100033 | Regulatory     | dnakP1 promoter: Heat shock inducible                              | Chris Peek        | 101    |
|               |   | BBa_J100034 | Regulatory     | groE promoter  | Margaret Stebbins | 44     |
| - 33          |   | BBa_J100036 | Regulatory     | Promoter induced by DNA damage                                     | Erich Baker       | 52     |
| 5.00          |   | BBa_J100039 | Regulatory     | GalP1 Promoter-Induced By Galactose                                | Anaiah Toby       | 75     |
| - 1           |   | BBa_J100040 | Coding         | Luxl with 3 clause 2-SAT problem                                   | Eric Sawyer       | 684    |
| - 33          |   | BBa_J100041 | Composite      | LuxI/GFP with 3 clause 2-SAT problem                               | Eric Sawyer       | 2803   |
|               |   | BBa_J100042 | Coding         | LuxI with 3 clause 3-SAT problem                                   | Eric Sawyer       | 702    |
| 7.77          |   | BBa_J100043 | Composite      | LuxI/GFP with 3 clause 3-SAT problem                               | Eric Sawyer       | 2821   |
| 3             | _ | BBa_J100044 | Coding         | Luxl with 4 clause 2-SAT problem                                   | Eric Sawyer       | 704    |
|               |   | BBa_J100045 | Composite      | LuxI/GFP with 4 clause 2-SAT problem                               | Eric Sawyer       | 2823   |
| 33            |   | BBa_J100046 | RNA            | lpp+tRNA CCAUC (10 bp anticodon loop)                              | Eric Sawyer       | 201    |
| 1.0           | _ | BBa_J100047 | Protein_Domain | TyrB2  | Julia Fearrington |        |
|               |   | BBa_J100048 | Protein_Domain | TyrB1  | Julia Fearrington | 930    |
| 31            | _ | BBa_K091231 | Composite      | LuxR producer and XOR gate   | Malcolm Campbell  | 2772   |
|               |   | BBa_K091232 | Composite      | LuxR producer and RFP(rev) + RBS(rev) + pLux (for)                 | Malcolm Campbell  | 1916   |

## Student Sample



#### dnakP1 promoter: Heat shock inducible

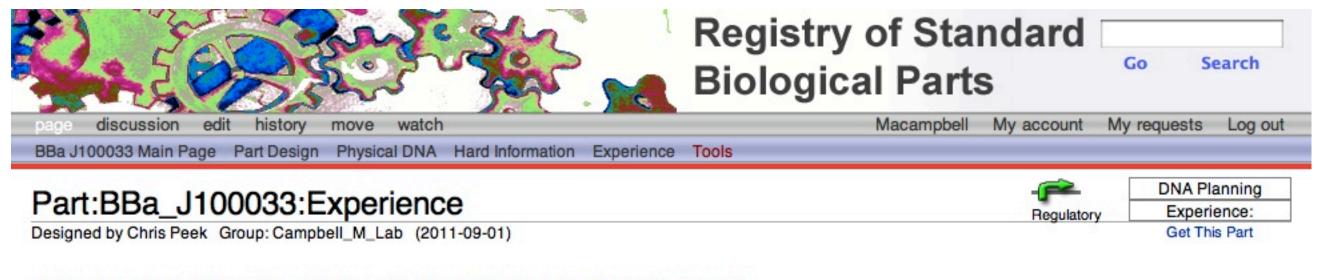
dnaKP1 is naturally off, but is induced when E. coli is heat shocked, resulting in transcription downstream from this promoter.

#### Sequence and Features

| Format: | Format: Subparts I Ruler I SS I DS   |    | Search: Length: 101 |    | 1 bp Context: Part only |    | nly G | Get selected sequence |    |
|---------|--------------------------------------|----|---------------------|----|-------------------------|----|-------|-----------------------|----|
| 1       | 11                                   | 21 | 31                  | 41 | 51                      | 61 | 71    | 81                    | 91 |
|         | tctgc gcaaaagcac<br>agacg cgttttcgtg |    |                     |    |                         |    |       |                       |    |
| 01 a    |                                      |    |                     |    |                         |    |       |                       |    |
| t       |                                      |    |                     |    |                         |    |       |                       |    |

Assembly Compatibility: 10 12 21 23 25

## Student Sample



This experience page is provided so that any user may enter their experience using this part.

Please enter how you used this part and how it worked out.

#### Applications of BBa\_J100033 [edit] Mean Fluorescence per Cell Density 10000 9000 Cell Desnity 8000 A: Experimental: 4000 1000 kPA1 (-) pTet (+) pLac - IPTG kPA1 (+) J10028 (-) pLac + IPTG Condition \* p < 0.01

cells containing dnaKP1 without heat shock (incubated at 37°C) B: Experimental: cells containing dnaKP1 with heat shock (incubated at 40°C) C: Negative control: part i100028 without pTet promoter (part i715039) with inducer (IPTG) F: pLac

# Our Current Challenge: Introductory Biology

#### Integrating Concepts in Biology

by

A. Malcolm Campbell, Laurie J. Heyer and Christopher J. Paradise

#### What's Wrong with Biology Education Now?

Globin gene family, 315, 316, 535

614, 651, 652, 664, 665

renal, 1099, 1100-1101, 1106

Gluconeogenesis, 154, 155, 175,

gluconeogenesis, 154, 155, 175,

Glucagon, 880, 887, 1087

forms of, 49, 50

overview of, 140, 142-144

Glycoproteins, 101

T cell receptors, 414

Glycosidic linkages, 50-51

634, 635, 636, 646

Glycosylation, 274

- Vocabulary is emphasized
- Experimental approaches are minimized

Germ line mutations, 275, 277

• Math is absent

Genetic drift, 494-495, 531

Genetic recombination, 223–224

- Memorization is rewarded
- Critical thinking is discouraged
- Information is irrelevant to students

#### If we currently cover all the important stuff....



...how can we add more content?

#### Too much content for the containers



#### Too much content for the containers

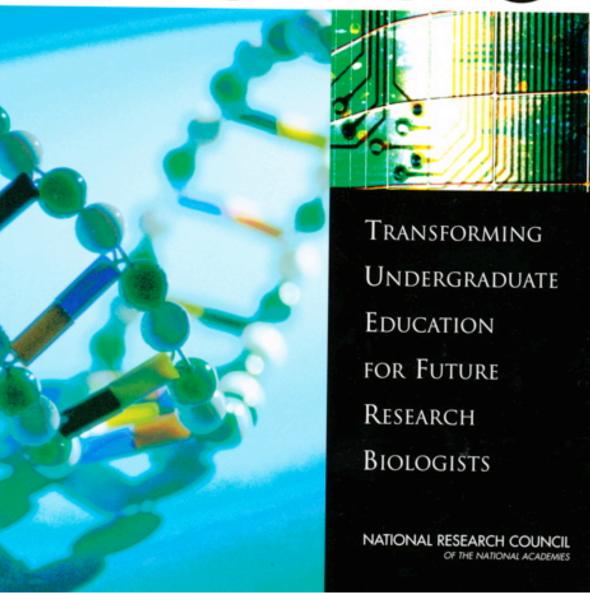


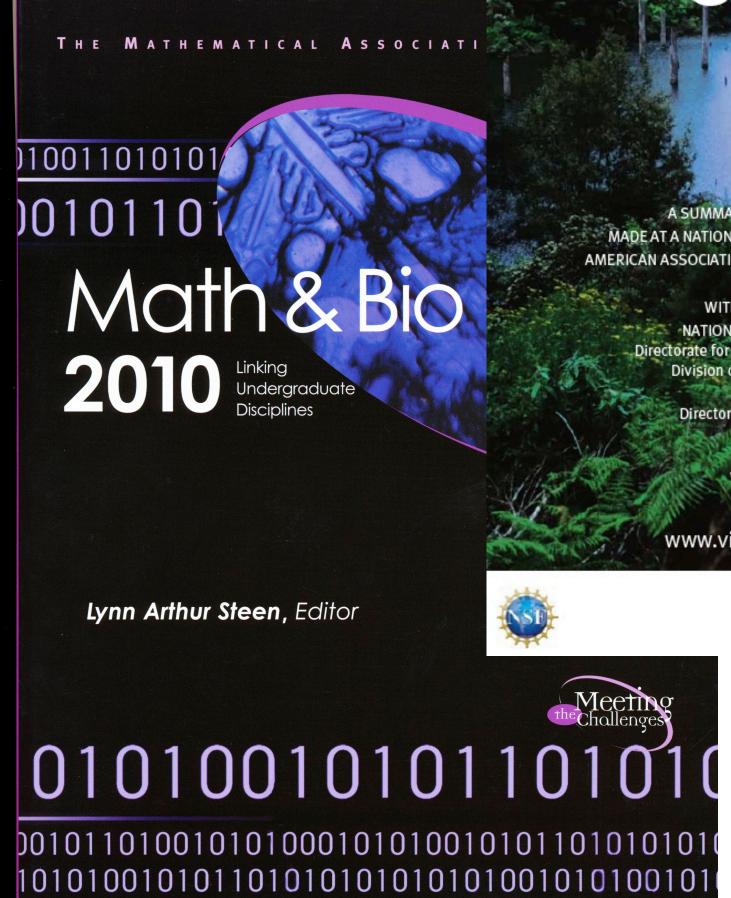


#### Start with the literature...



BI(







Expanded Edition

NESLANCII

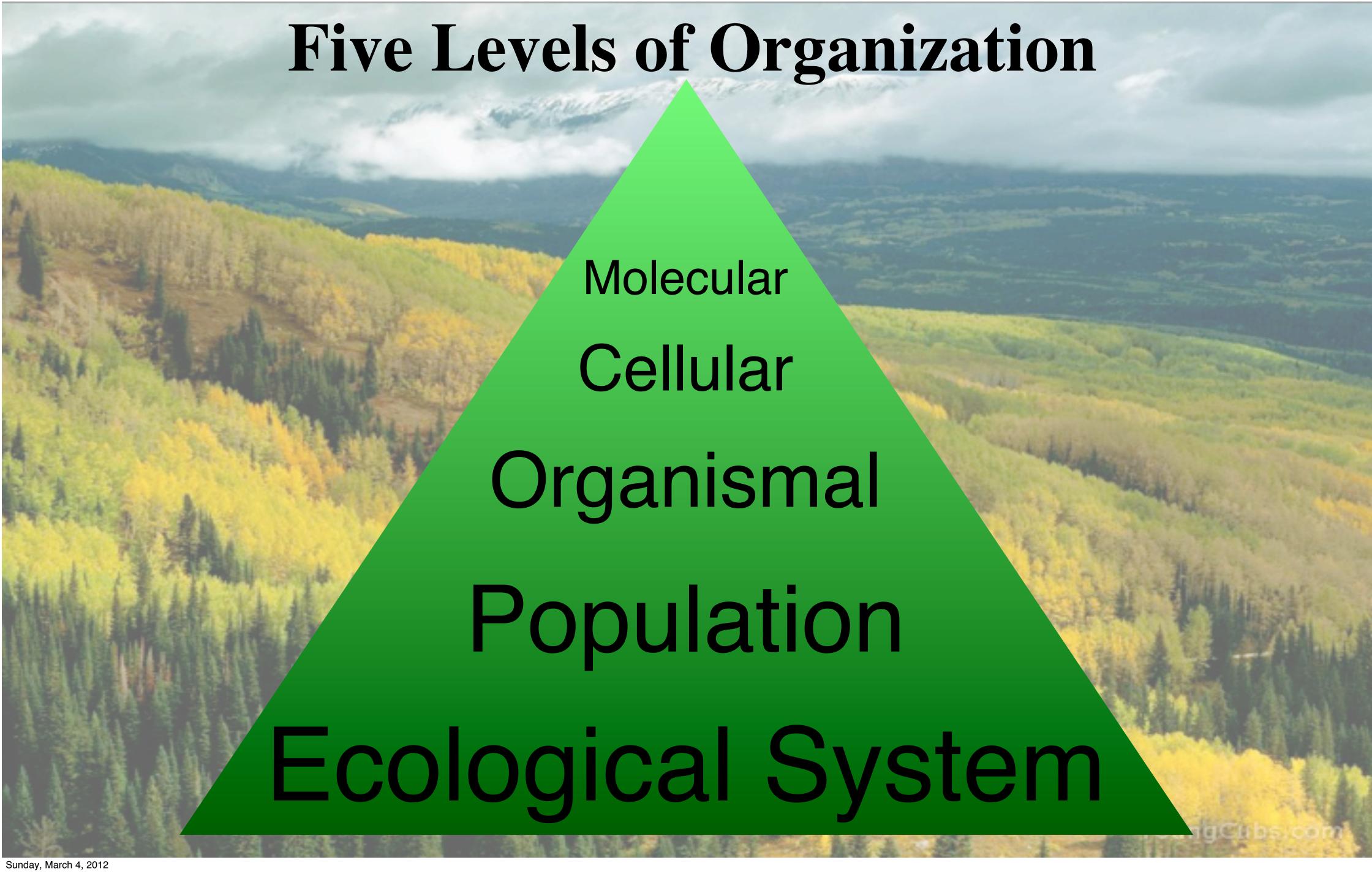
NATIONAL ACADEMY OF SCIENCES, NATIONAL ACADEMY OF ENGINEERING, AND INSTITUTE OF MEDICINE OF THE NATIONAL ACADEMIES

