

# Synthetic Biology: Insights into Natural Biology and New Technologies



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# Three Rules for Our Lab

1. Everyone has to learn.
2. Everyone has to have fun.
3. We try to contribute to the body of science.

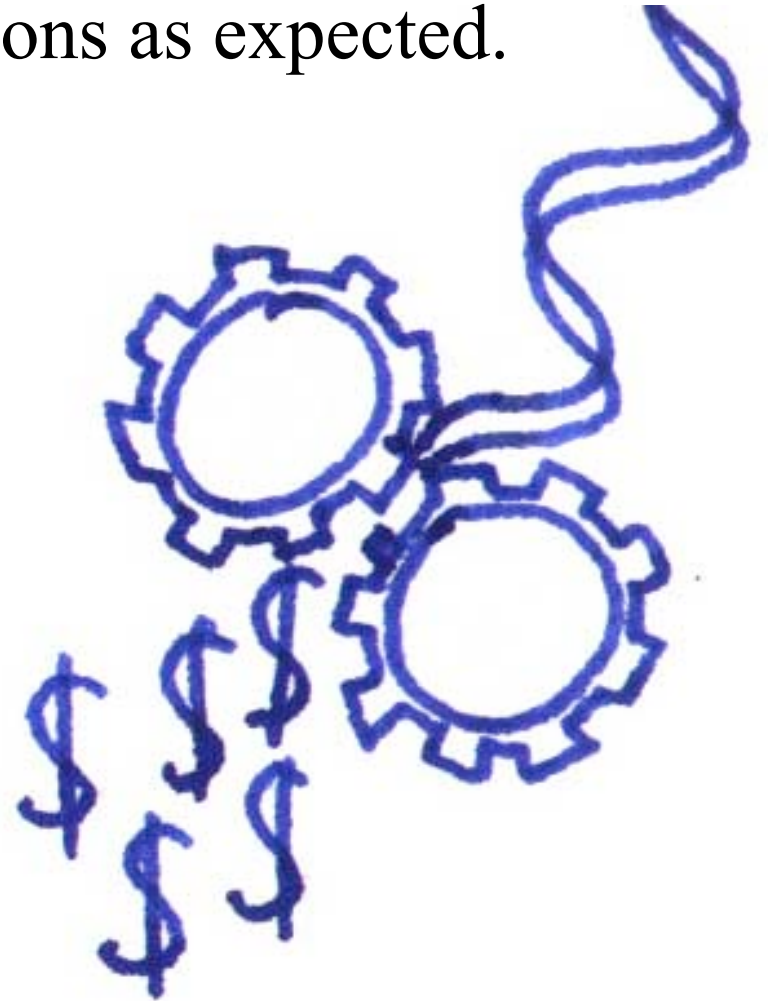
# What is Synthetic Biology?

Application of **engineering principles** and **mathematical modeling** to the design and construction of **biological parts, devices, and systems** with applications in energy, medicine, and technology.

[www.bio.davidson.edu/projects/gcat/Synthetic/What\\_Is\\_SynBio.html](http://www.bio.davidson.edu/projects/gcat/Synthetic/What_Is_SynBio.html)

# Synthetic Biology: Win-Win

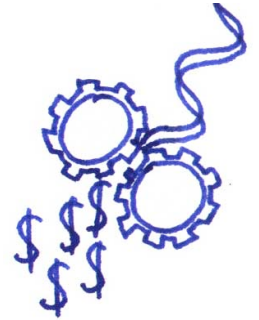
Win #1: your design functions as expected.



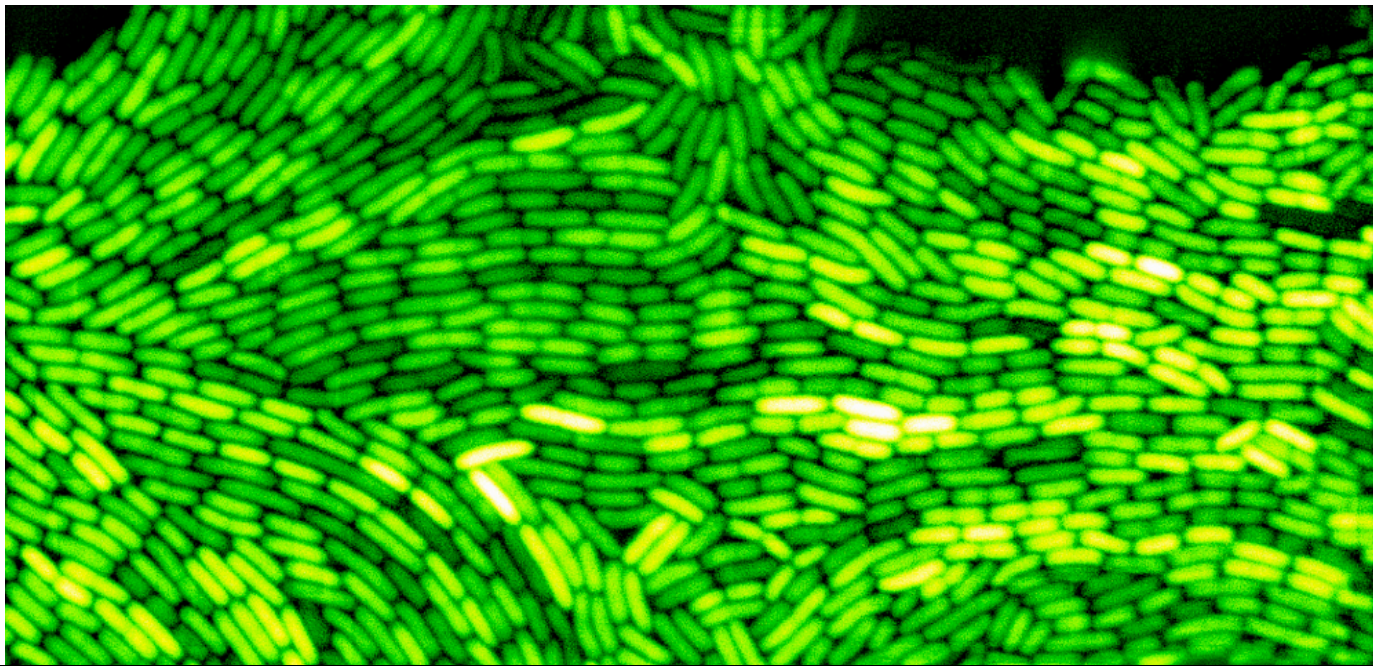
# Synthetic Biology: Win-Win Research



Win #1: your design functions as expected.



Win #2: your design fails but you uncover basic biology



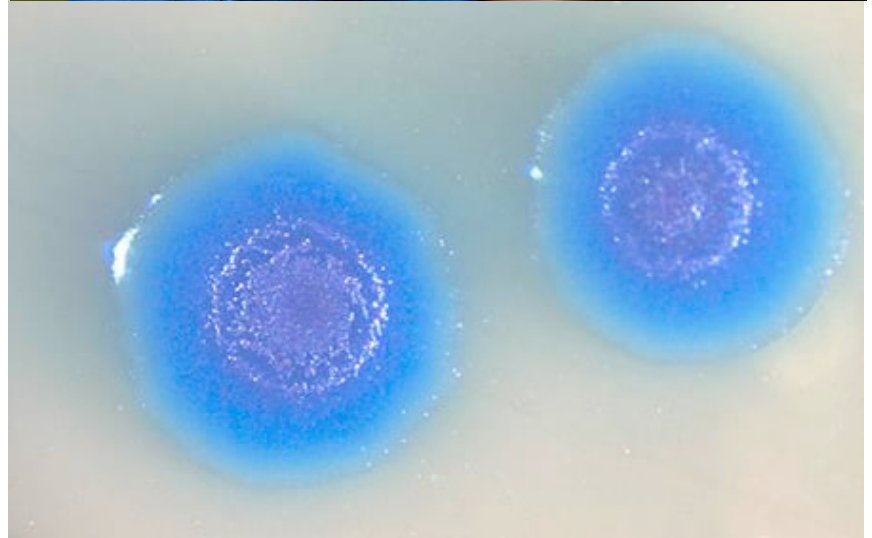
# How is Synthetic Biology Different?

Abstraction

Modularity

Standards

Designing and modeling



# Abstraction



# Abstraction





# Modularity



USB ports on computers

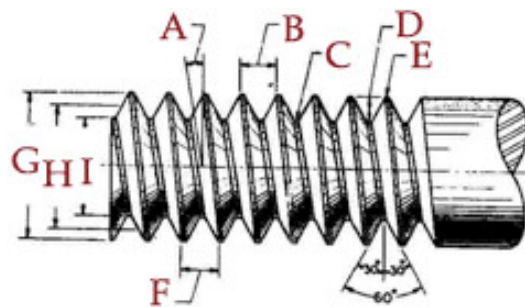
# Modularity



# Standardization

## *On a Uniform System of Screw Thread*

“In this country, no organized attempt has as of yet been made to establish any system, each manufacturer having adopted whatever his judgment may have dictated as best, or as most convenient for himself.”



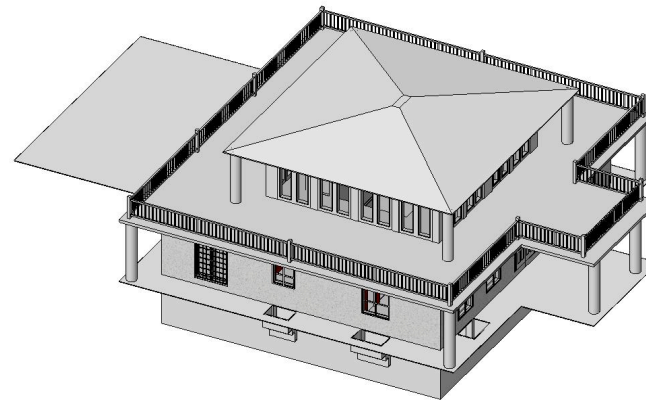
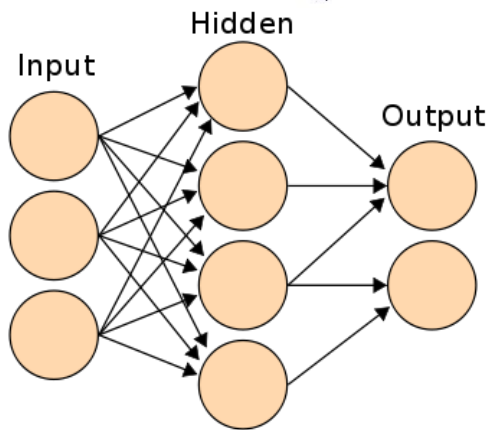
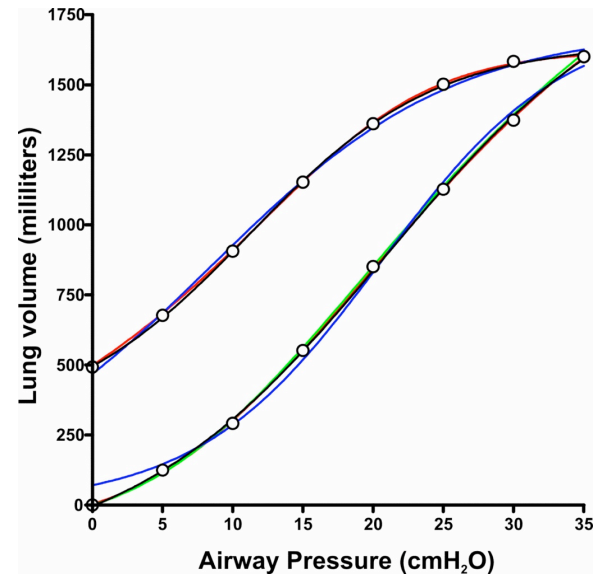
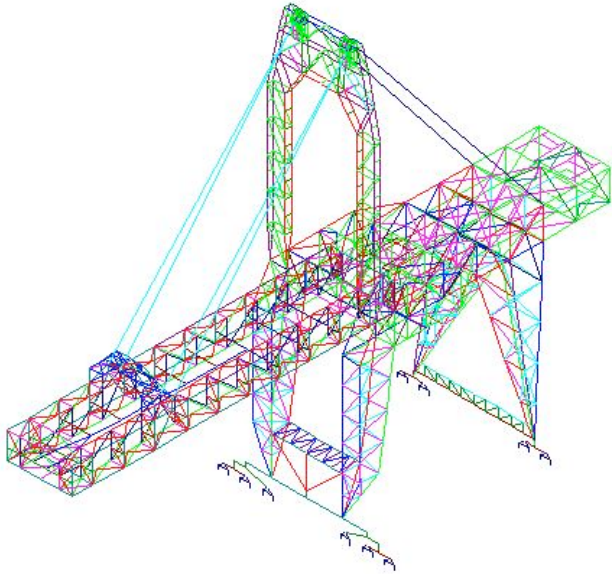
William Sellers April 21, 1864

# Standardization



*On a Uniform System of Screw Thread*

# Modeling of Designs



# Real World Applications of Synthetic Biology

# Land Mine Detection



# Land Mine Detection





# Synthetic Biology Land Mine Detection



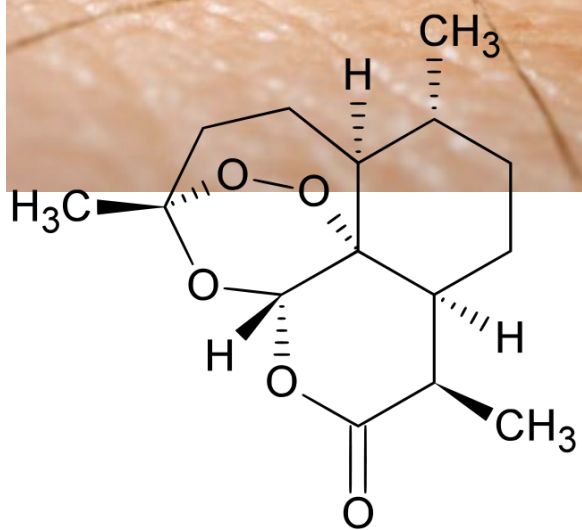
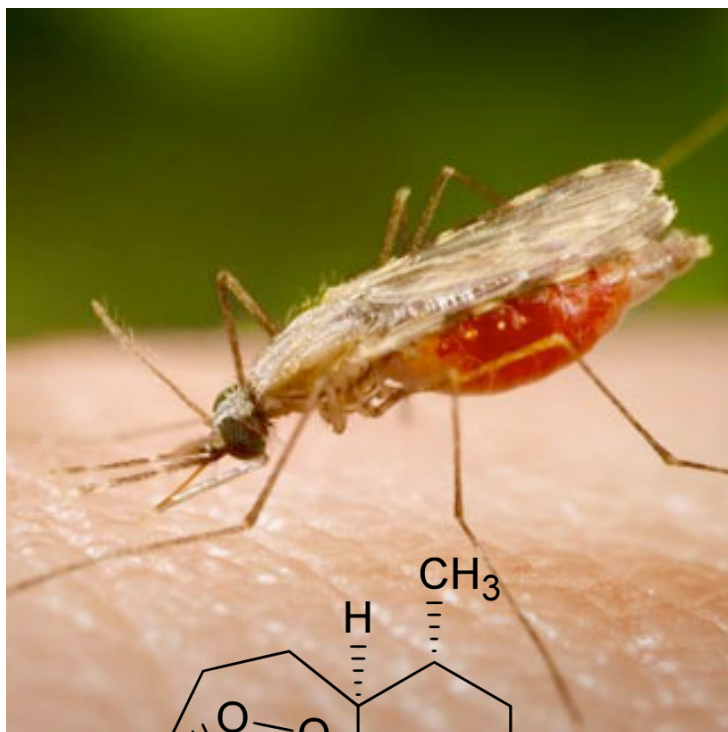
**WARNING SIGN:** The bioengineered Thales cress turns red when exposed to a mine byproduct.