#### Present information and data...

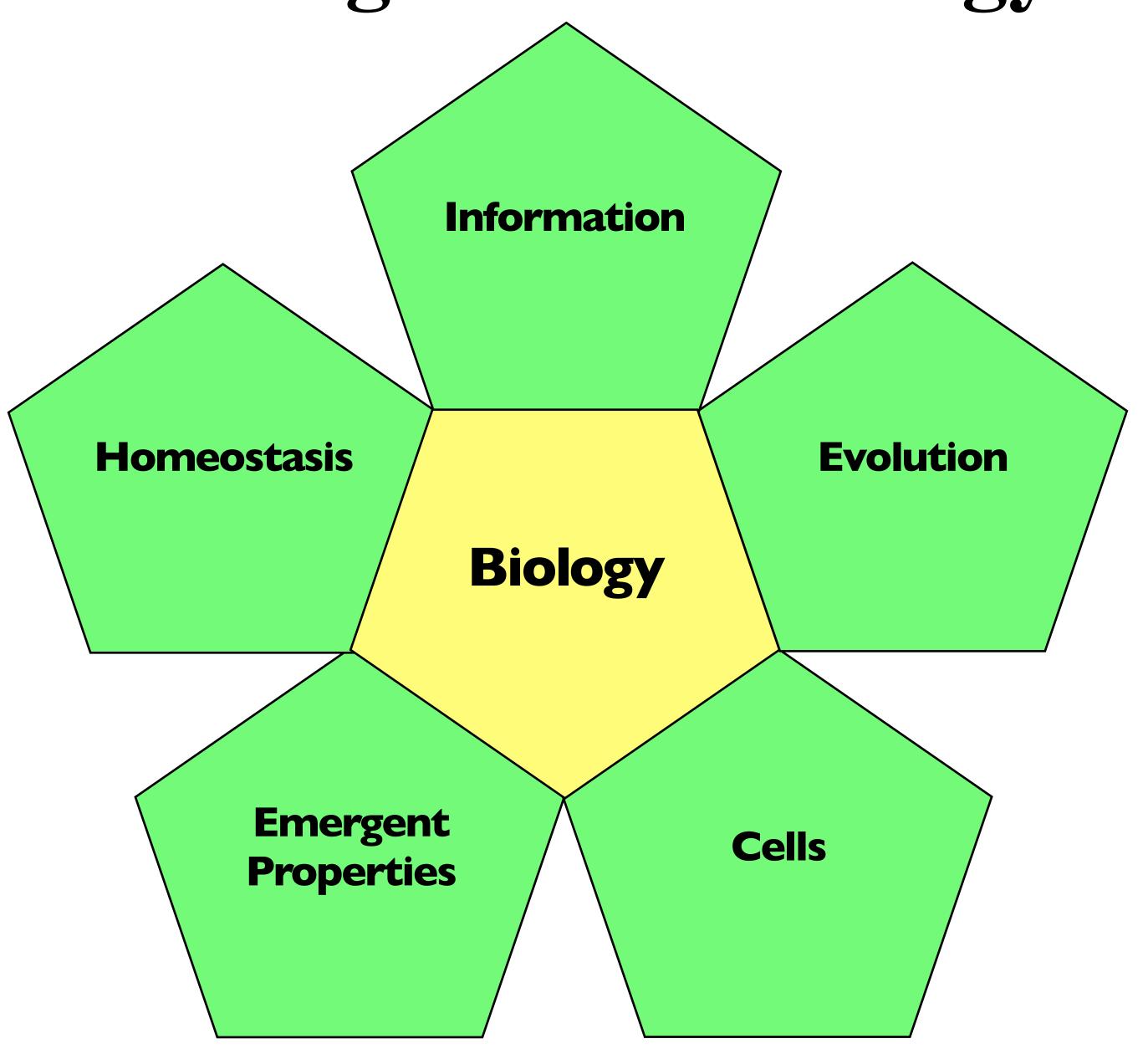


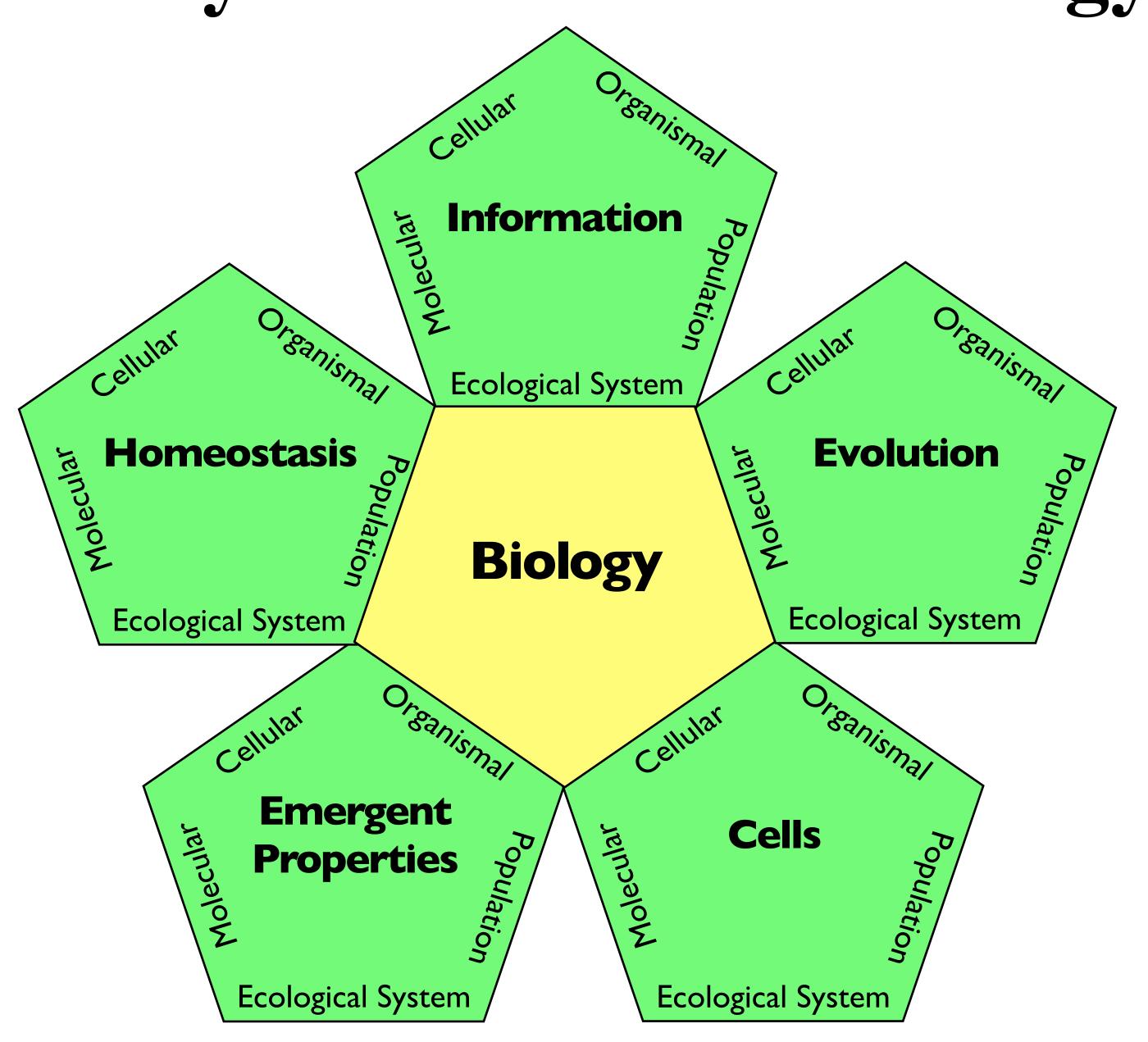


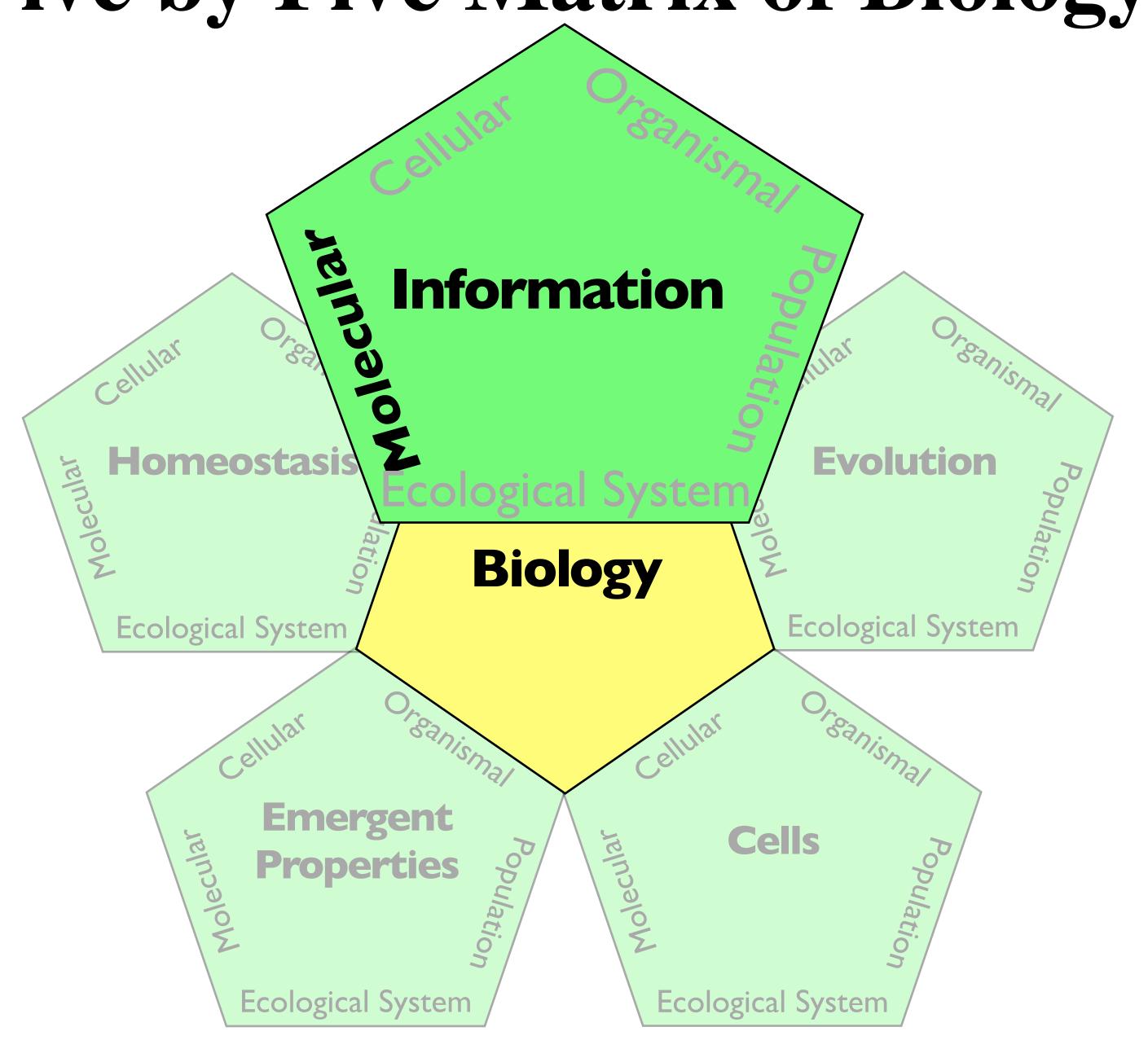
# Artificial Divide within Biology Small Biology Big Biology Sunday, March 4, 2012