COURTESY OF ARESA  
BIODETECTION

## New weed may flag land mines

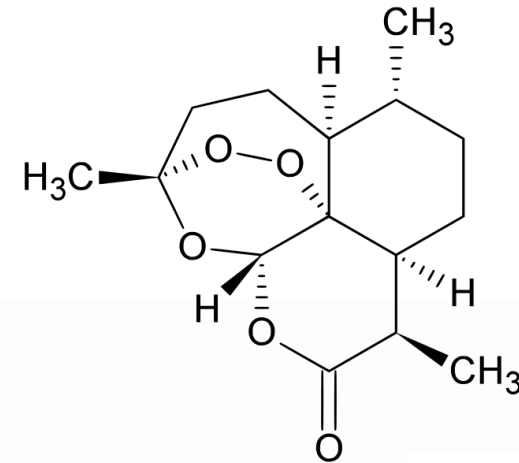
By John K. Borchardt | *Contributor to The Christian Science Monitor*

# Production of Medicines

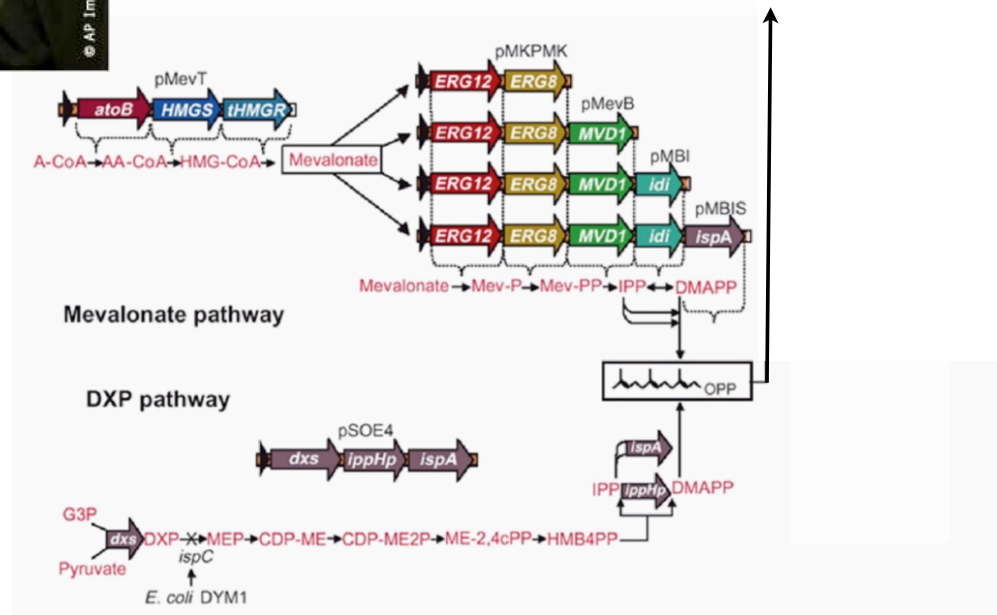


\$1 per pill

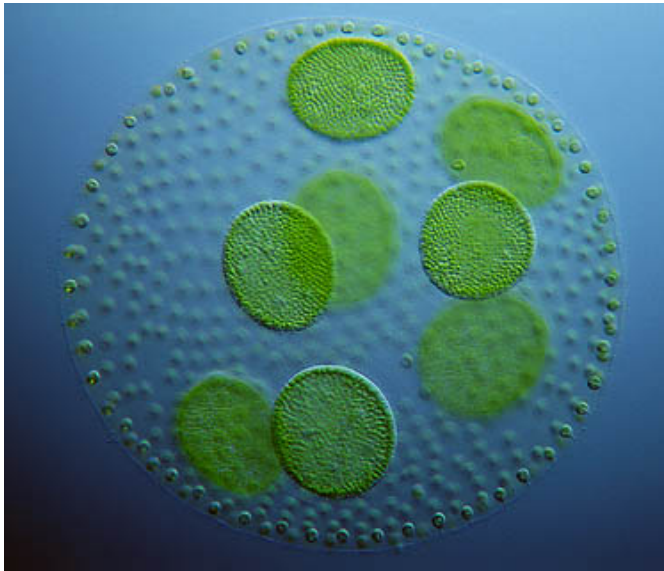
# Production of Medicines



10¢ per pill



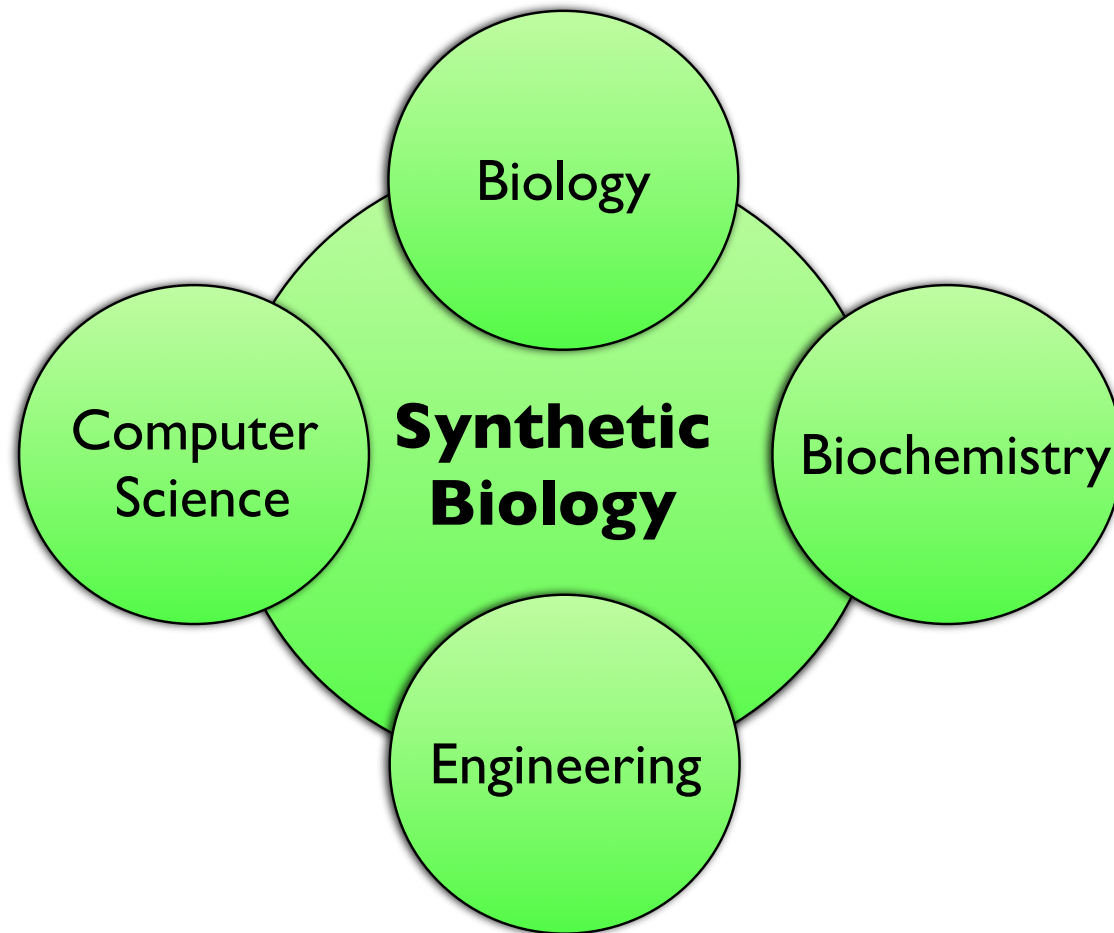
# Biofuels from Algae



CO<sub>2</sub>-neutral

1,000,000 gallons in 2008

# Synthetic Biology



# Synthetic Biology



# Synthetic Biology




 Ribosome Binding Sites ?

 Protein Coding ?

 Regulatory ?

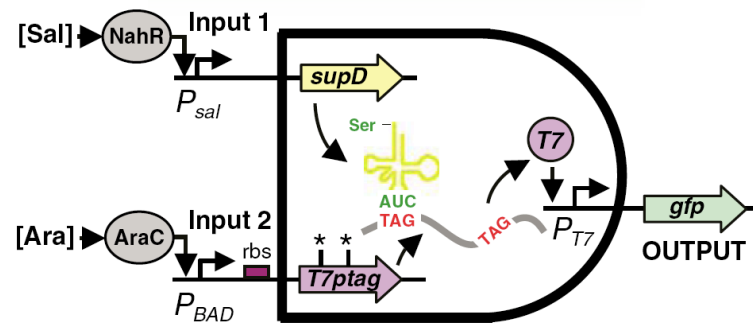
 Terminators ?

 RNA ?

 Conjugation ?

 DNA ?

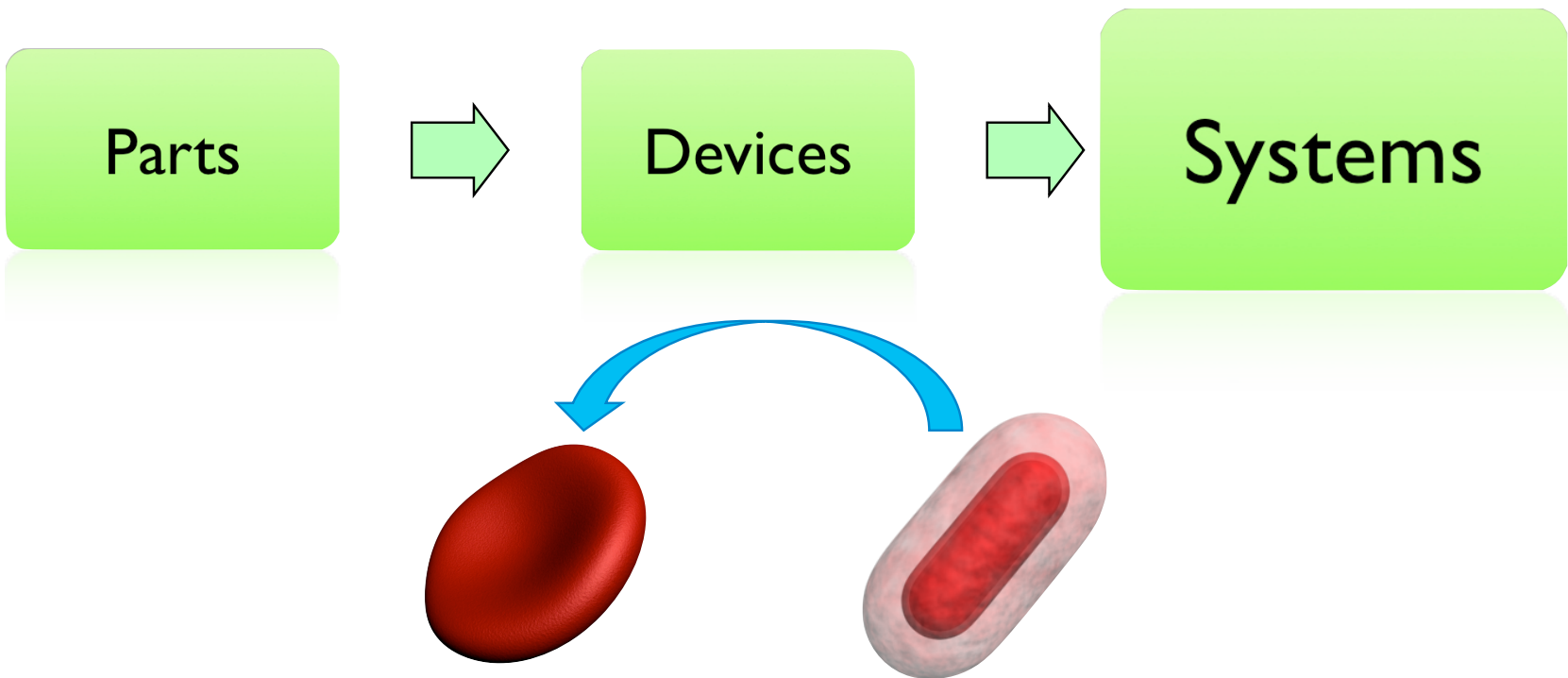
# Synthetic Biology



Anderson et al. *Mol Sys Bio.* 2007.



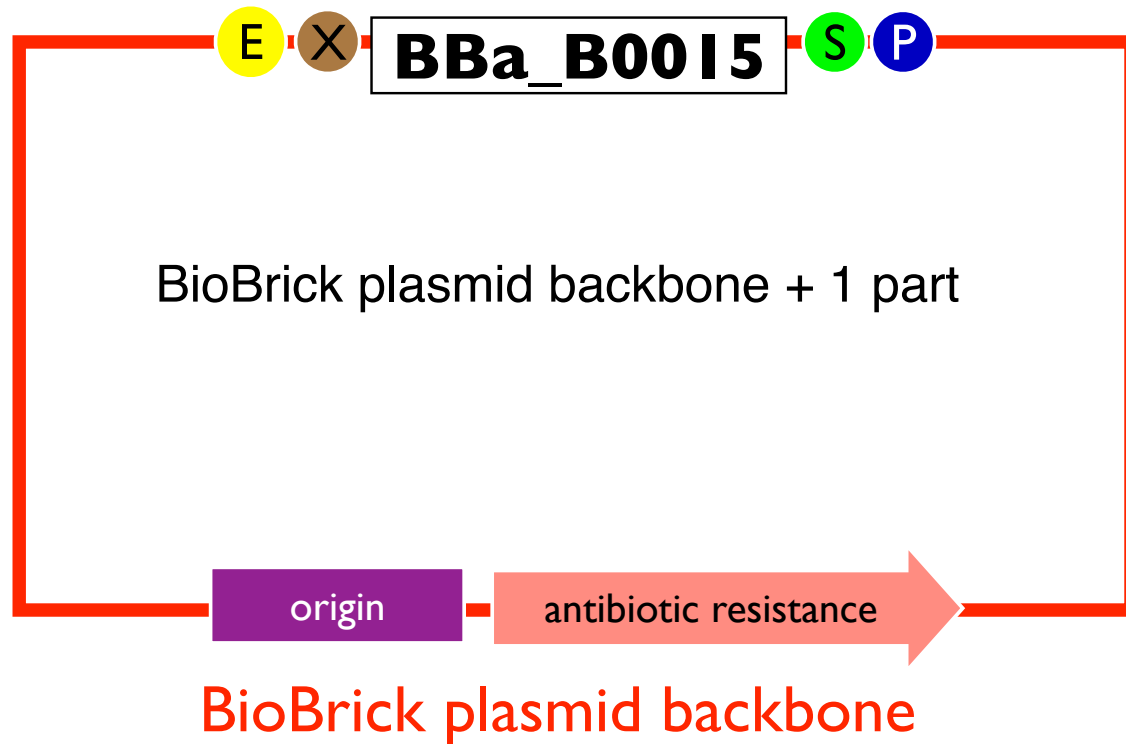
# Synthetic Biology



How do we clone DNA?

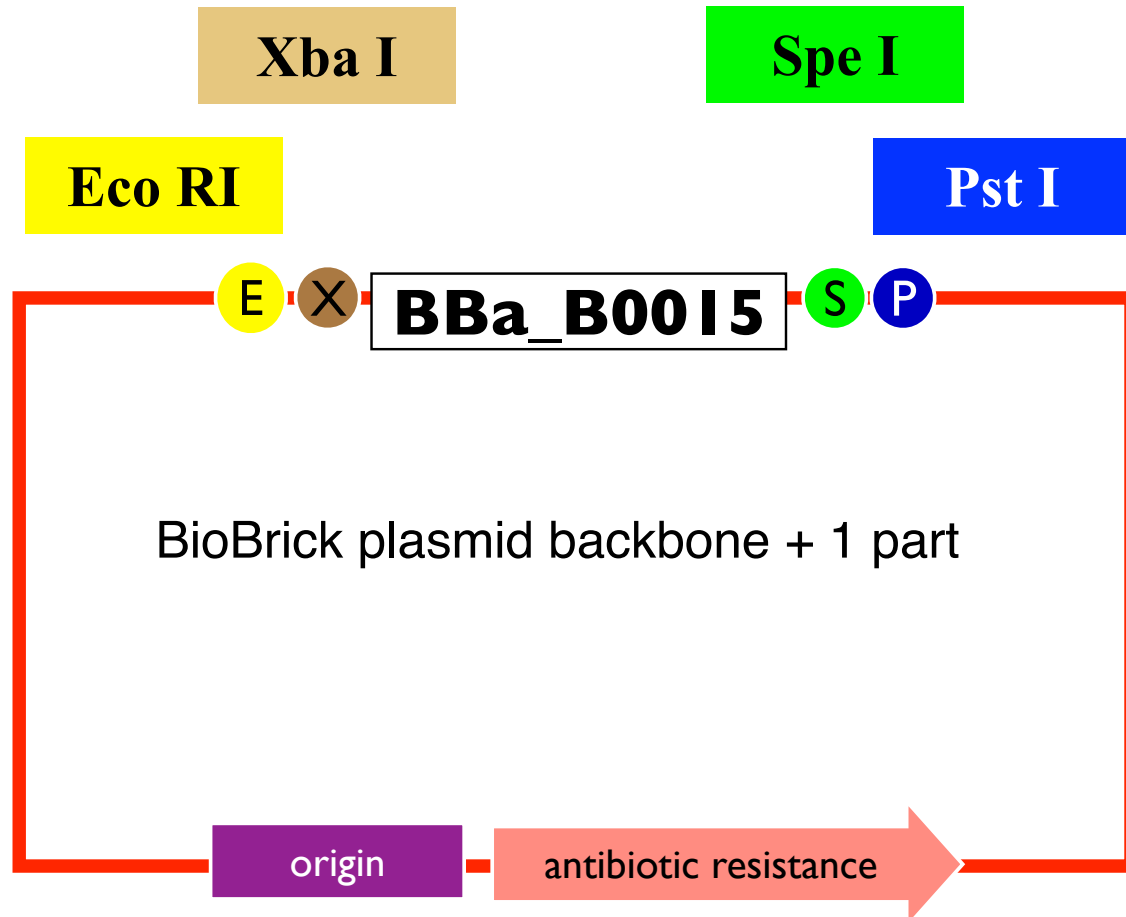
# BioBricks

## BioBrick Part



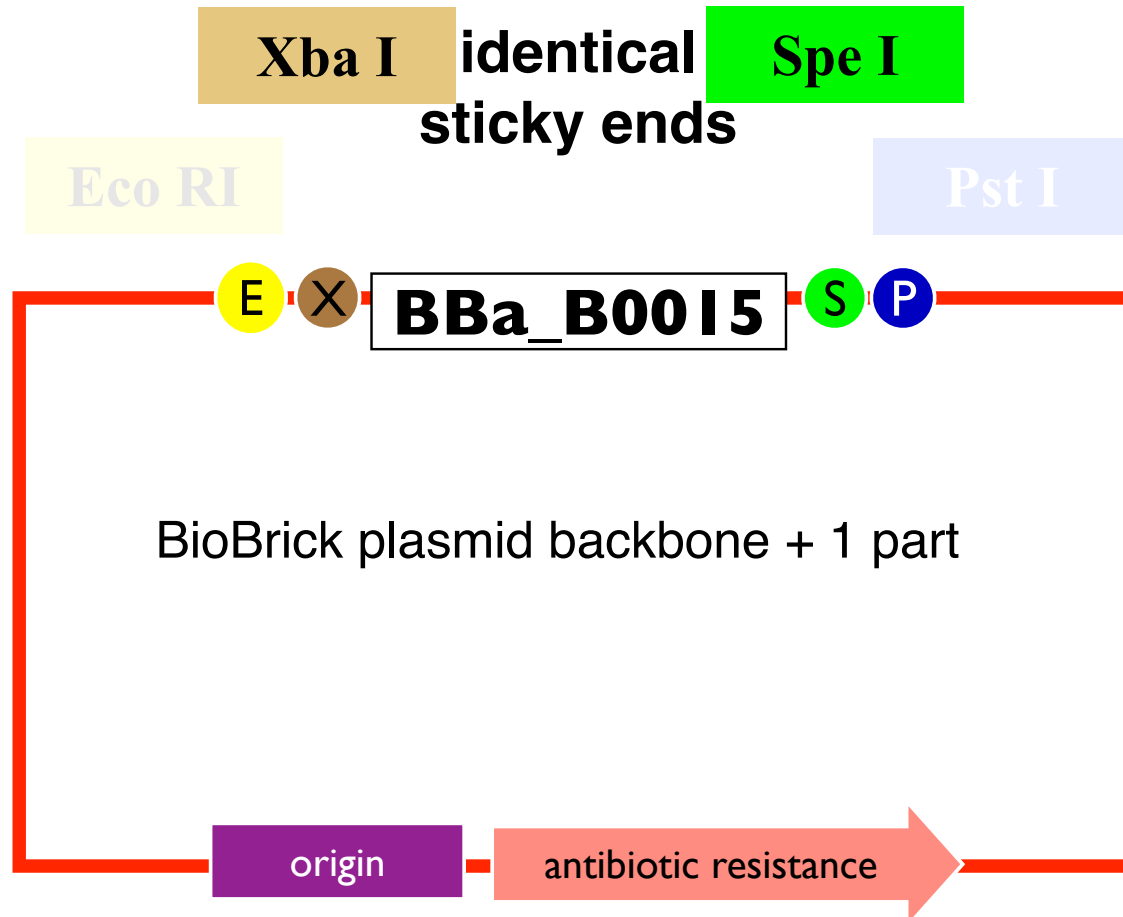
(<http://partsregistry.org/Plasmids>)

# BioBricks



(<http://partsregistry.org/Plasmids>)

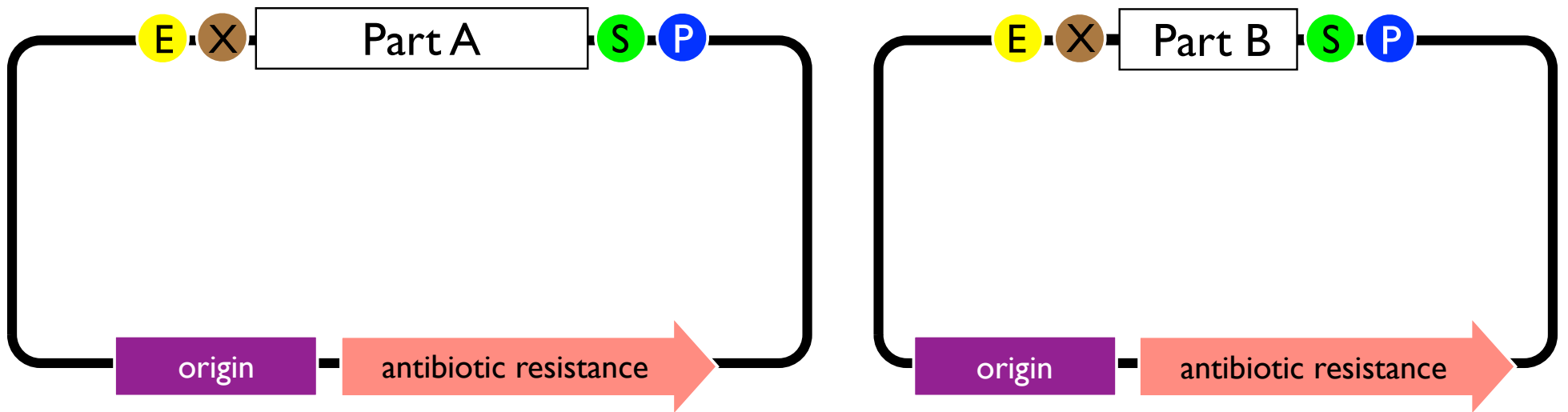
# BioBricks



(<http://partsregistry.org/Plasmids>)

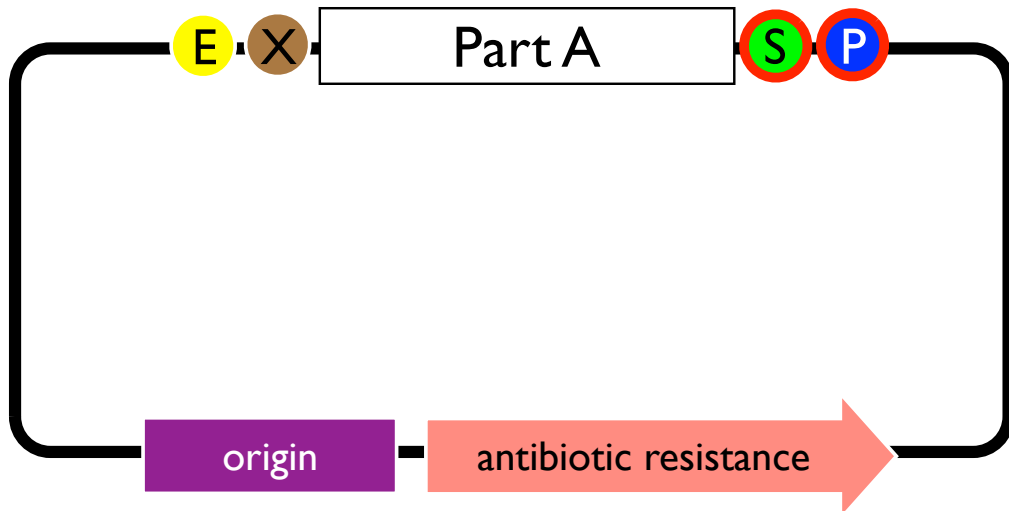
# BioBricks

put B downstream of A

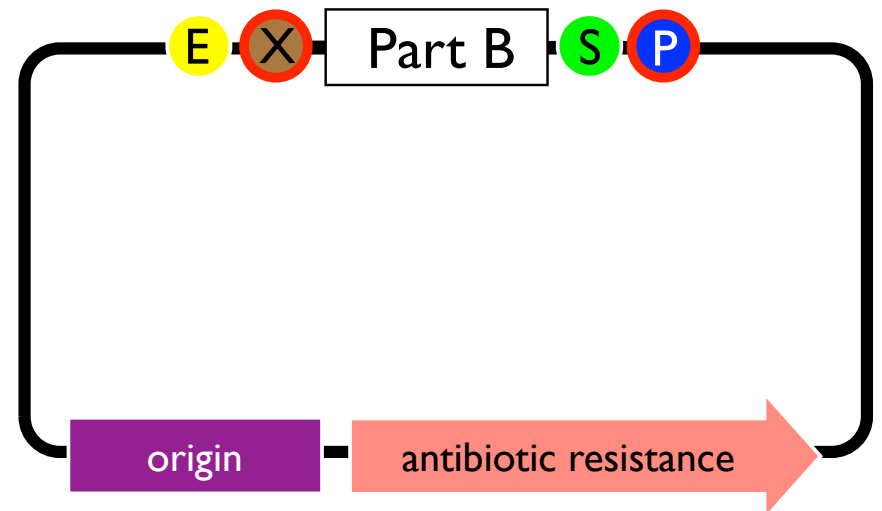


# BioBricks

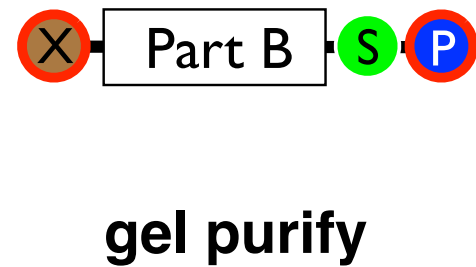
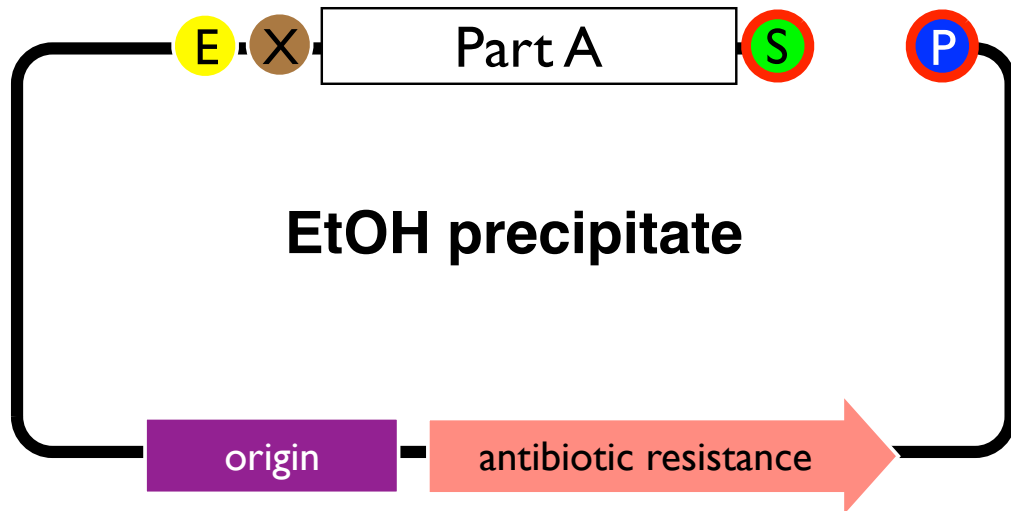
cut with **Spe** and **Pst**