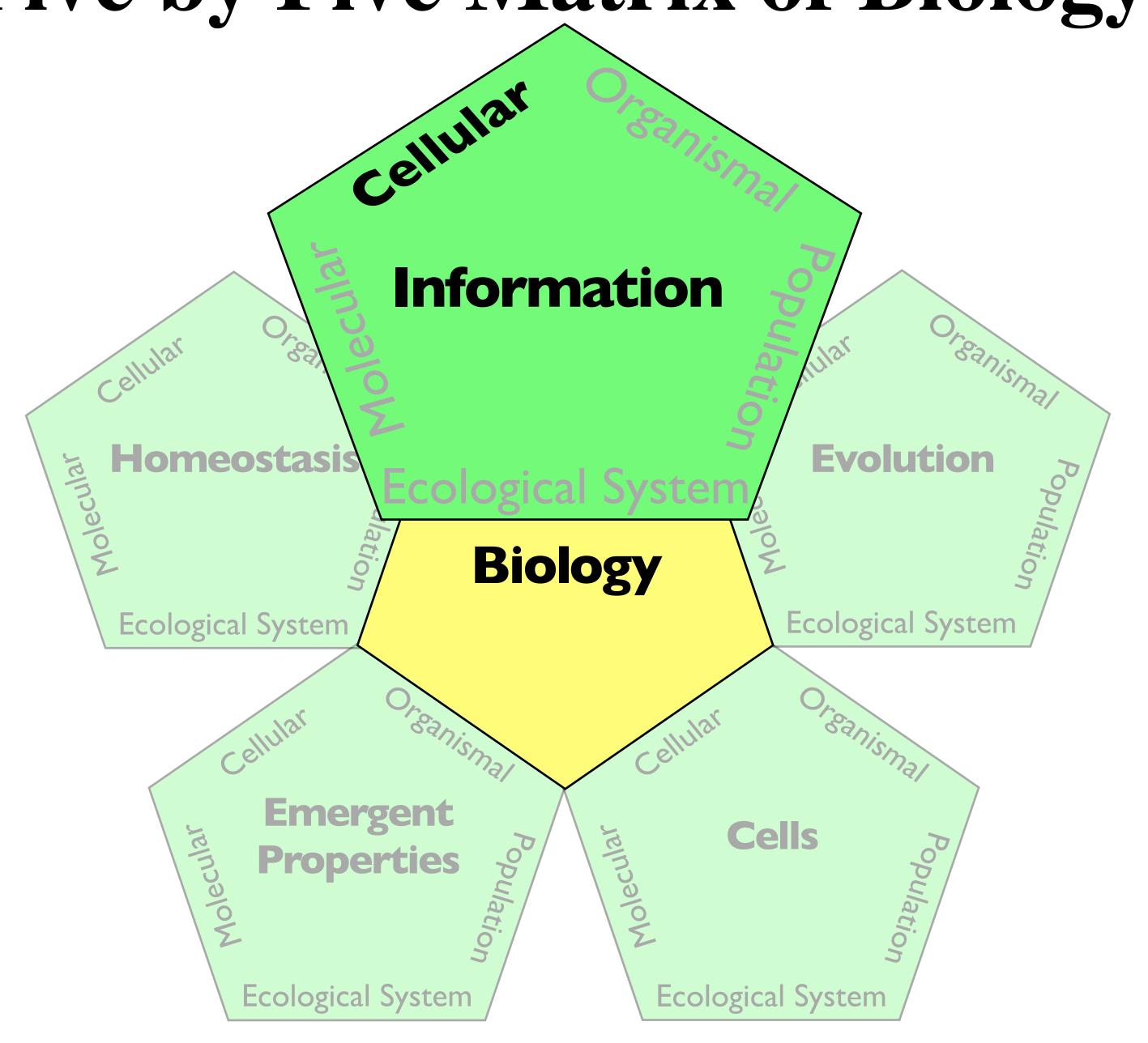


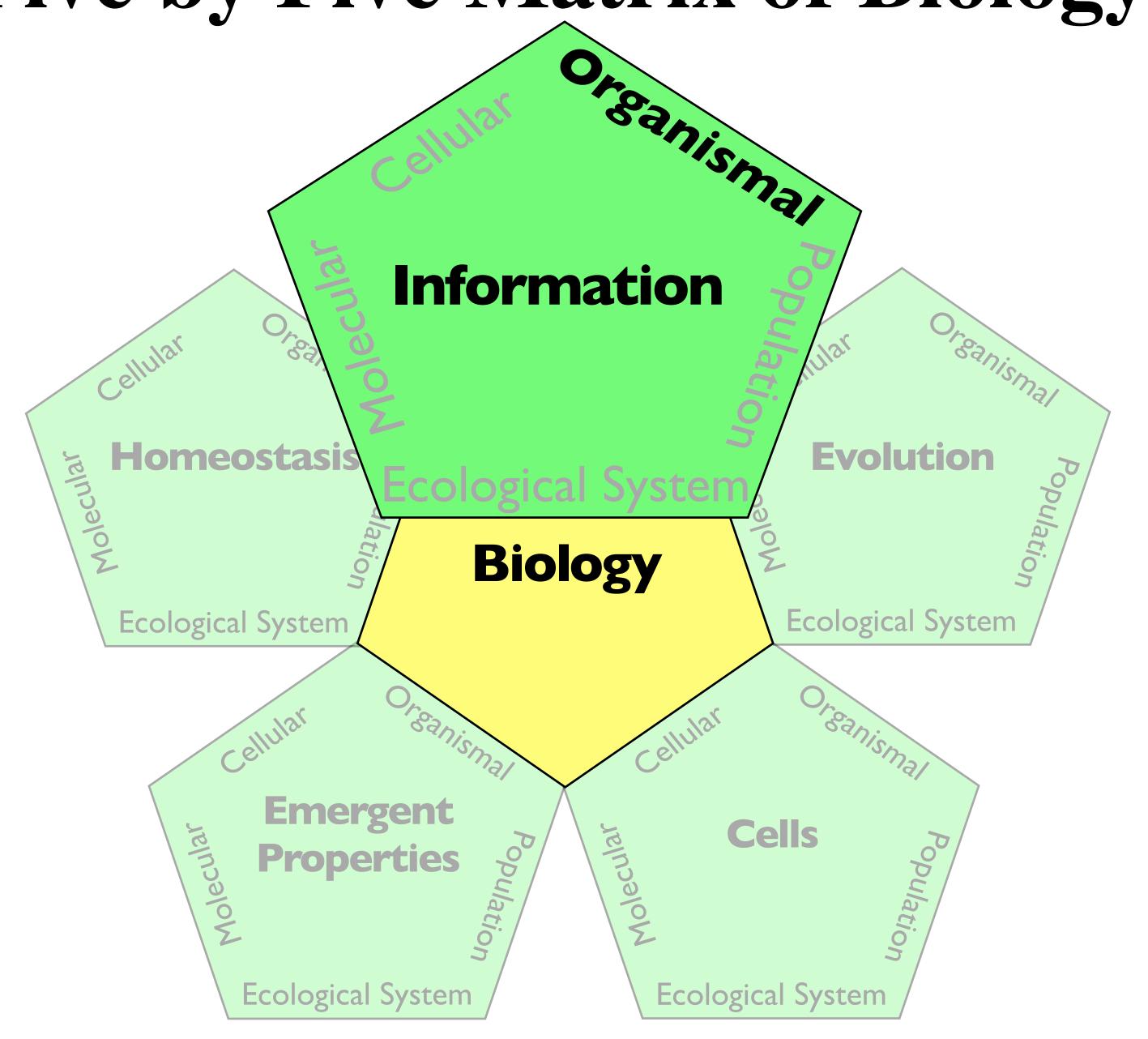
#### Five Big Ideas of Biology

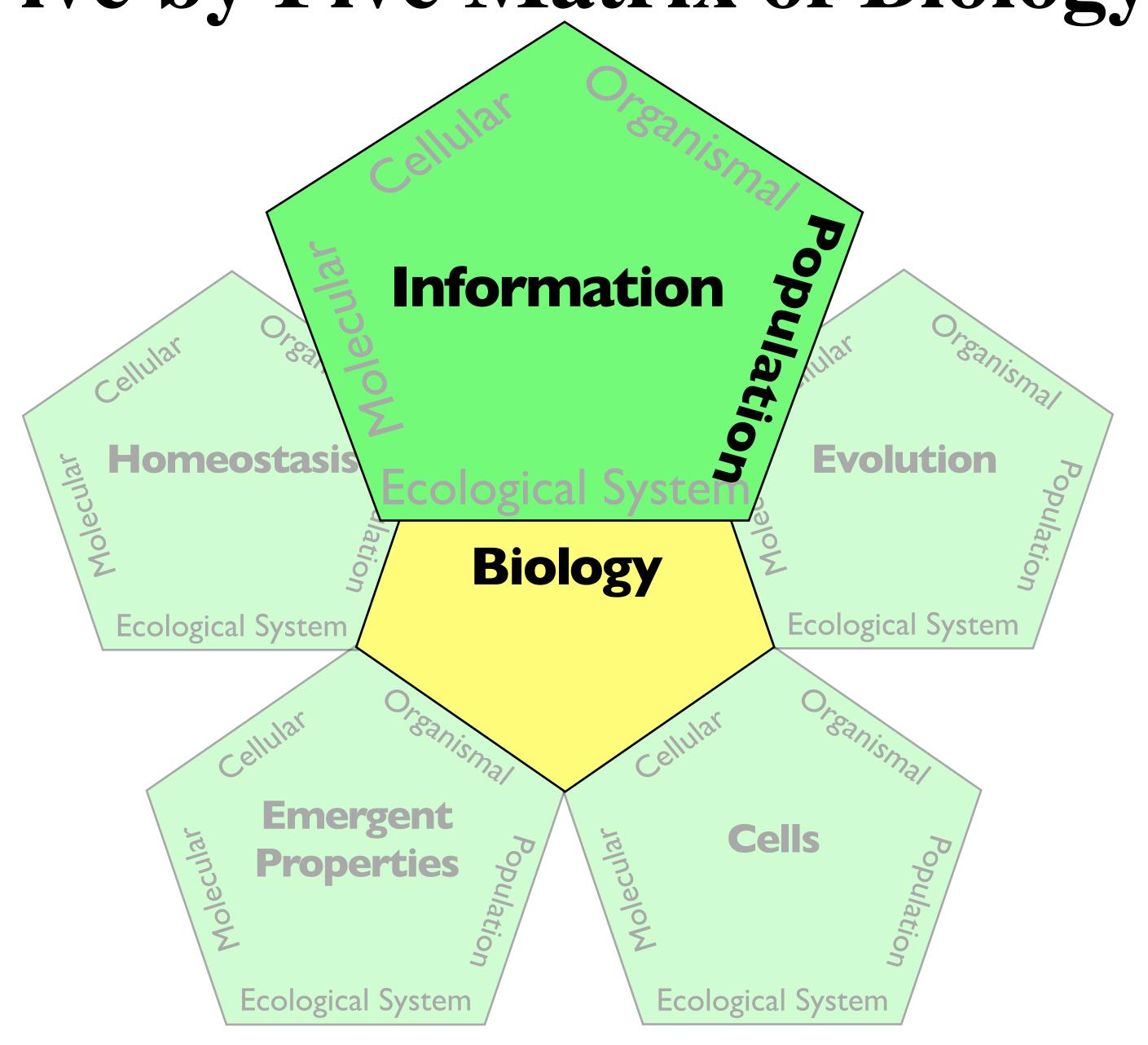


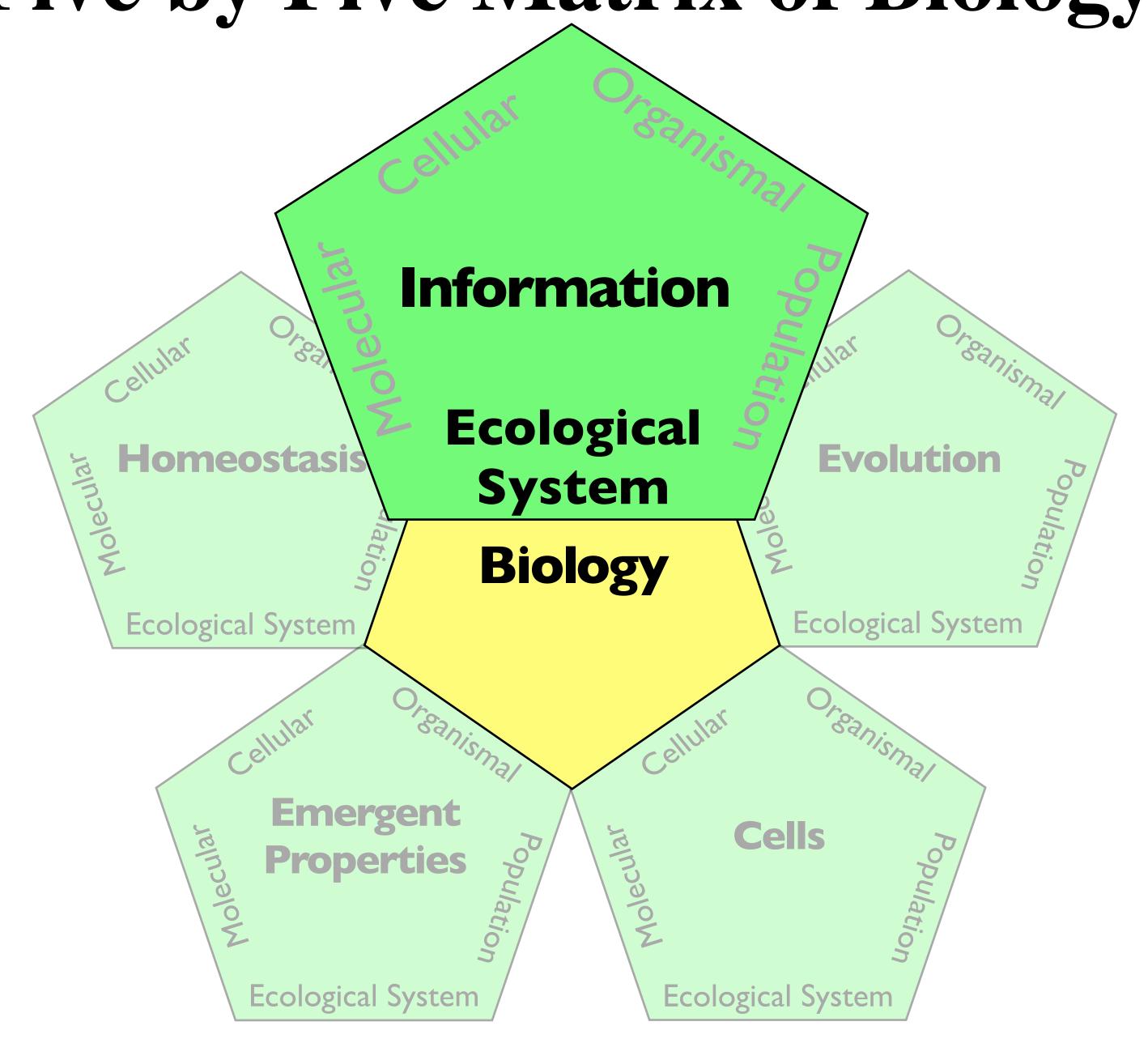


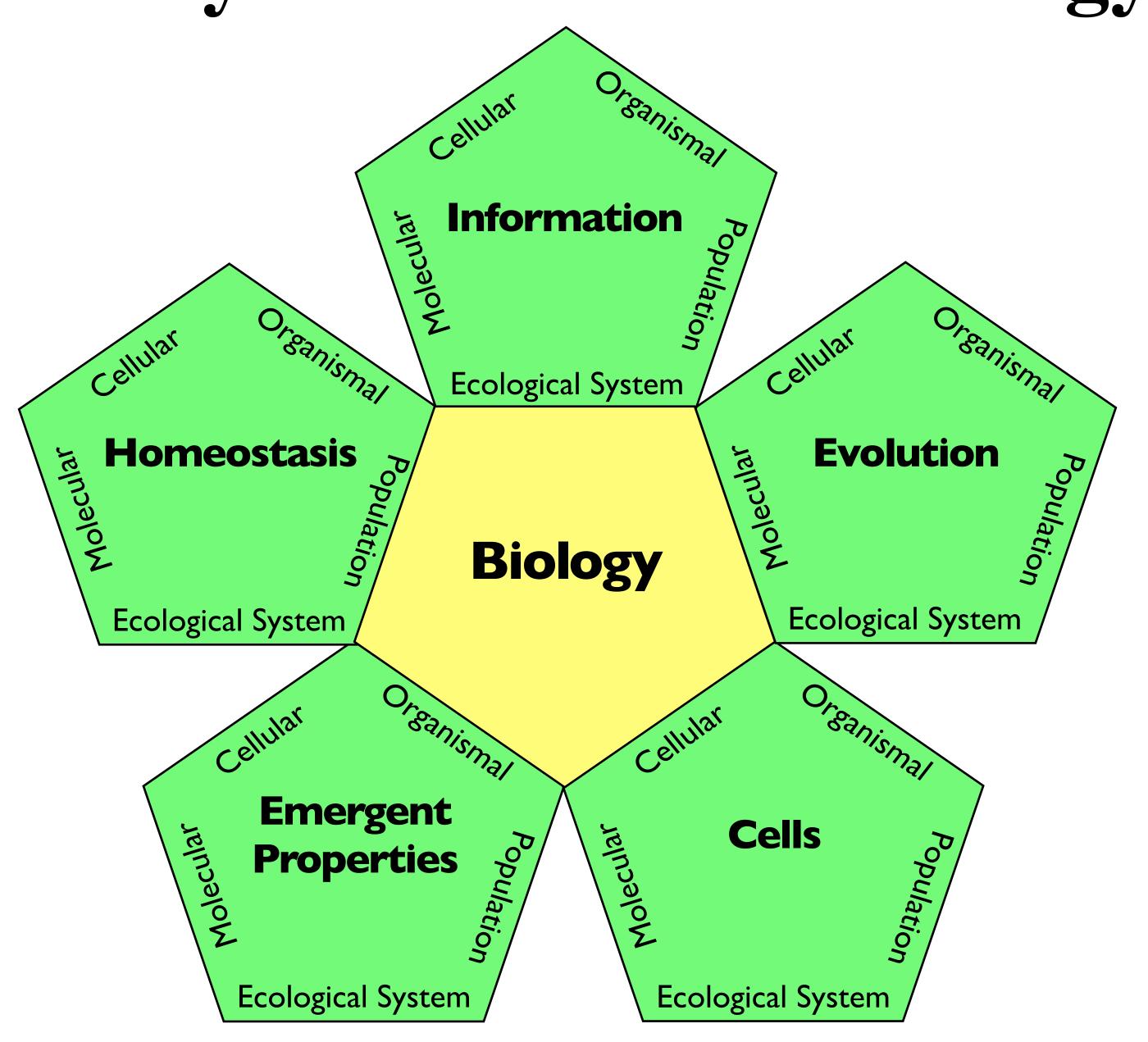








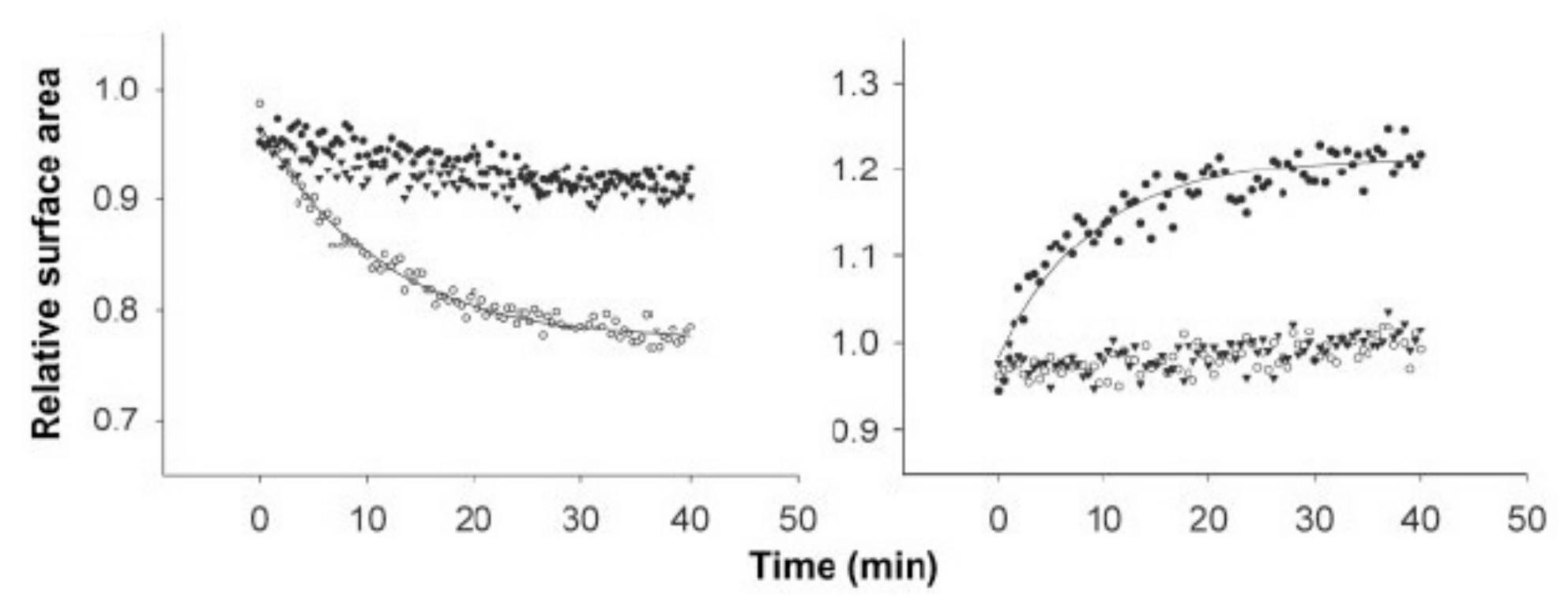