cut with **Xba** and **Pst**

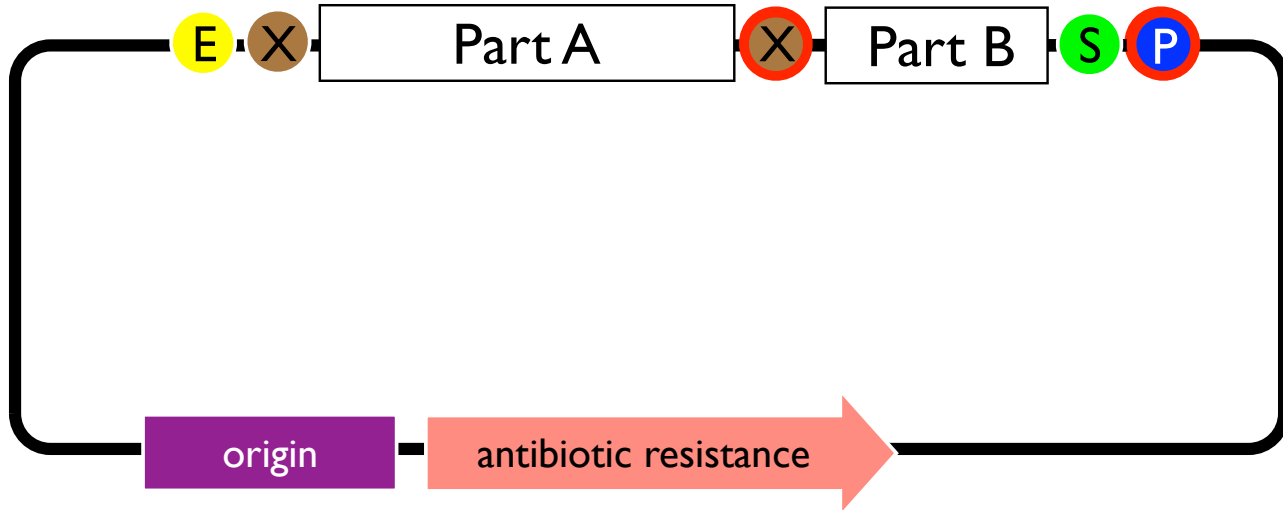


# BioBricks





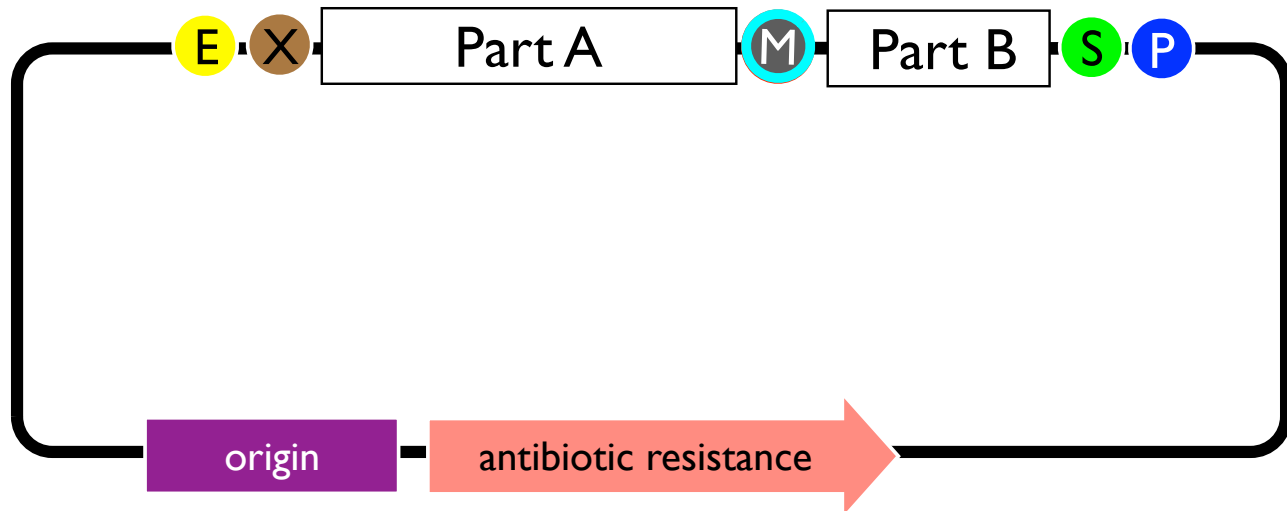
# BioBricks



ligate

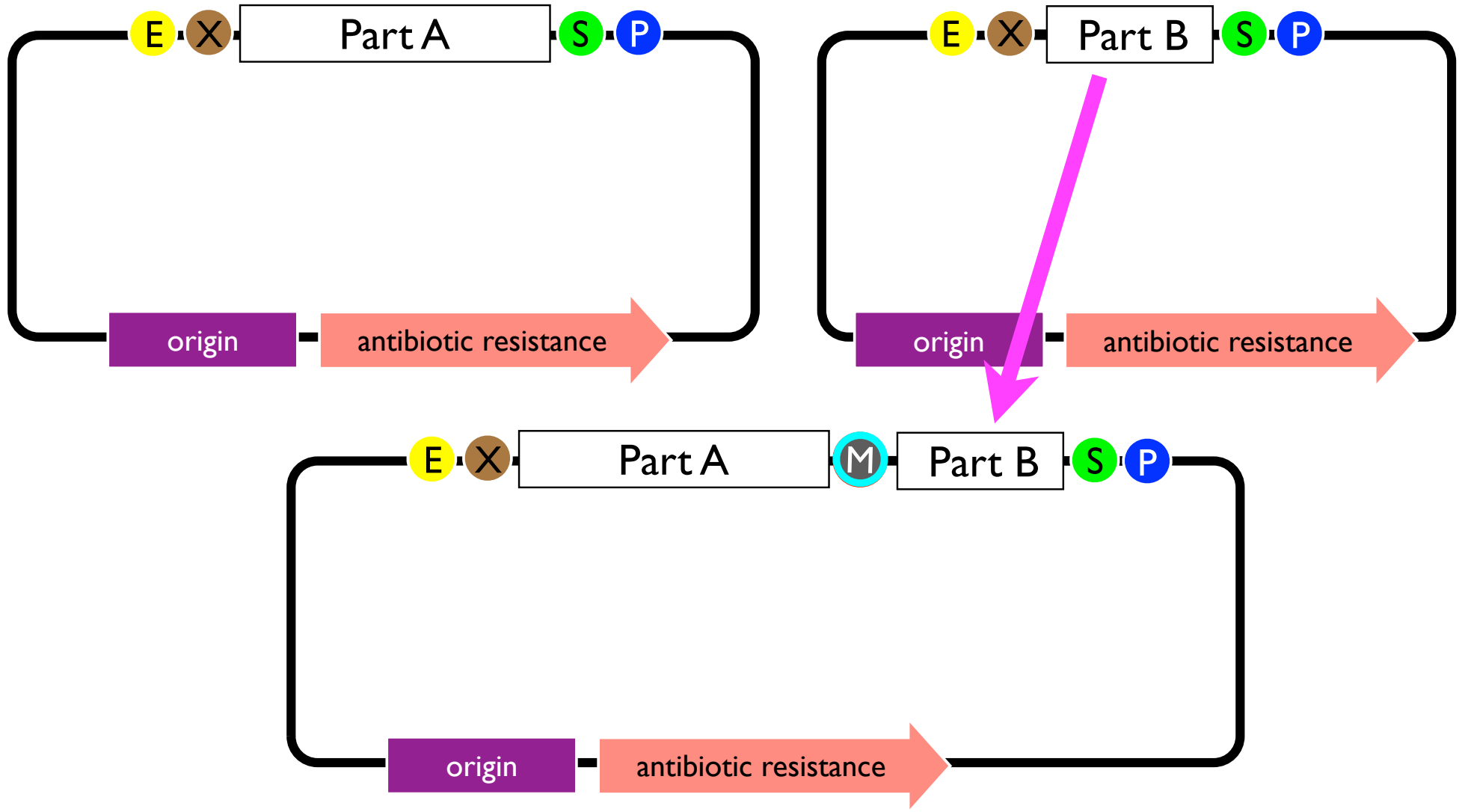
# BioBricks

mixed site = scar



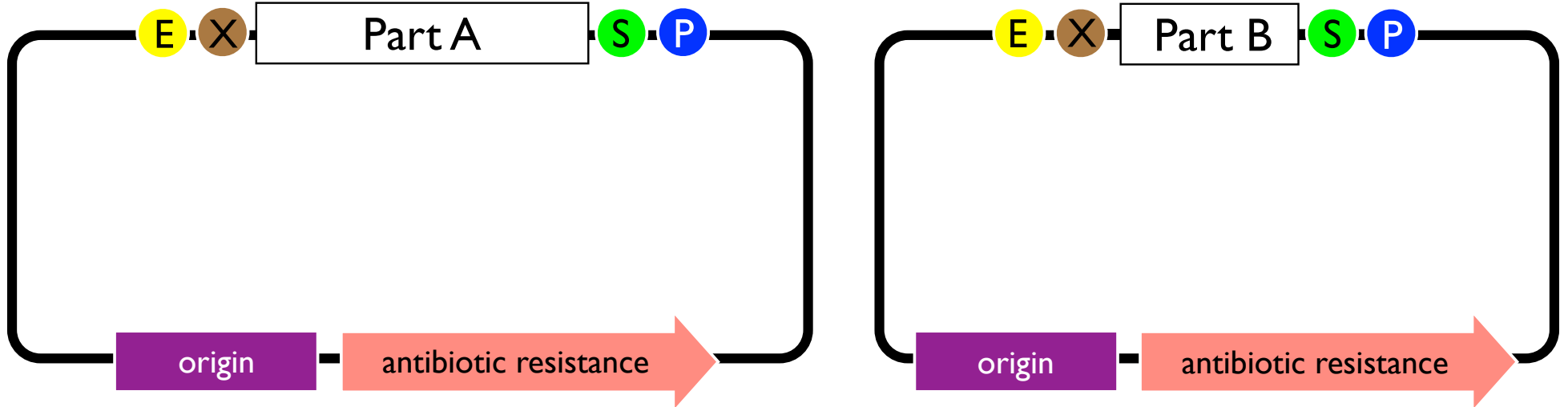
transform

# BioBricks



# Challenge:

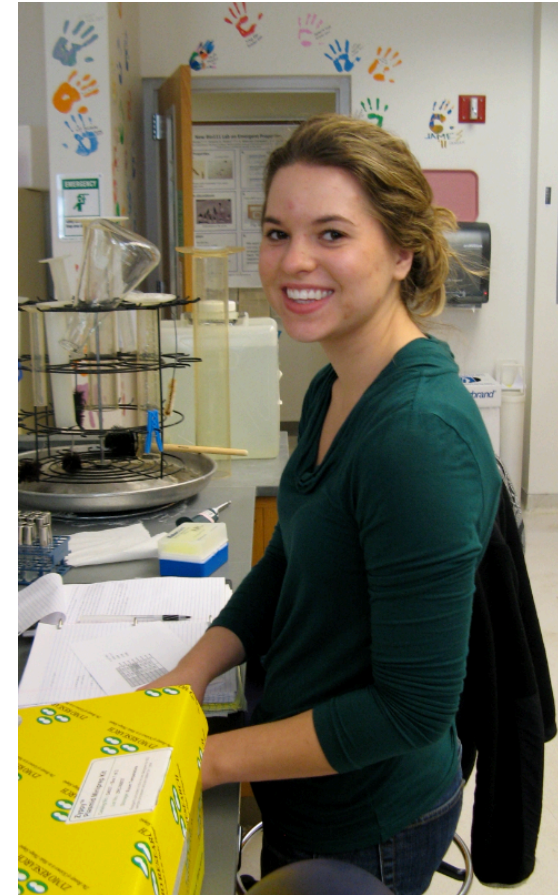
put A upstream of B



# Too Many Projects to Talk About Today



Catherine Doyle '14



Julia Fearrington '13

# Building Bacterial Computers



# Advantages of Bacterial Computation

Software → Hardware → Computation



Computation



Computation

<http://www.dnamnd.med.usyd.edu.au/>

<http://www.turbosquid.com>

# Advantages of Biological Computers

go anywhere - arctic, thermal vents, inside organisms