#### BioMath Explorations

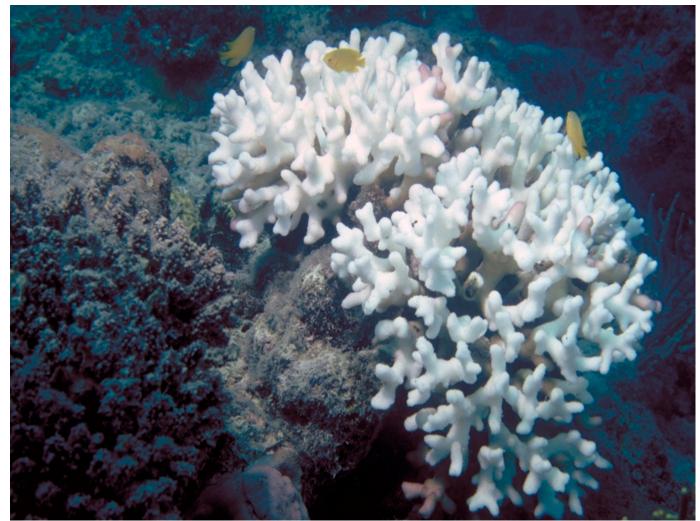
BioMath Exploration 6.3

How can you fit exponential curves to data?



#### Ethical, Legal and Social Implications



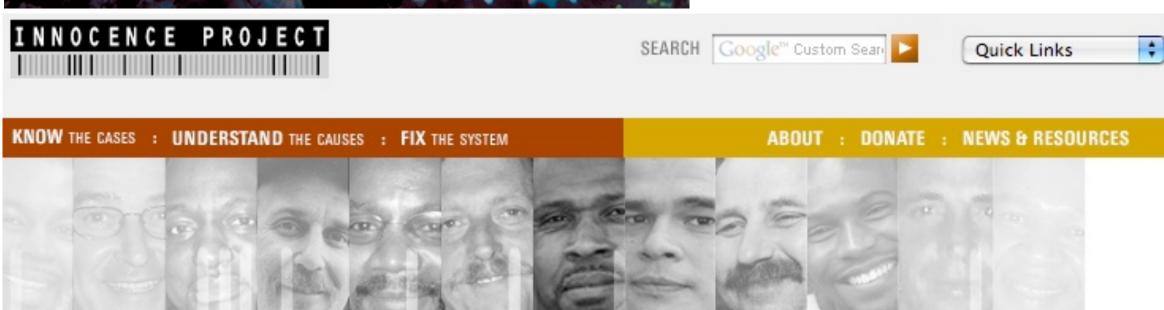


Are religion and evolution compatible?

Is science possible if you are uncertain about what is true?

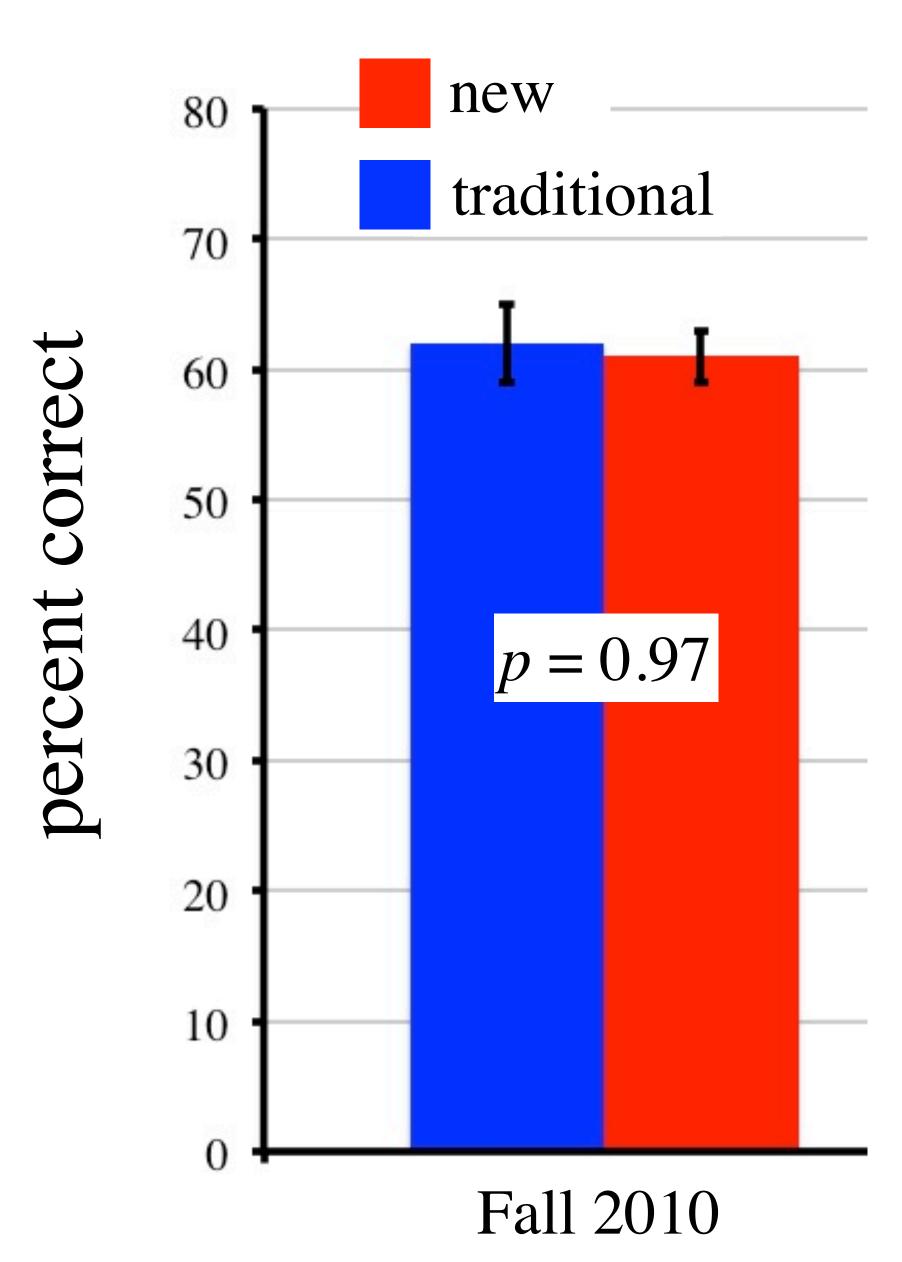
Does basic biology have any impact on the real world?

Who owns your DNA?



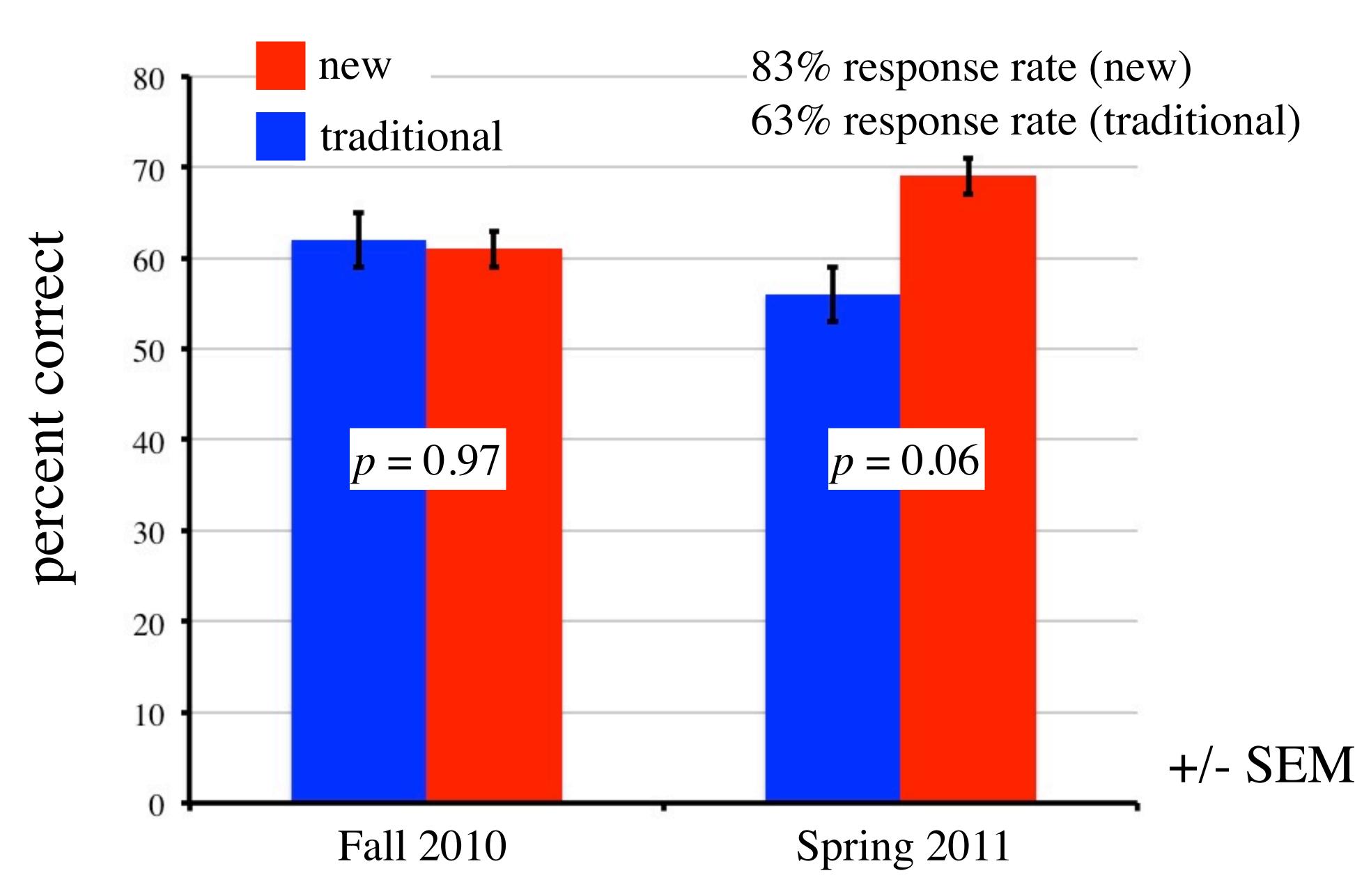
#### Did my students learn less content?