no electricity

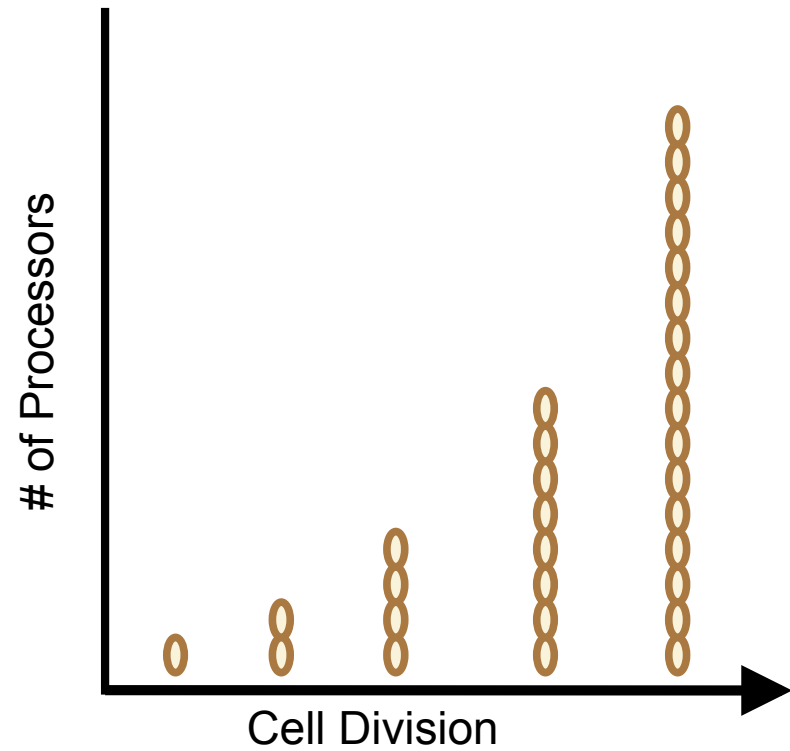
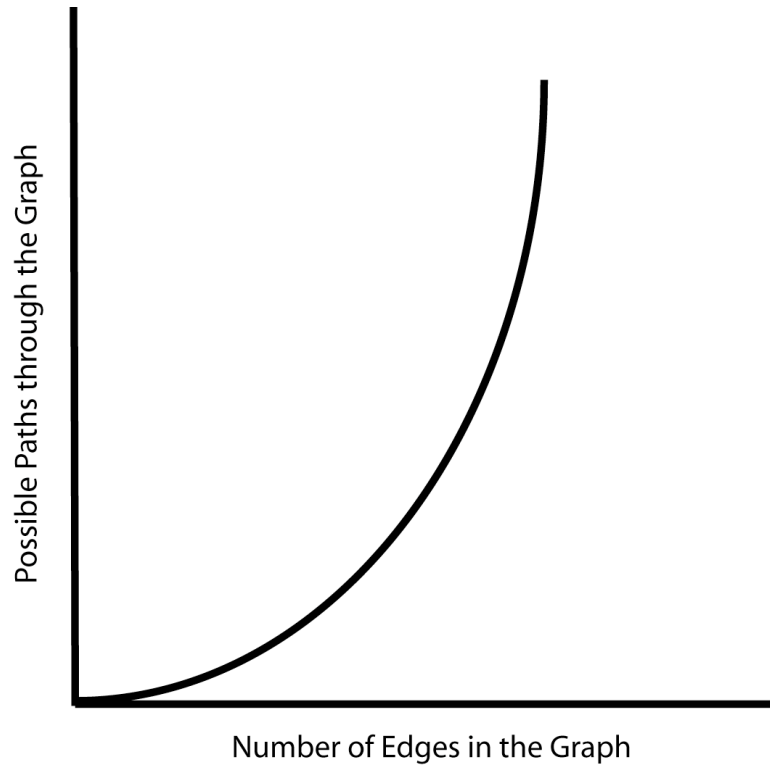
self-replicating

no immune rejection





# Self-replicating Computers

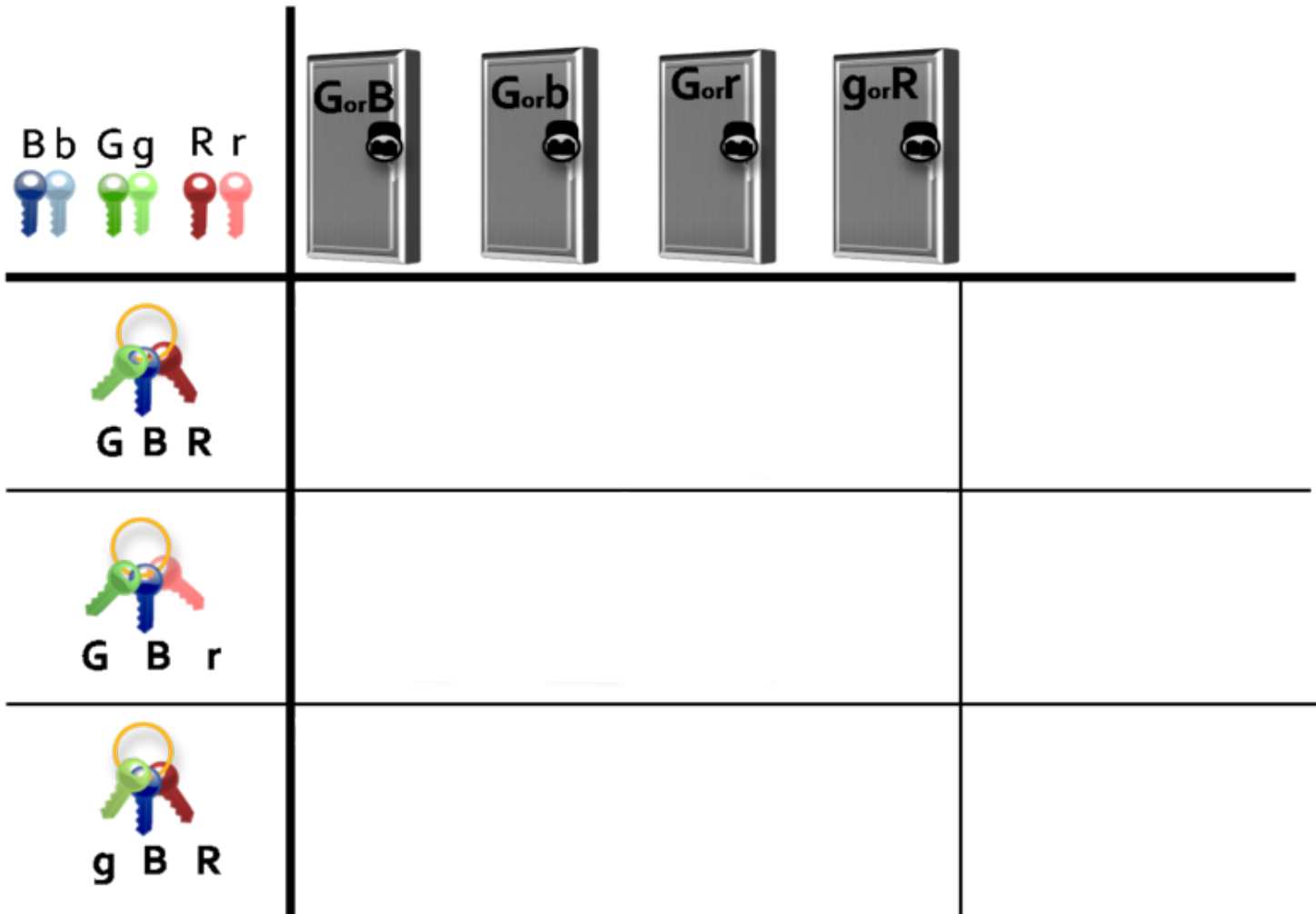


# **One Research Project - the SAT problem**




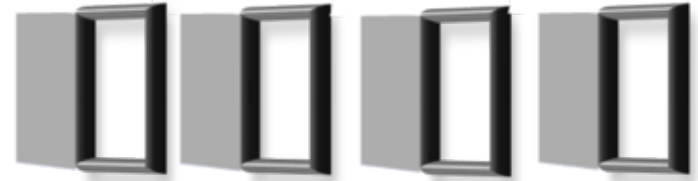

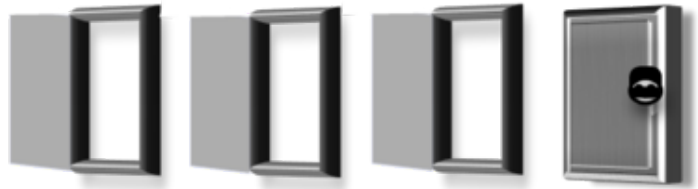


# Define the SATisfiability Problem



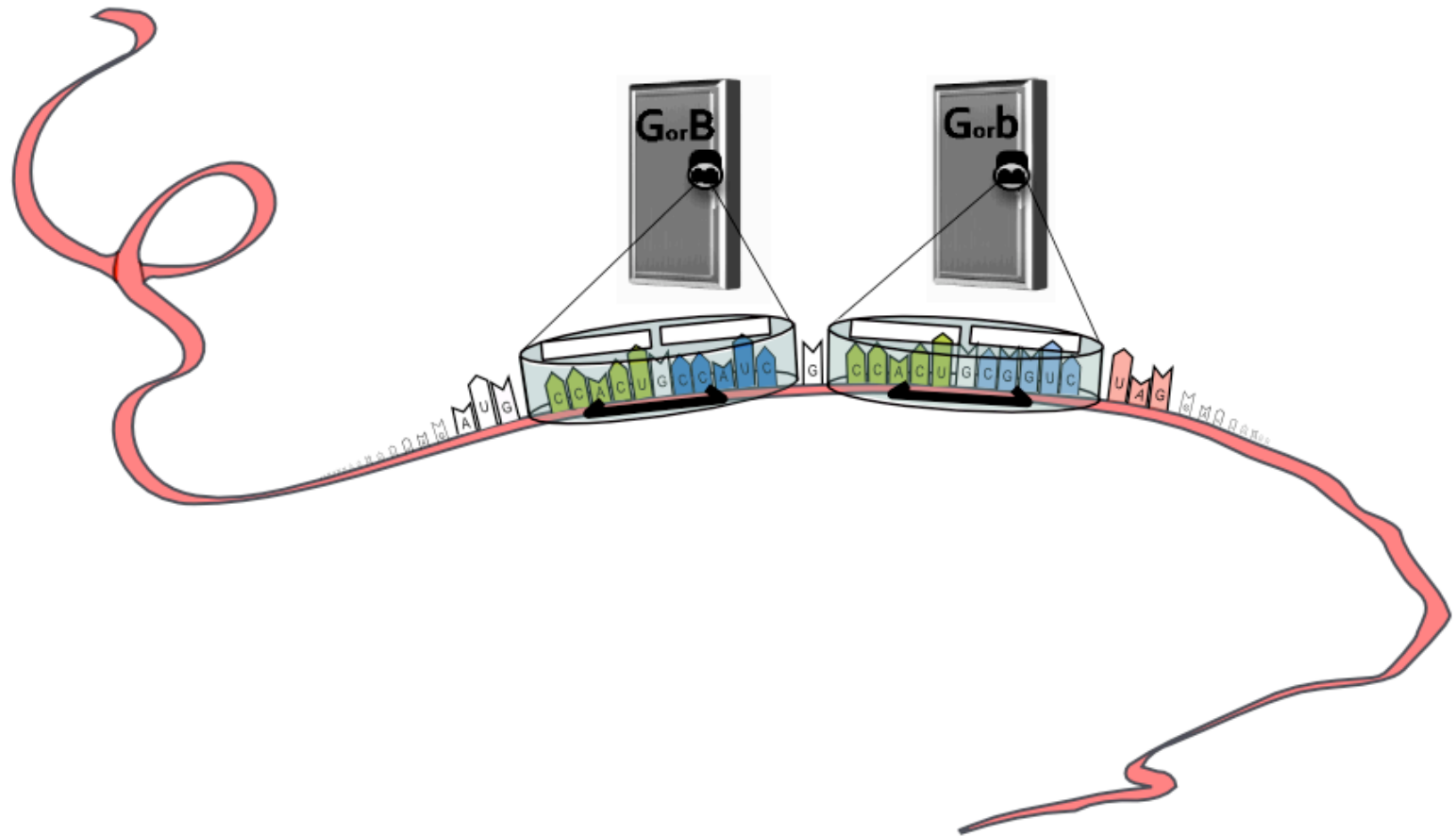
# Define the SATisfiability Problem



# Define the SATisfiability Problem

 <p>Bb Gg Rr</p>	 <p>G<sub>or</sub>B G<sub>orb</sub> G<sub>or</sub>r g<sub>or</sub>R</p>	
 <p>G B R</p>		<b>4</b> open doors
 <p>G B r</p>		<b>3</b> open doors
 <p>g B R</p>		<b>2</b> open doors