### Student Content Assessment



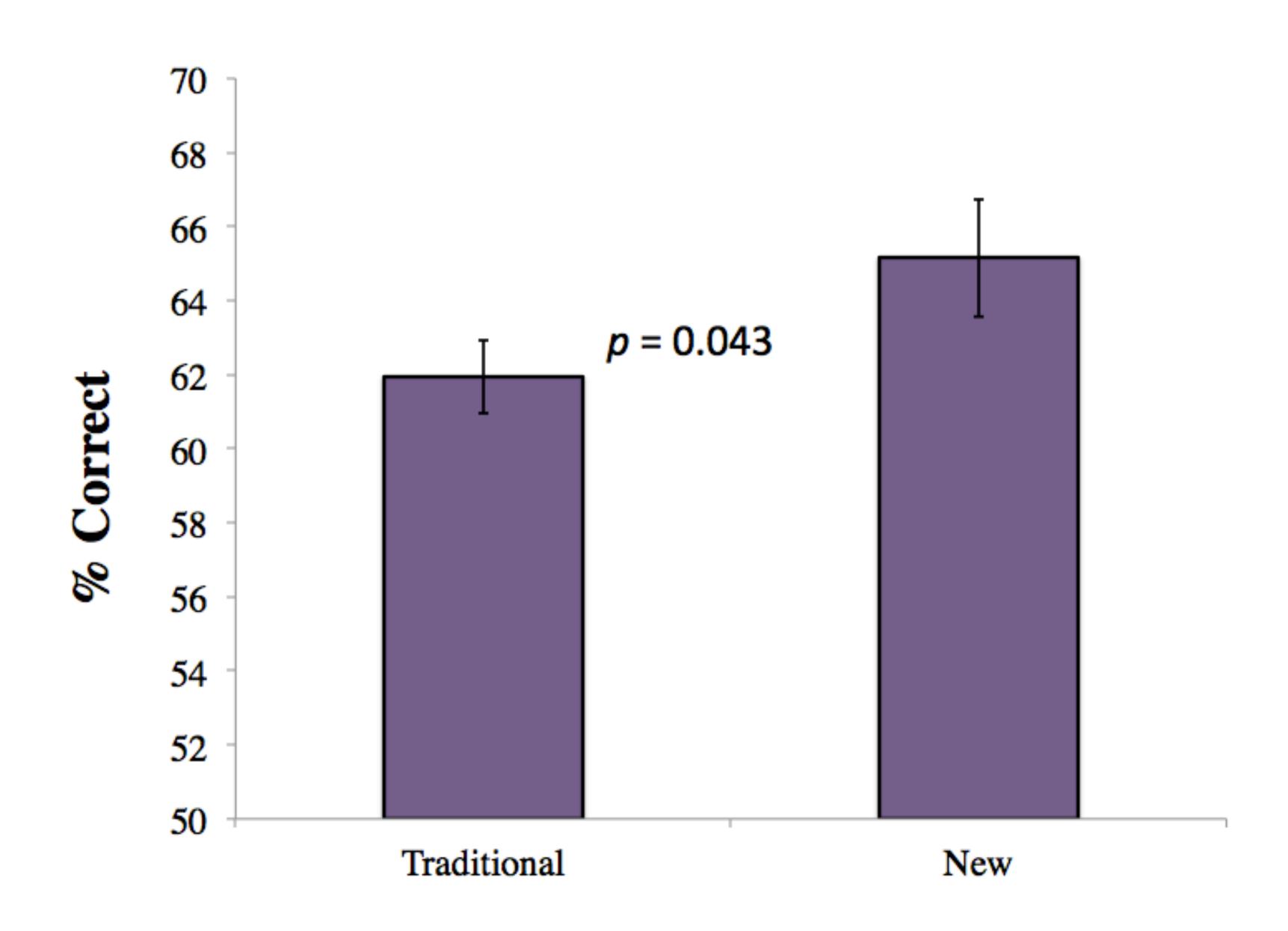
+/- SEM

### Student Content Assessment

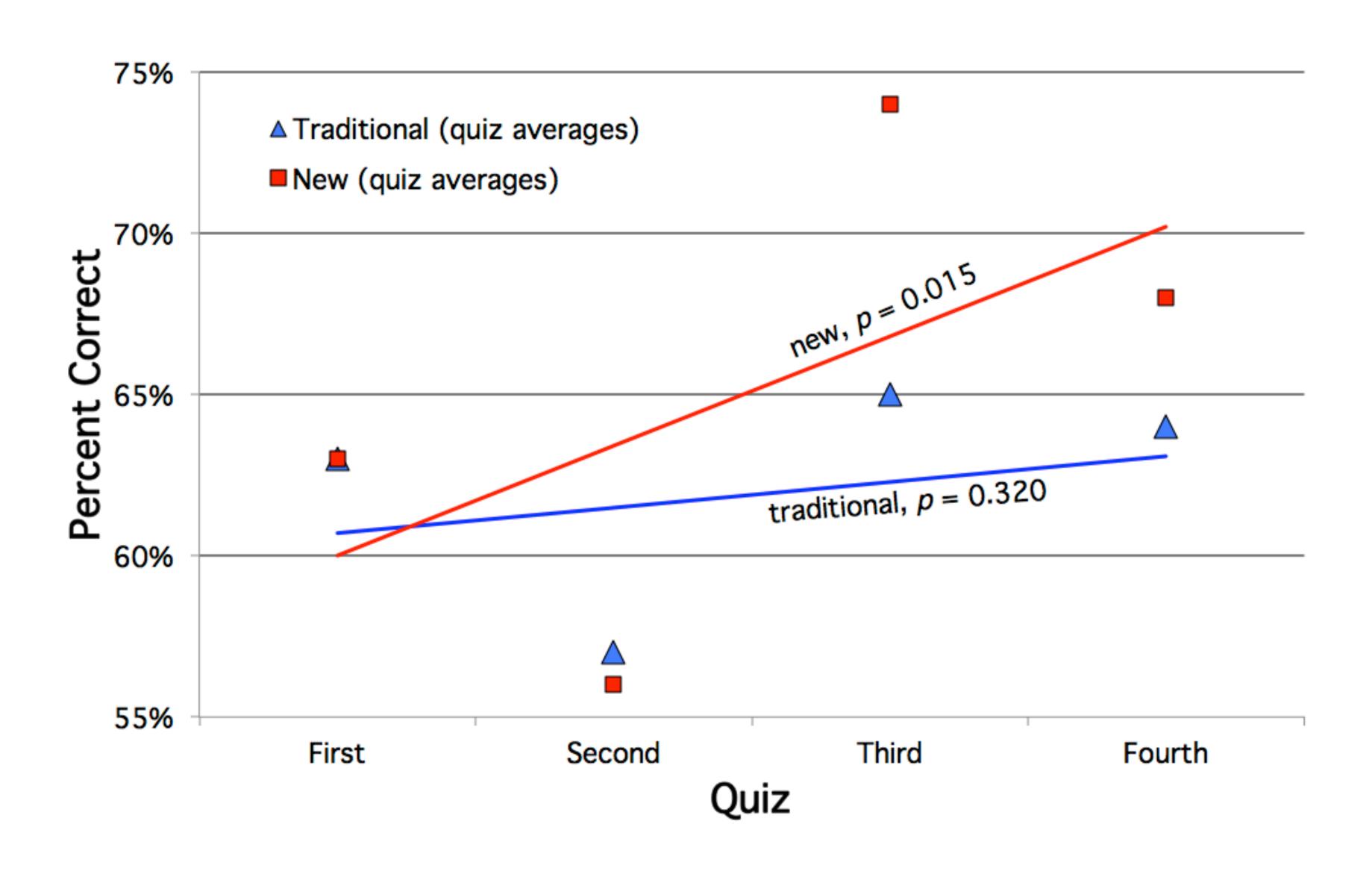


### Can my students analyze data better?

### Student Skills Assessment

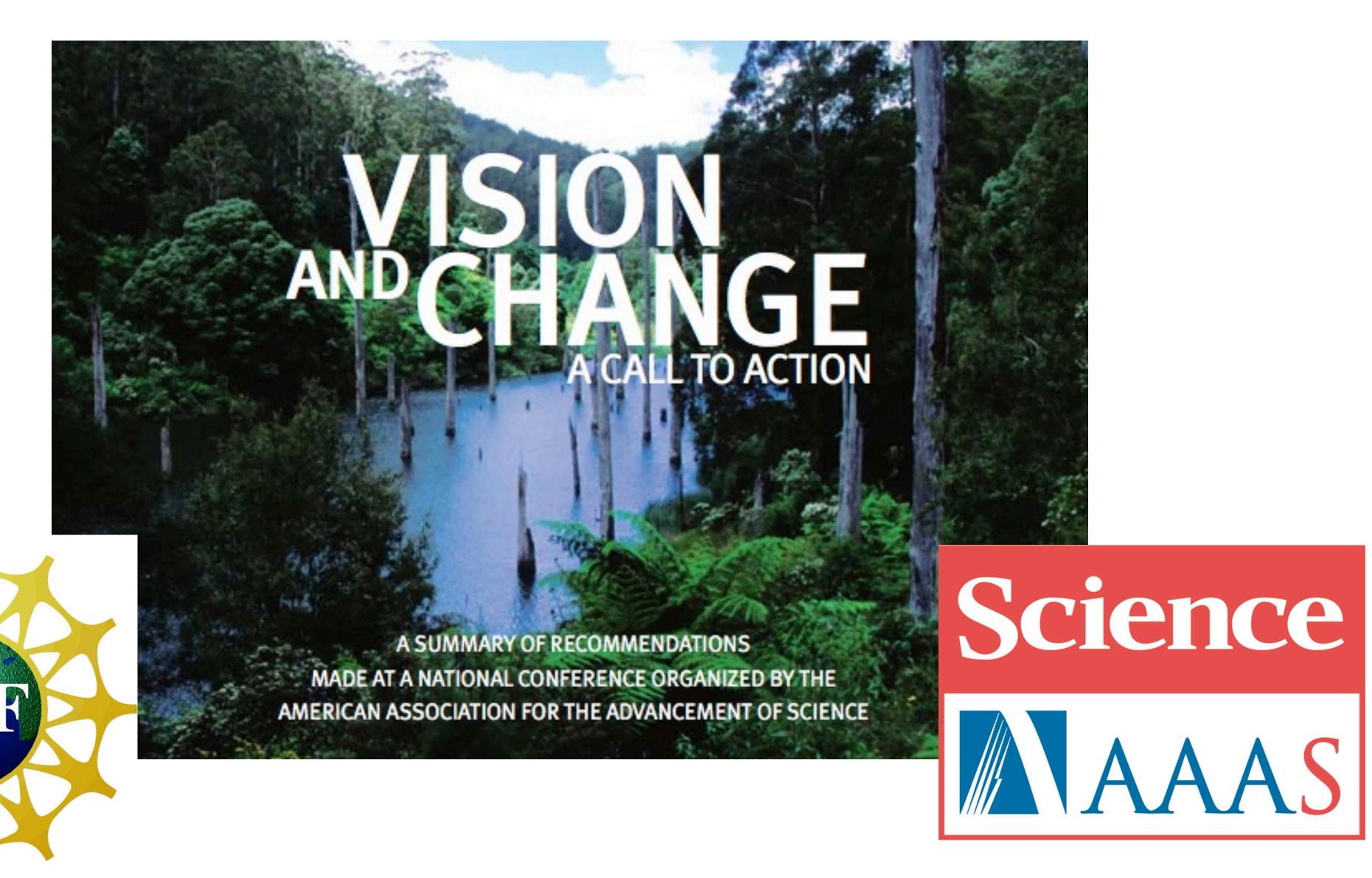


### Student Skills Assessment



### Why bother changing?

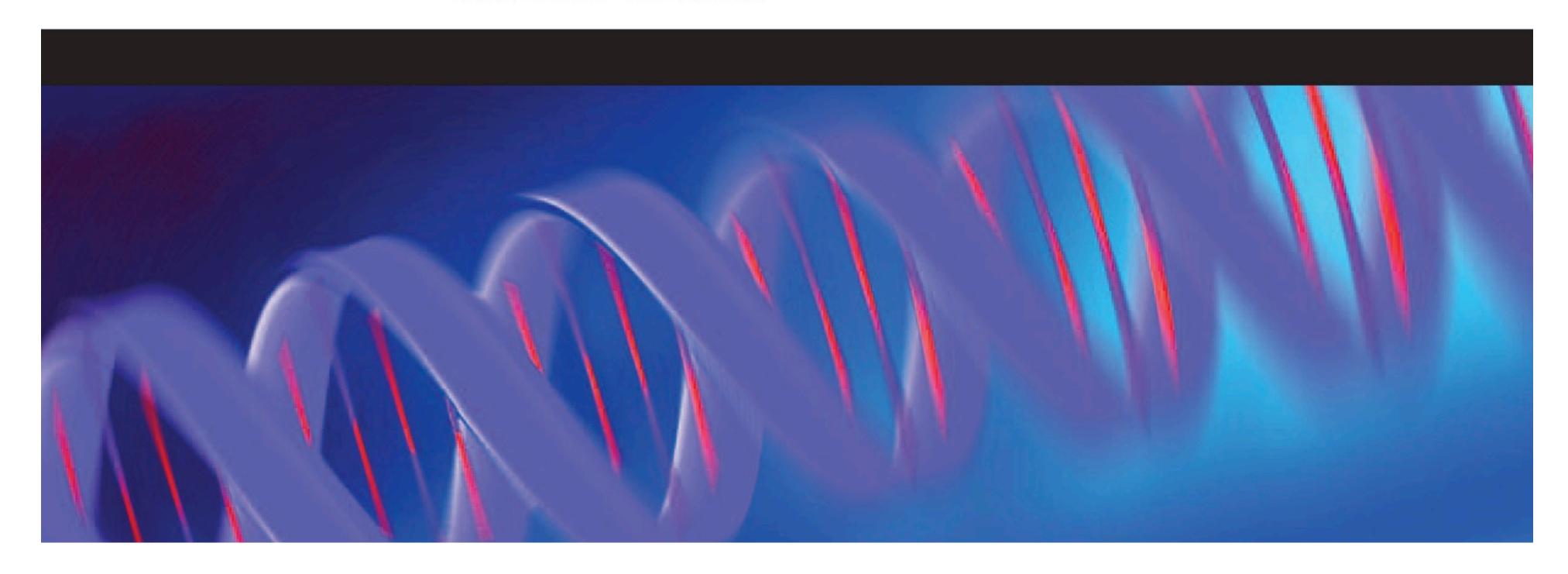
### National Recognition of Need to Change



### AP Biology is Changing to Match Our Design

\*\*\*AP® BIOLOGY

Curriculum Framework
2012–2013



# Acknowledgements

Faculty: Laurie Heyer, Jeff Poet, Todd Eckdahl, Karmella Haynes, Pat Sellers, Mark Barsoum

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Davidson College James G. Martin Genomics Program
MWSU SGA, Foundation & Summer Research Institute