# Converting Math to Biology



# Central Dogma

DNA  
atgccctactcactacctatagcgcat



transcription

mRNA  
**aug** ccc uac uca cua ccu aua ccg cau

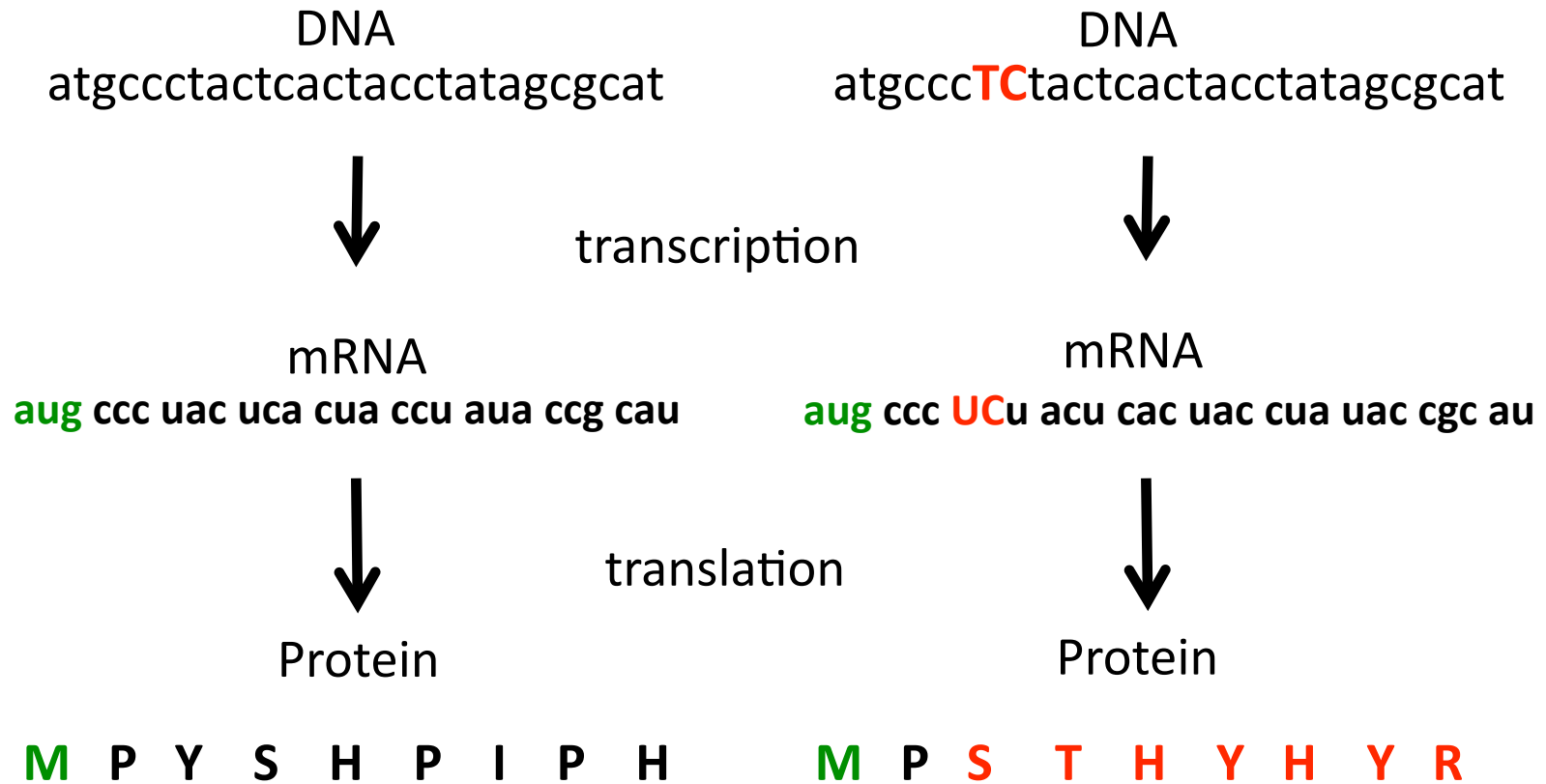


translation

Protein

**M** P Y S H P I P H

# Frameshift Mutation





# Frameshift Suppression

DNA  
atgccctactcactacctatagcgc



mRNA  
aug ccc uac uca cua ccu aua ccg cau



Protein  
M P Y S H P I P H

DNA  
atgcc**TC**tactcactacctatagcgc



5 base  
suppressor tRNA  
aug **cccUC** uac uca cua ccu aua ccg cau

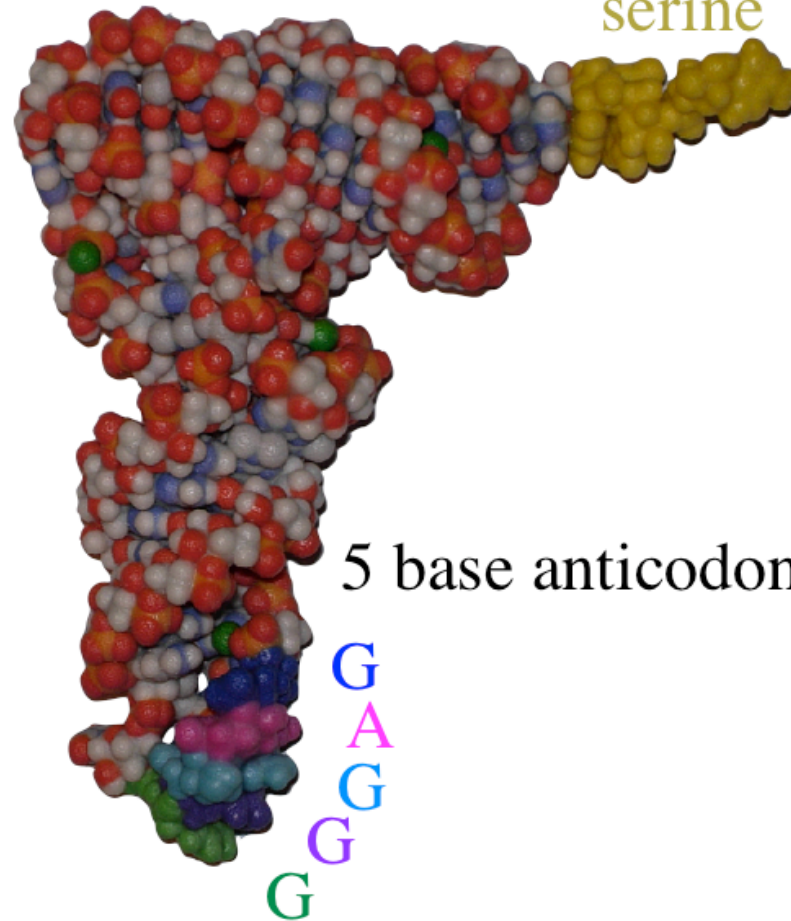


Protein  
M **S** Y S H P I P H

# Suppressor tRNA

core tRNA  
nucleotides

serine



# Coding 2-SAT Clause



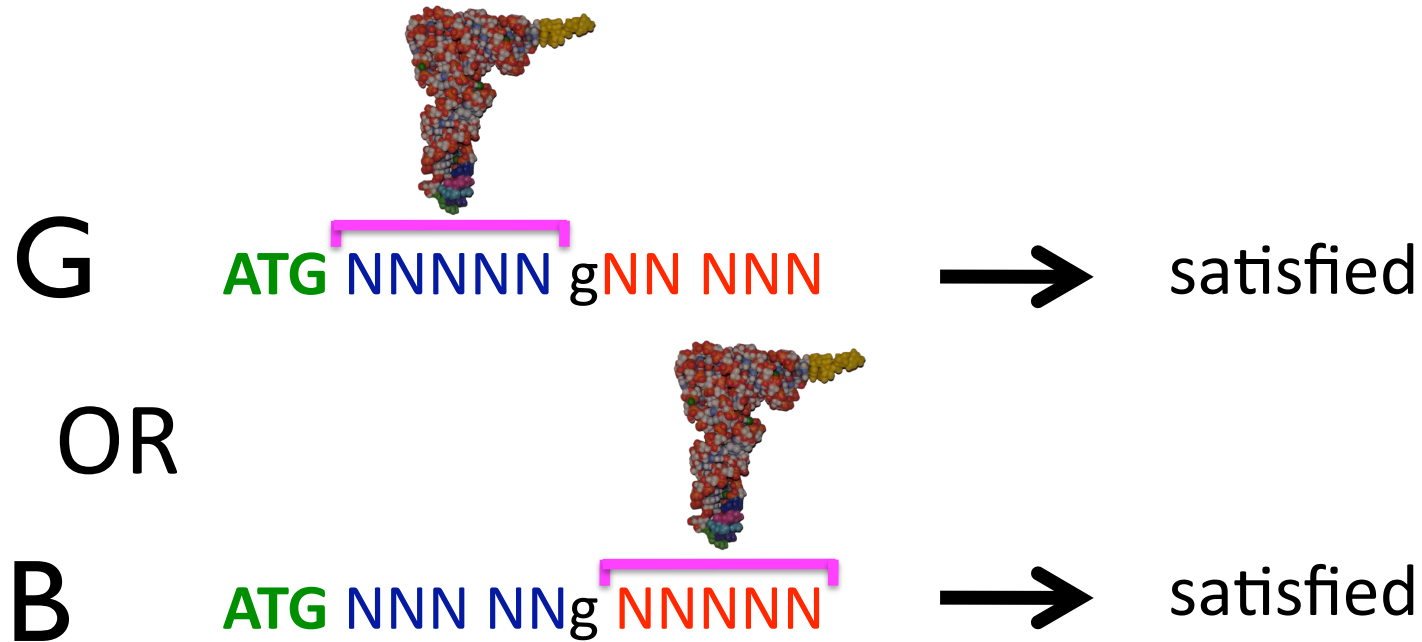
G

ATG NNNNN gNN NNN

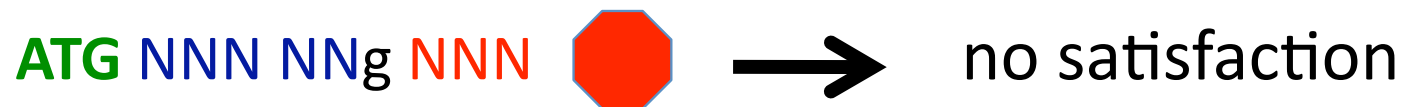
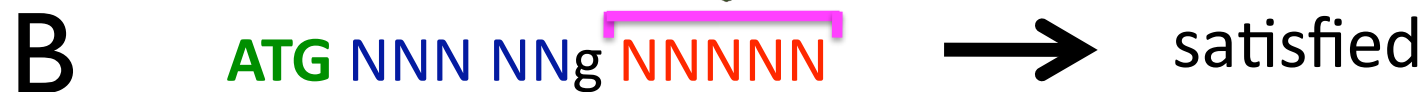
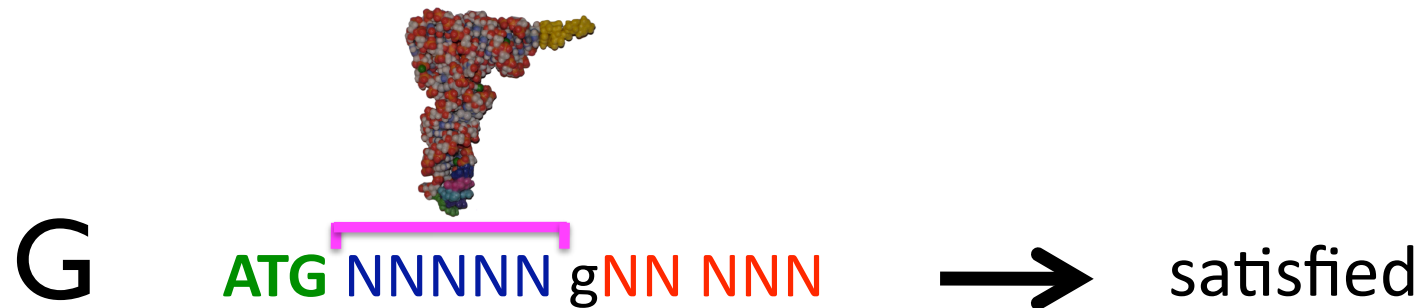