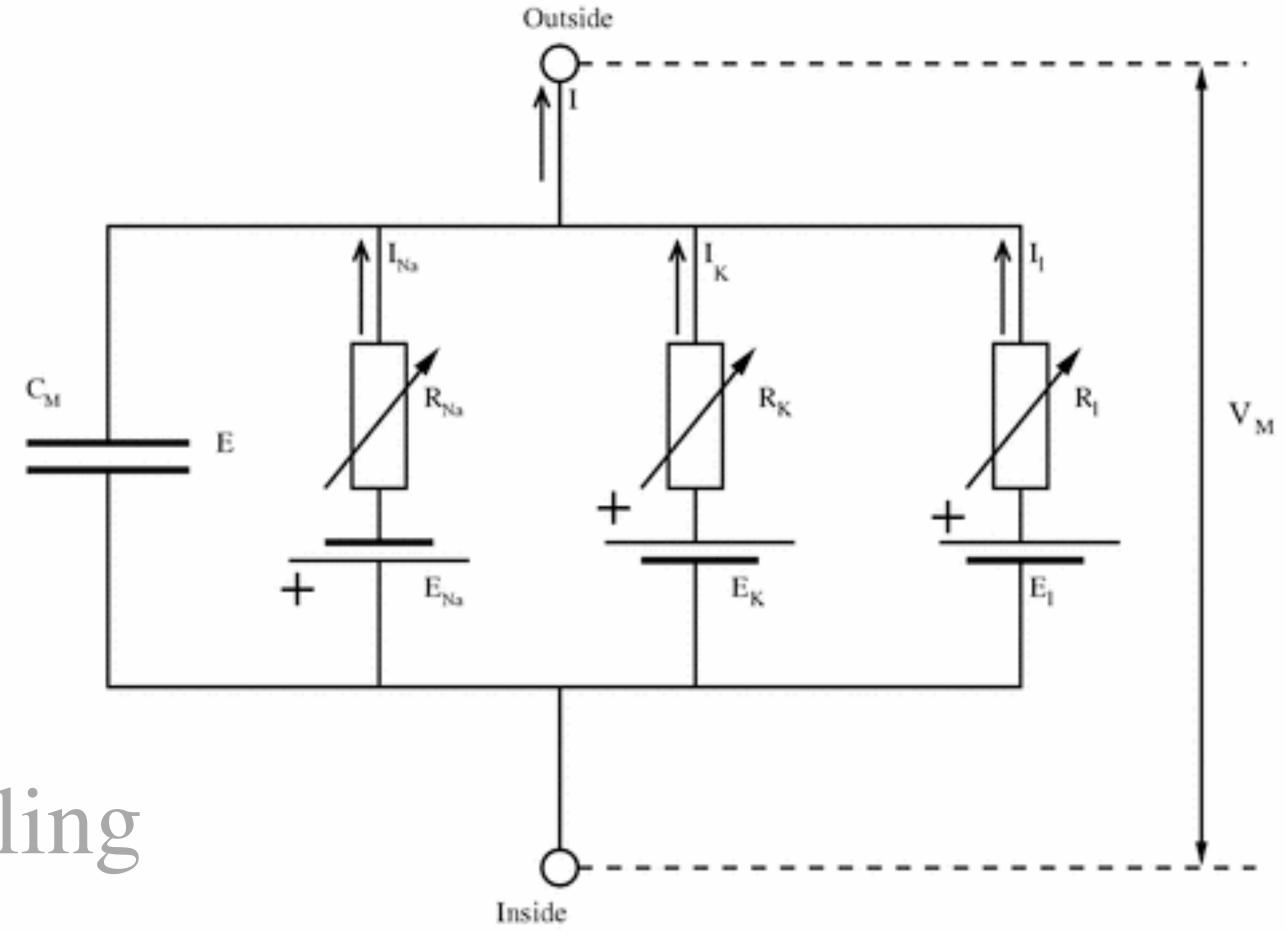




### Abstraction

Modularity

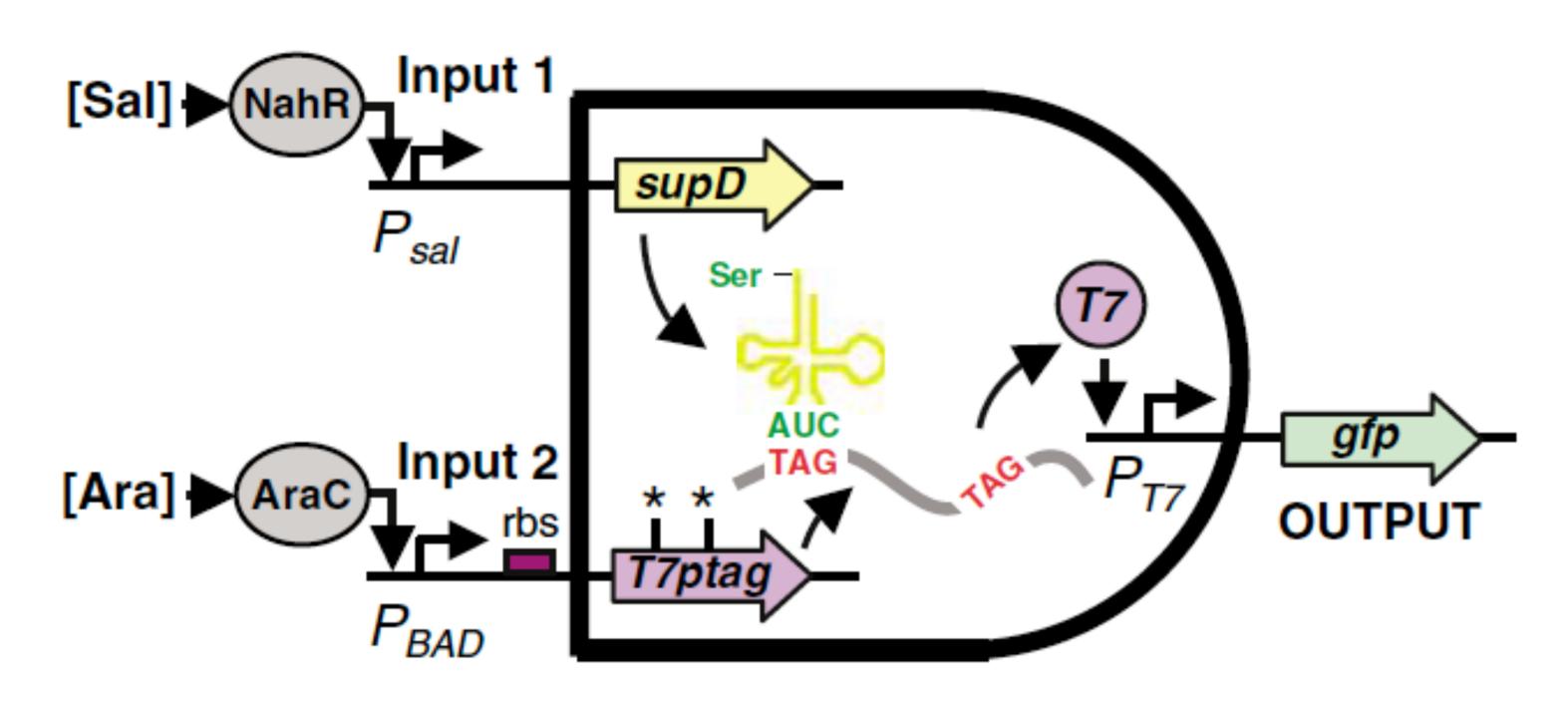
Standards



Abstraction

Modularity

Standards

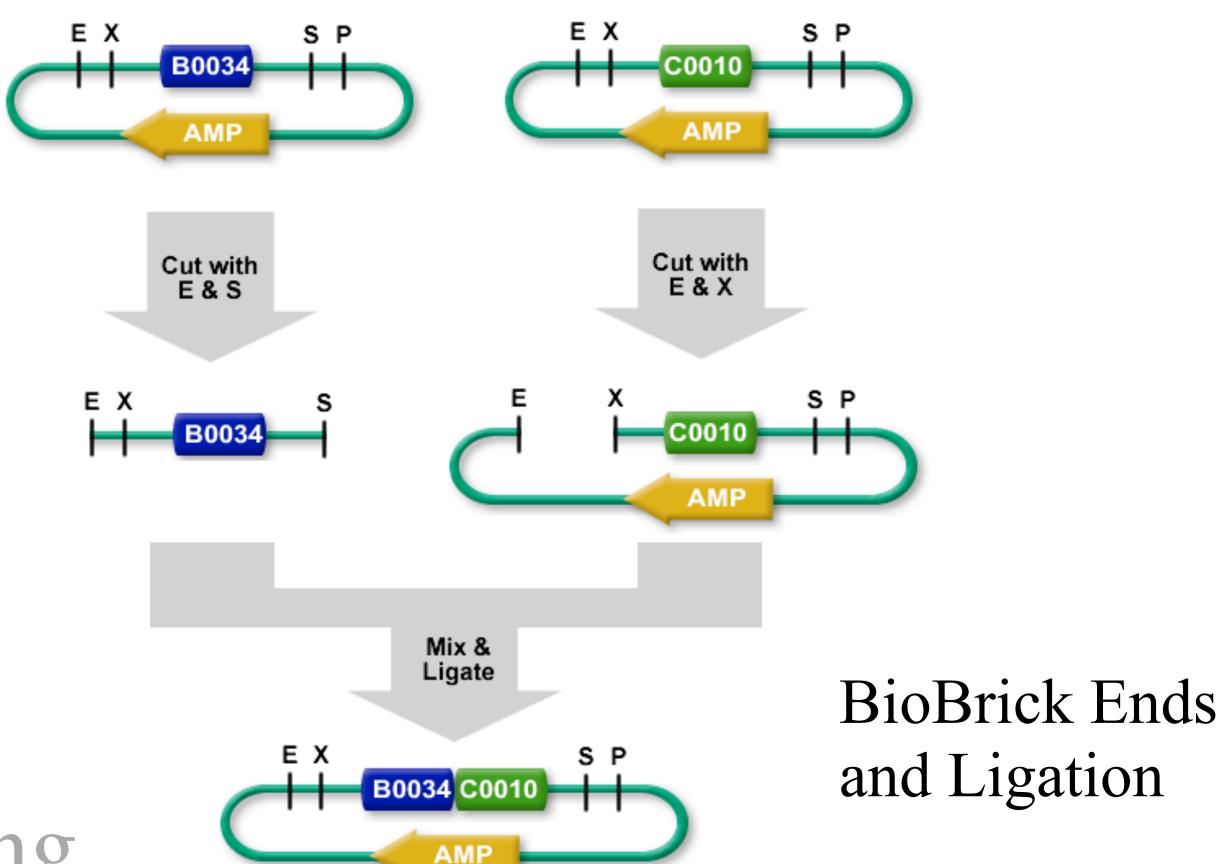


AND Logic Gate

Abstraction

Modularity

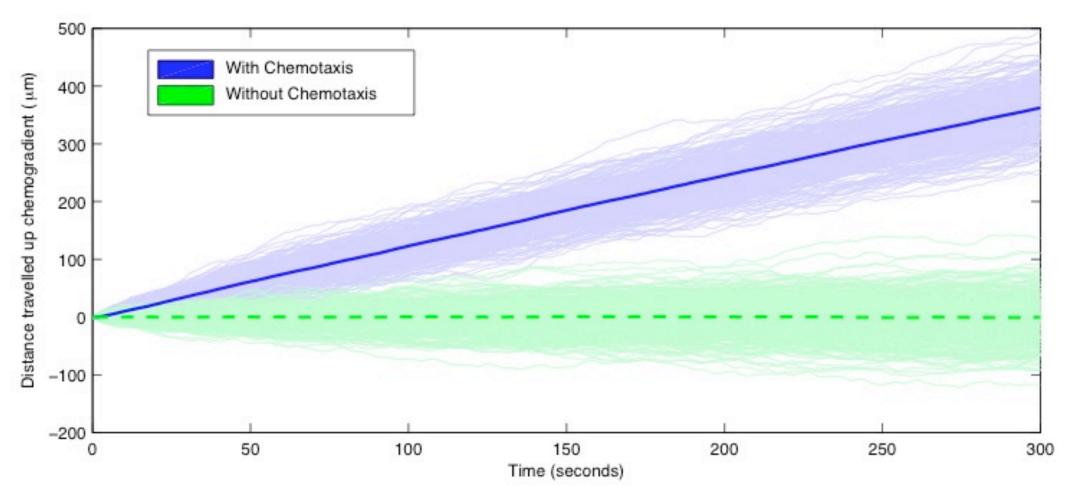
### Standards

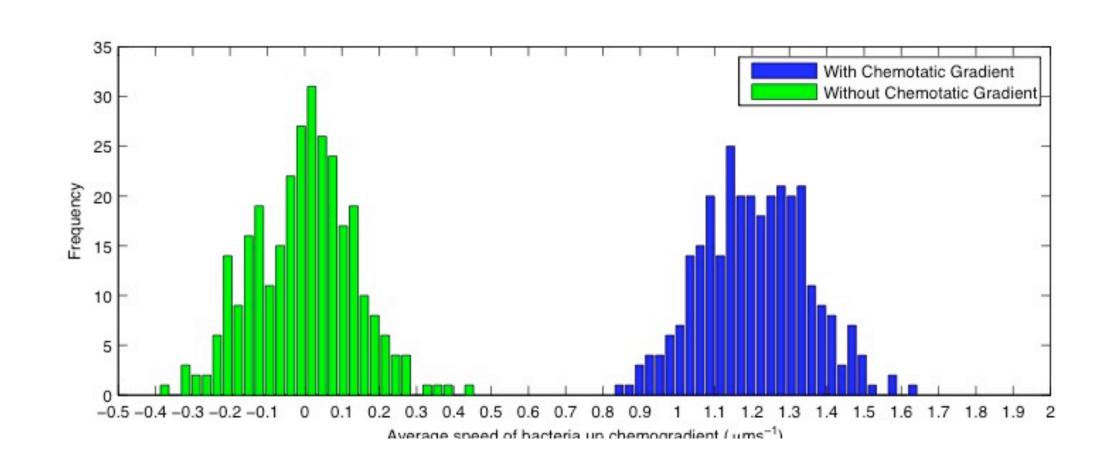


Abstraction

Modularity

Standards





### Increased Student Diversity

### 56 undergraduates in 7 years

| African  | Hispanic | First      | Asian    | Asian    |
|----------|----------|------------|----------|----------|
| American |          | Generation | Minority | Majority |
| 14       | 2        | 9          | 2        | 7        |