satisfied

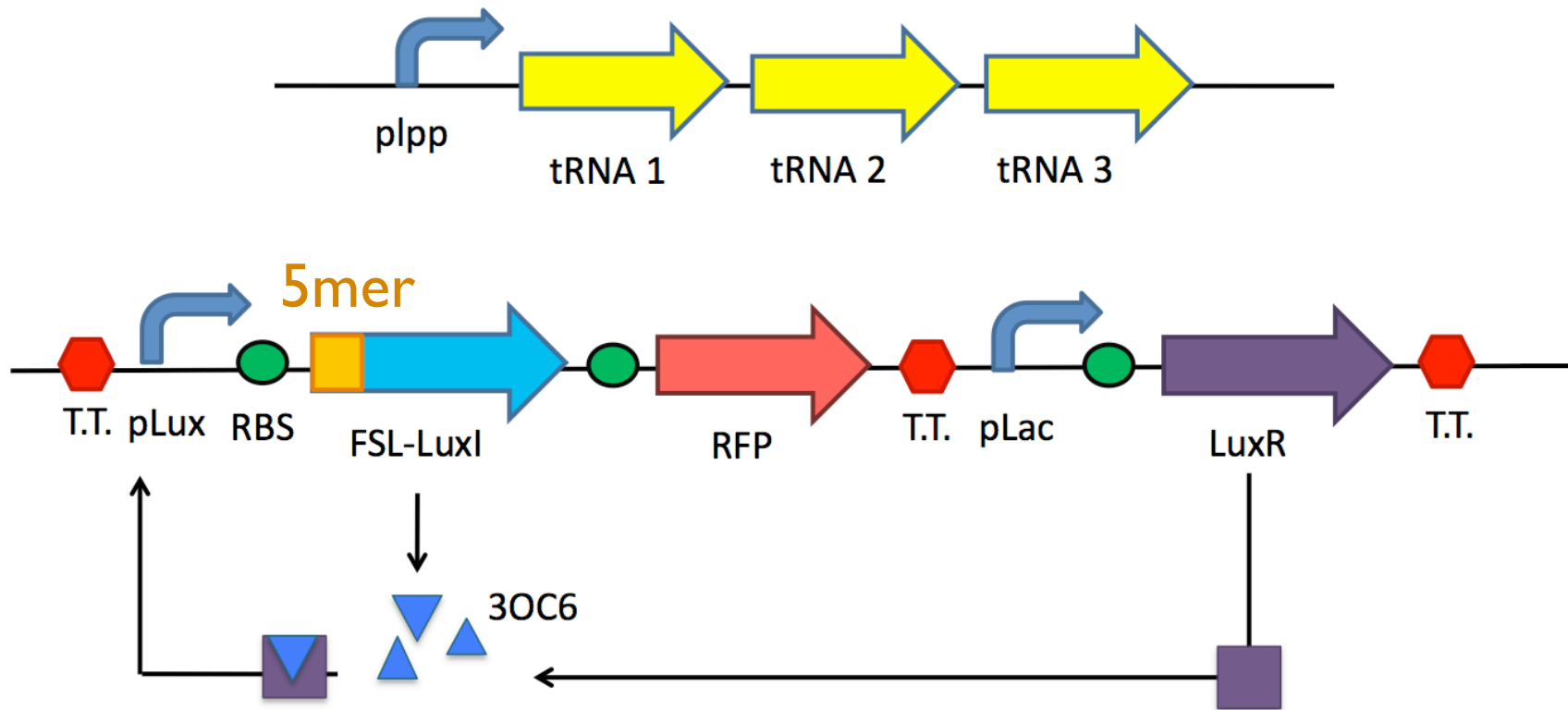
# Coding 2-SAT Clause



# Coding 2-SAT Clause



# Redesign System v2.0



# Outcomes of v 2.0

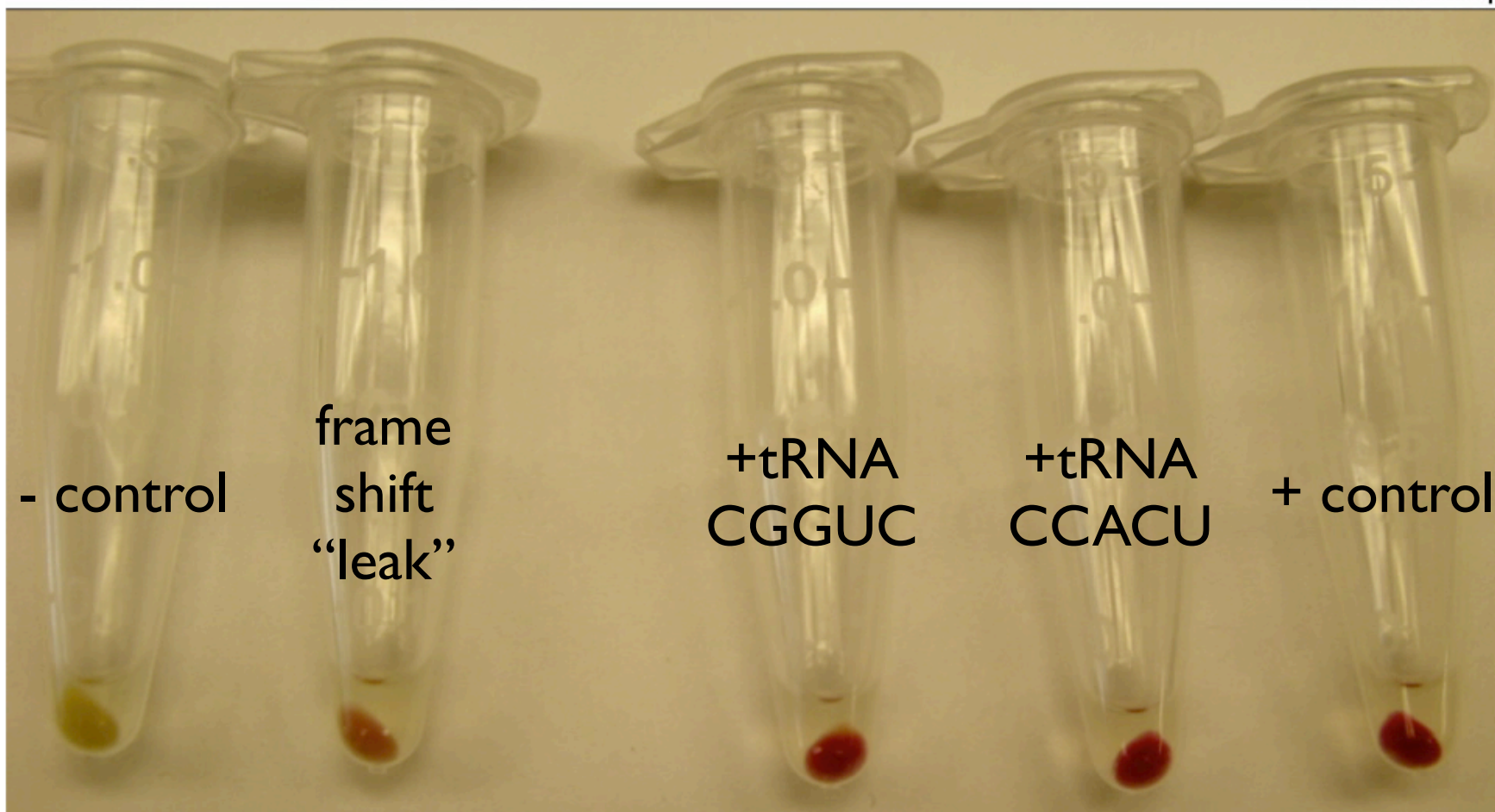
no RFP

5mer CGGUC

5mer CGGUC  
+ tRNA

5mer CCACU  
+ tRNA

optimized  
feedback loop



- control

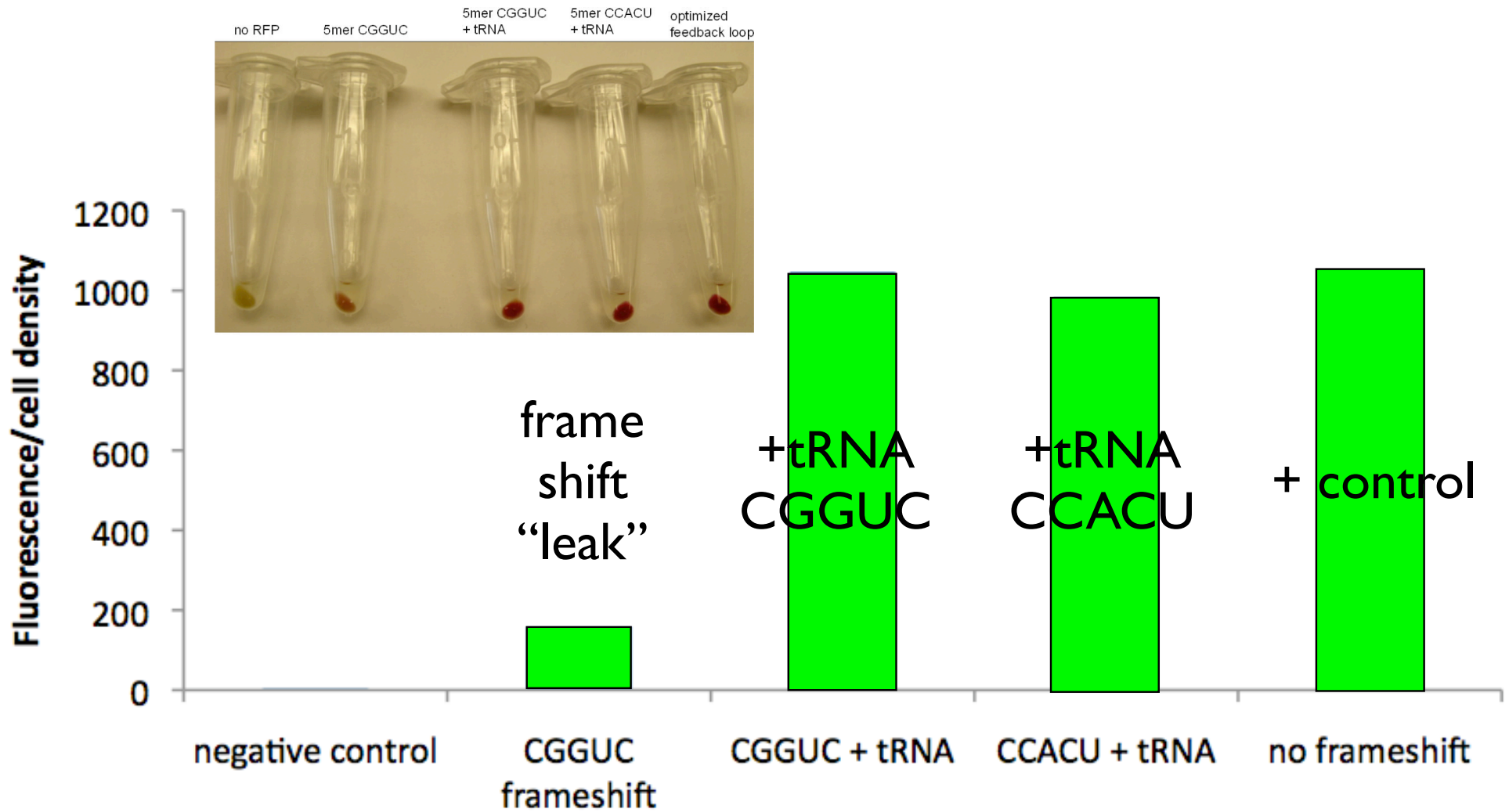
frame  
shift  
"leak"