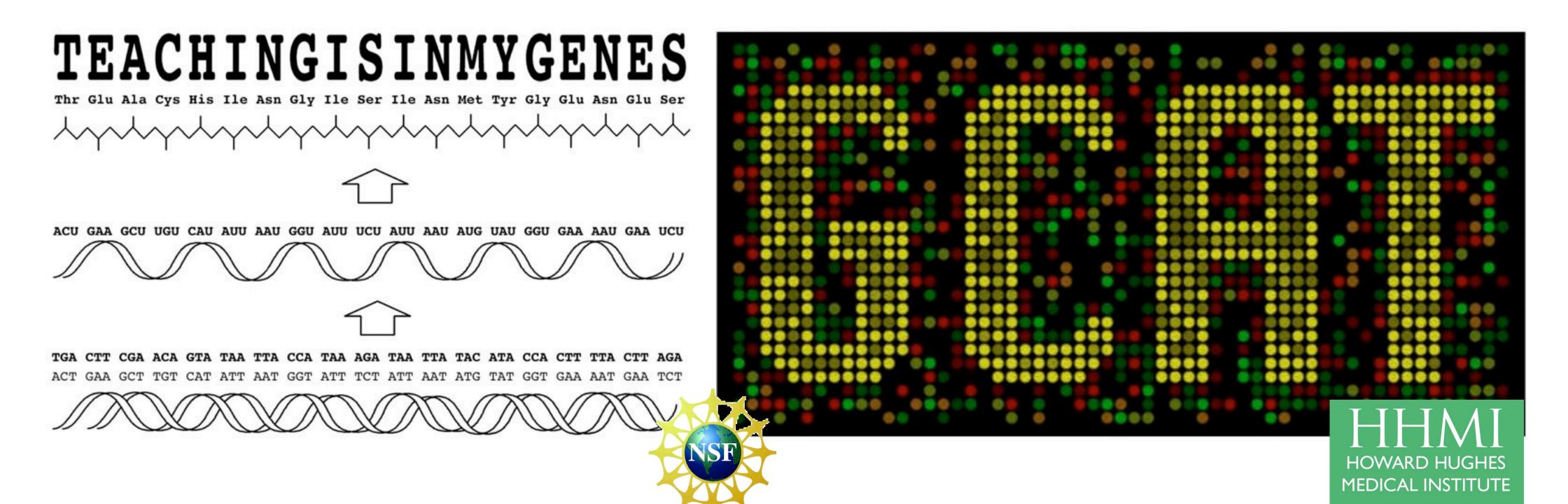
| PhD | Dual<br>degree | MD | MPH | Jobs | MD | at DC |
|-----|----------------|----|-----|------|----|-------|
| 13  | 2              | 2  | 3   | 5    | 7  | 27    |

campus: 74% Caucasian

biology majors: 87% Caucasian

# GCAT Faculty Workshop Synthetic Biology

15 pairs of faculty
1 Bio + 1 Other
NSF & HHMI



# What did my students think about this approach to intro bio?

"The method of learning, placing emphasis on the interpretation of data, has helped me not only in this class, but also in others."

anonymous student course evaluation, Dec. 2010

"I found it much more beneficial using this approach compared to straight memorization. It allowed me to gain interpretation skills I was lacking before."

anonymous student course evaluation, Dec. 2010

"The data-driven approach is brilliant. It alleviates the issues that I've always had of asking, 'How do we know that? What's the supporting data?'"

anonymous student course evaluation, Dec. 2010

"Emphasis on big picture and understanding how to pull information from real data was an easier and more beneficial format than memorization of facts (which used to be a struggle for me)."

### How did I test student learning?

### Four Exams Per Semester

#### 8 pts.

9) Limit your answers to a maximum of 2 sentences for each part.

a) Explain why it is adaptive for each eukaryotic organelle to be composed of a different lipid

composition. Use data to support your answer.

Each one has a particular surface area to volume ratio and different lipids have different bending capacity. Rigid lipids produce larger volumes while relaxed lipids produce bends and small volumes inside membranes.

| Lipid Name                               | Rat Liver<br>ER* | Rat Liver Plasma<br>Membrane* | Rat Liver<br>Golgi* | Mouse Skin plasma<br>membrane <sup>b</sup> | Yeast Inner<br>Mitochondria <sup>b</sup> | Yeast Outer<br>Mitochondria <sup>b</sup> | Yeast Inner<br>Nuclear <sup>b</sup> |
|--|------------------|-------------------------------|---------------------|--|--|--|-------------------------------------|
| phosphatidylcholine                      | 58               | 39                            | 50                  | 43.0                                       | 38.4                                     | 45.6                                     | 44,6                                |
| phosphatidylethanolamine                 | 22               | 23                            | 20                  | 16.1                                       | 24.0                                     | 32.6                                     | 26.9                                |
| sphyngomyelin                            | 3                | 16                            | 8                   | 12.2                                       | 0  | 0  | 0                                   |
| phosphatidylinositol                     | 10               | 8                             | 12                  | 7.6  | 16.2                                     | 10.2                                     | 15.1                                |
| phosphatidylserine                       | 3                | 9                             | - 6                 | 6.4  | 3.8                                      | 1.2                                      | 5.9                                 |
| phosphatidic acid                        | 0                | 0                             | 0                   | 0.0  | 1.5                                      | 4.4                                      | 2.2                                 |
| cholesterol                              | n.d.             | n.d.                          | n.d.                | 13   |  |  |                                     |
| cholesterol or<br>diphosphatidylglycerol | n.d.             | n.d.                          | n.d.                | -  | 16.1                                     | 5.9                                      | 1.0                                 |

a: from Gerrit van Meer, 1998, Table 1. b: from Orientations of Proteins in Membranes, 2010, http://opm.phar.umich.edu/stlas.php. "relaxed" vesicle

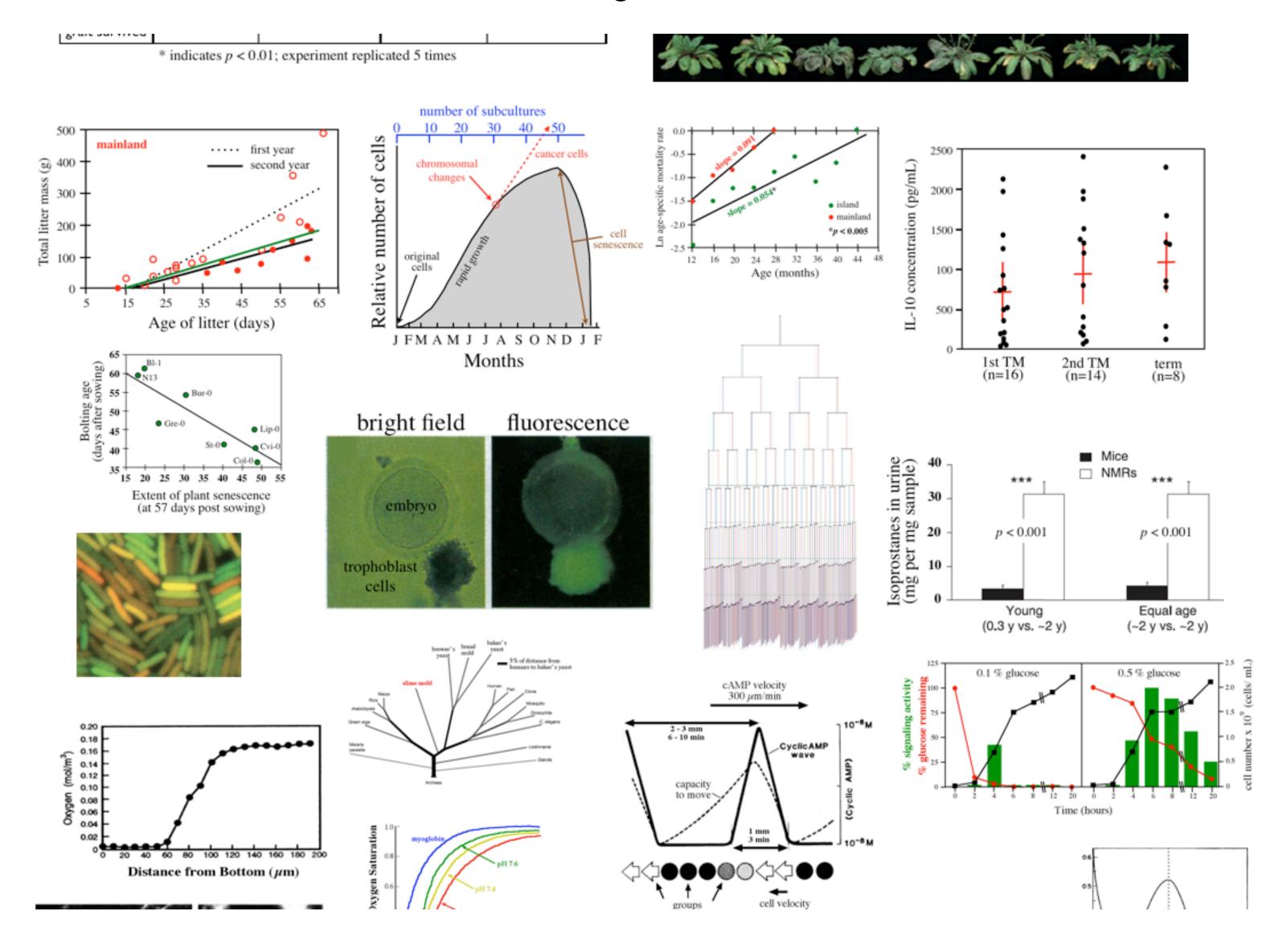
"relaxed" vesicle

Time (s)

b) Would you predict that the secretory vesicles containing epinephrine would contain more rigid lipids, or flexible lipids? Use data to support your answer.

relaxed due to large surface area to volume ratio

## Data Gallery for Answers



# When did the students feel they were learning something different than in high school?

### Table of Contents

#### Chapter 7 Evolution at the Cellular Level

- 7.1: How are new species formed? Discover how genomes can change dramatically to produce new species.
  - BME 7.1: What information is in a dot plot? Discover how to construct and interpret a dot plot for comparing whole genomes.
  - ELSI 7.1: Are GMOs safe?
- 7.2 Why doesn't your stomach digest itself? Analyze experimental results showing that eukaryotes evolved a shared mechanism to retain proteins inside the endoplasmic reticulum.
  - BME 7.2: Cause or effect? Explore the meaning of correlation, and how it is quantified.
- 7.3 Why do my allergies get worse each year? Determine that B cells evolve in days to produce stronger immune responses.
  - ELSI 7.2: Banning PB&J: How far should a society go to protect the rights of an individual?
- 7.4 Why are corals dying around the world? Realize that species can coevolve as symbionts and become interdependent.
  - BME 7.3: Can you predict coral bleaching? Evaluate the fit and predictive ability of a trendline.

### Table of Contents

### Chapter 17 Emergent Properties at the Cellular Level

- 17.1 Do unicellular species have to work solo? Realize that microbes use quorum-sensing, biofilms and communal behavior to enhance their functions.
- 17.2 How can changes in two cells affect an entire plant? Appreciate how guard cells change their shape to regulate plant gas exchange through stomata.
  - BME 17.1: Can local decisions have global effects? Model the opening of stomata using a simulation of local rules.
- 17.3 How do brain cells store memories? Discover how long-term memories are formed by analyzing classic experiments on *Aplysia* learning.
  - ELSI 17.1: If pills could make you remember or forget, would you take them?
- 17.4 Does the genome allow random actions by cells? Learn how random movements of molecules determine cell phenotypes which can be transmitted across generations.

BME 17.2: What is chaos?

### Table of Contents

### Chapter 22 Homeostasis at the Cellular Level

- 22.1 Why is paraquat used in America but illegal in Europe? Analyze classic experiments to deduce how light energy is captured by plant cells.
- 22.2 How does Brazil's rainforest affect Greenland's glaciers? Determine how carbon dioxide is fixed by photosynthetic cells into biological molecules.
  - ELSI 22.1: How do you compromise when a policy hurts one country but helps another?
- 22.3 Is there anywhere on earth devoid of life? Explore inhospitable niches where microbes have evolved homeostatic mechanisms to survive harsh conditions.

### Student Skills Assessment