+tRNA  
CGGUC

+tRNA  
CCACU

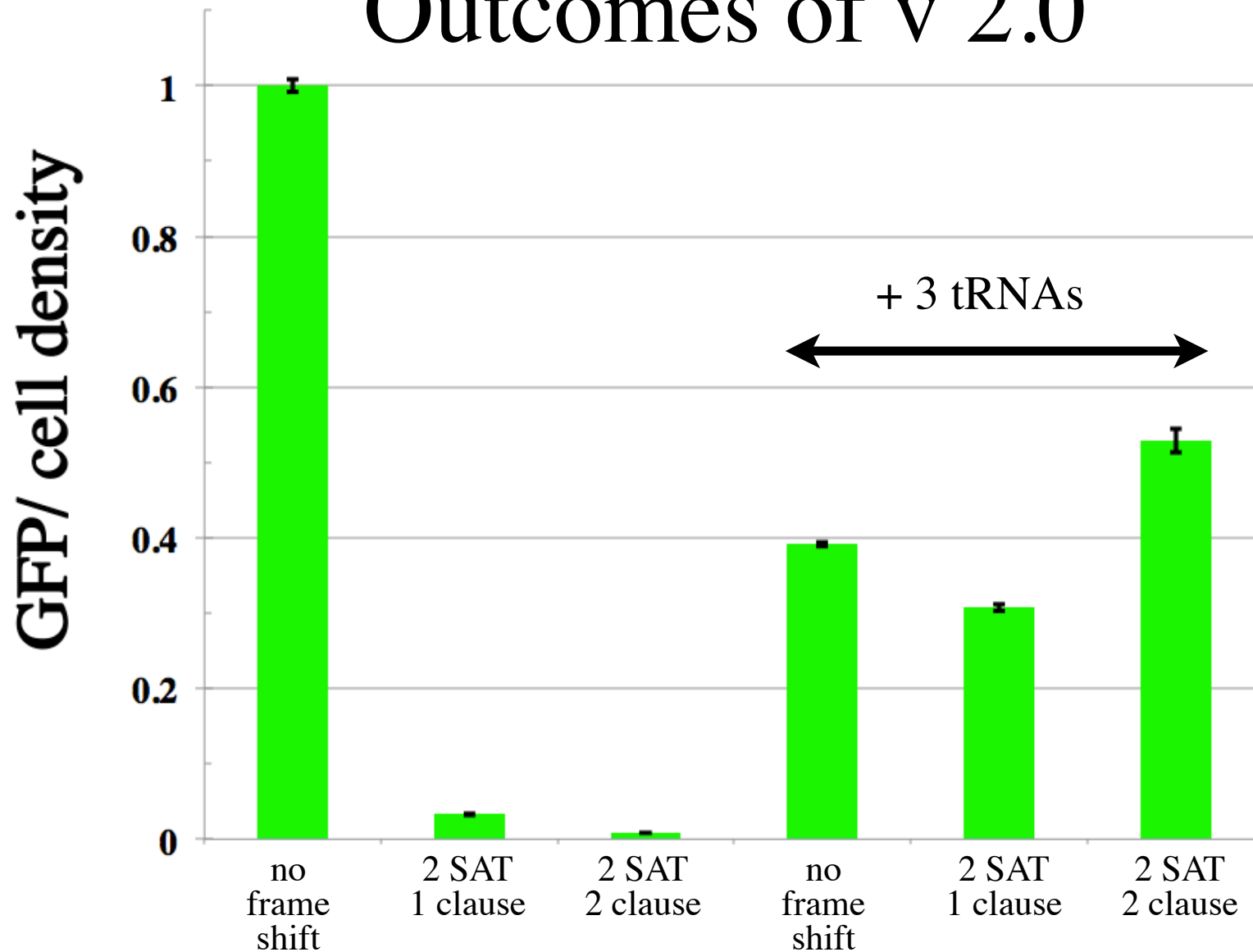
+ control

# Outcomes of v 2.0

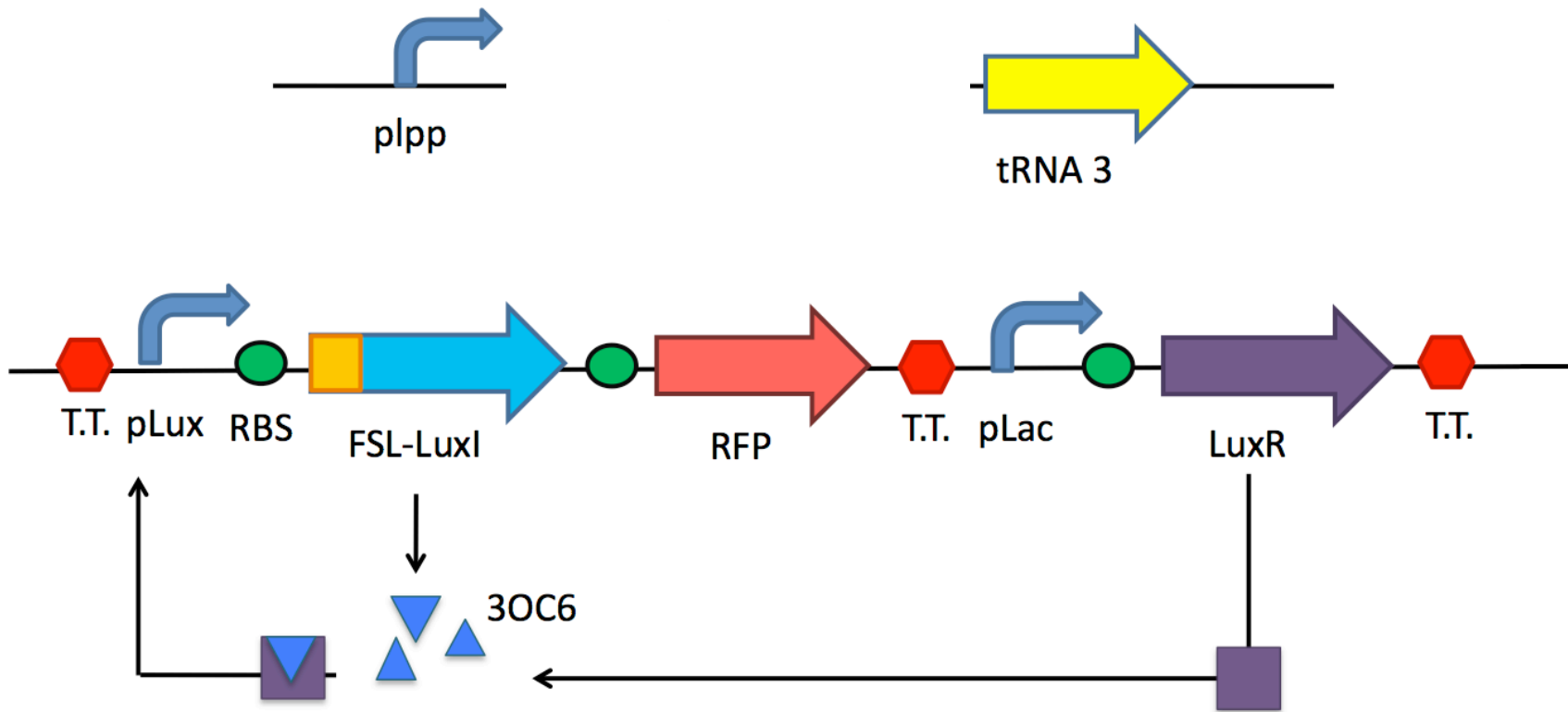




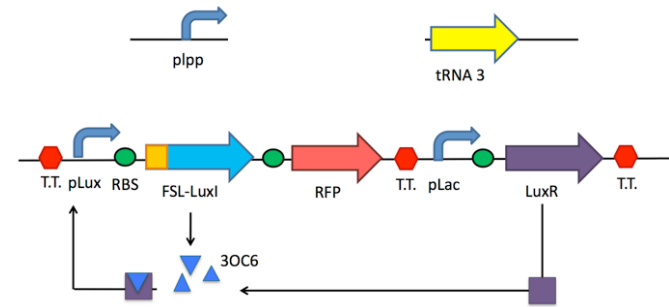
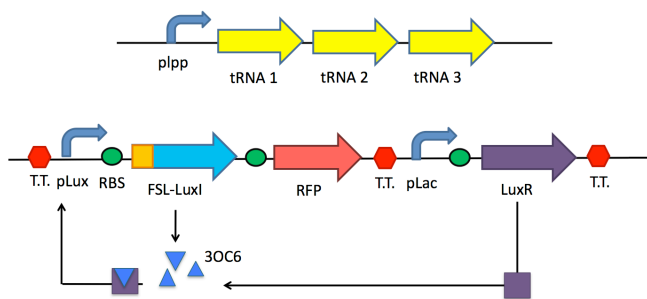
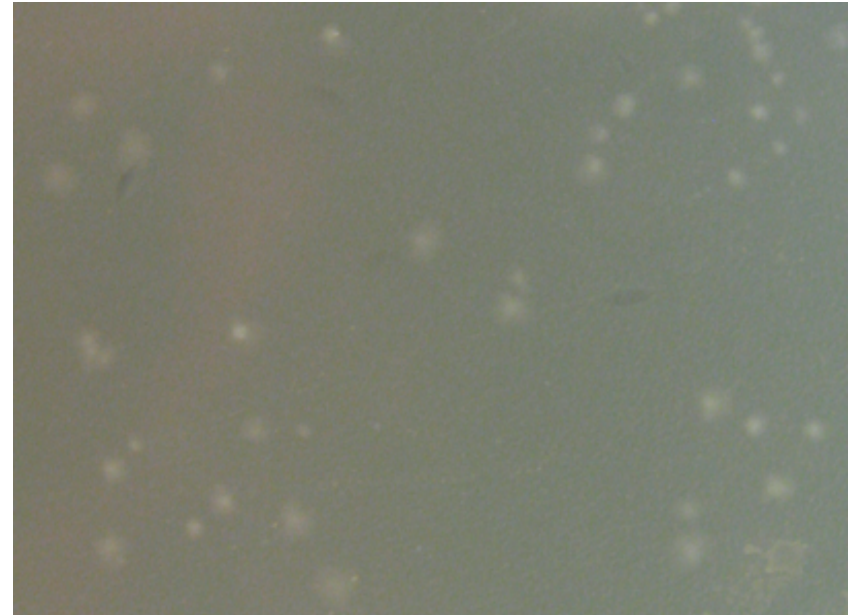
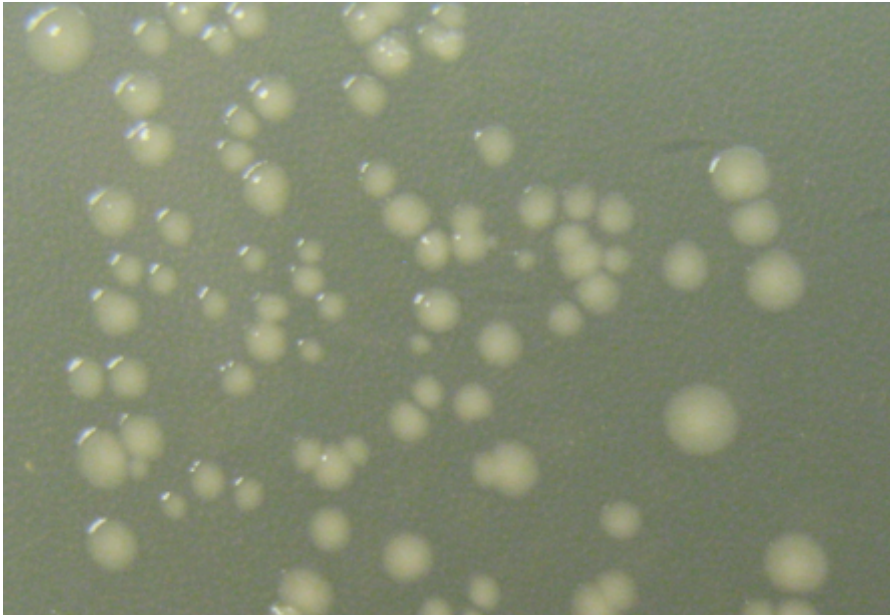
# Outcomes of v 2.0



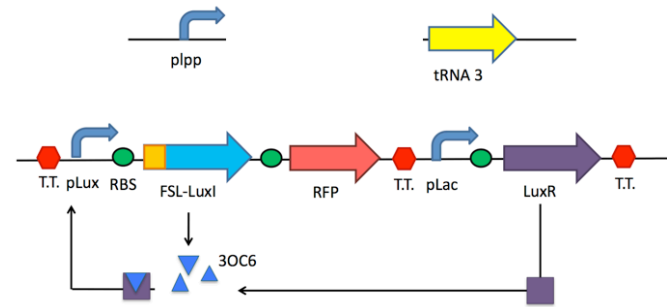
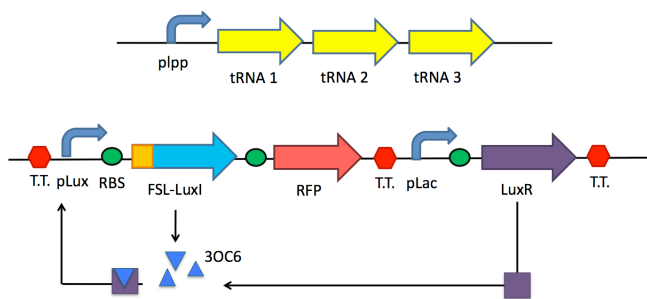
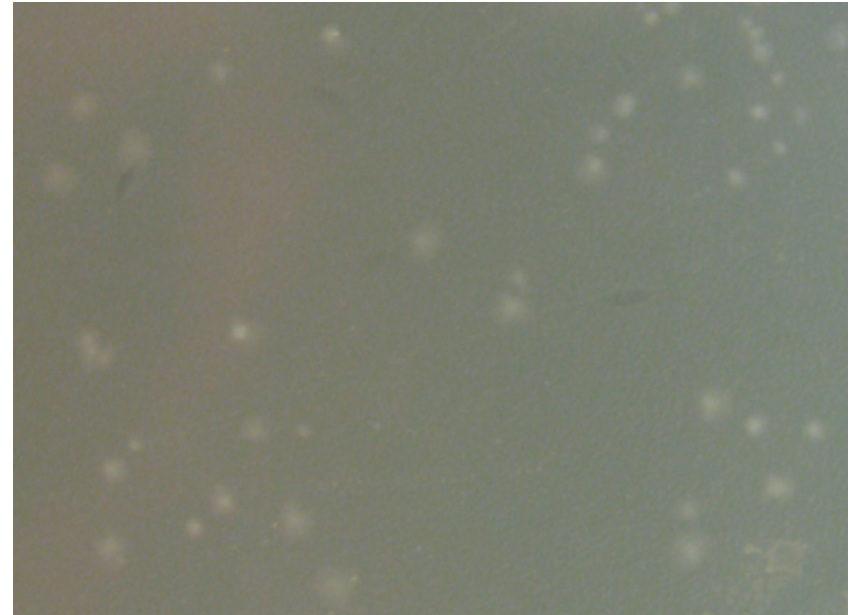
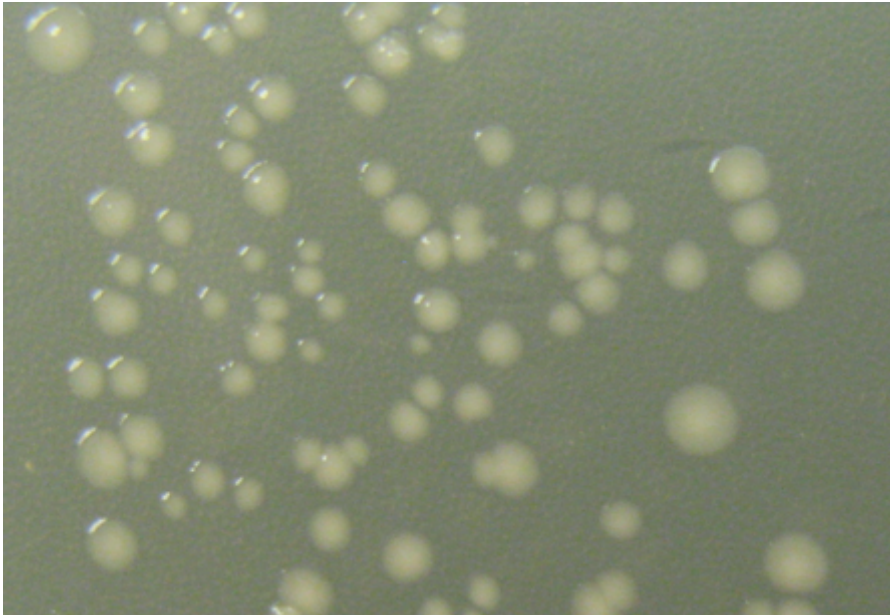
# Controls....



# Controls....

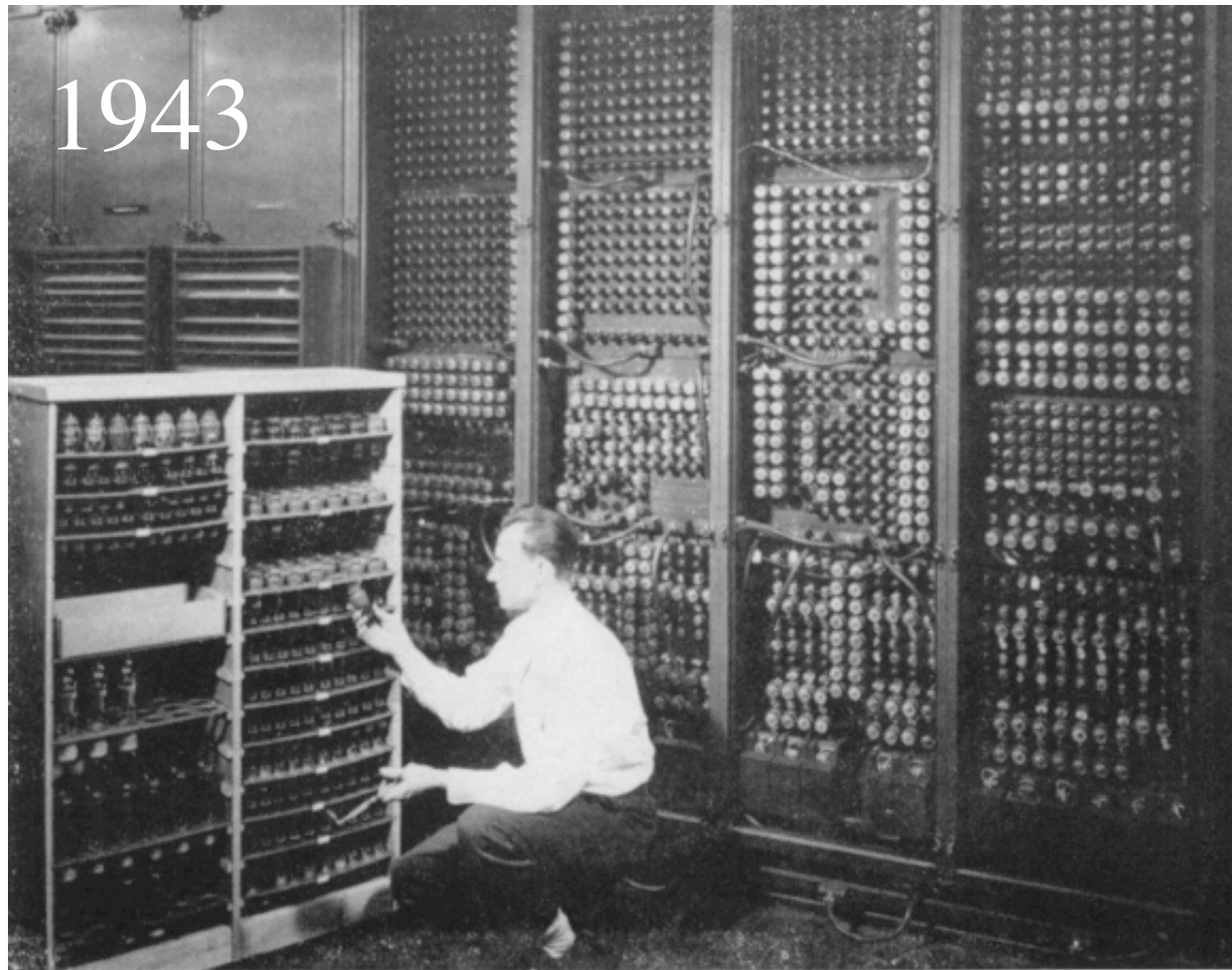


# Still working on this...



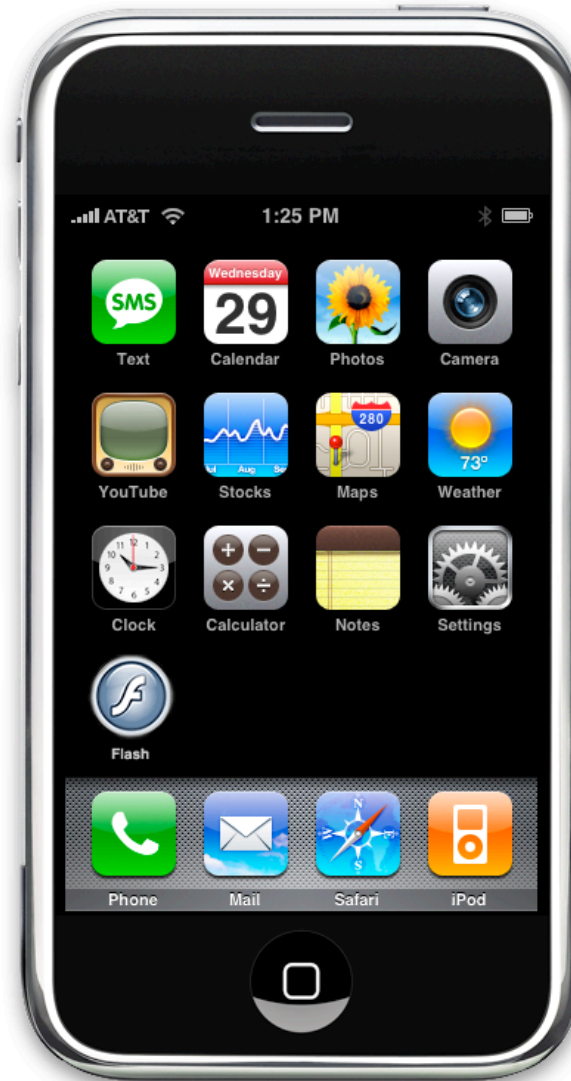
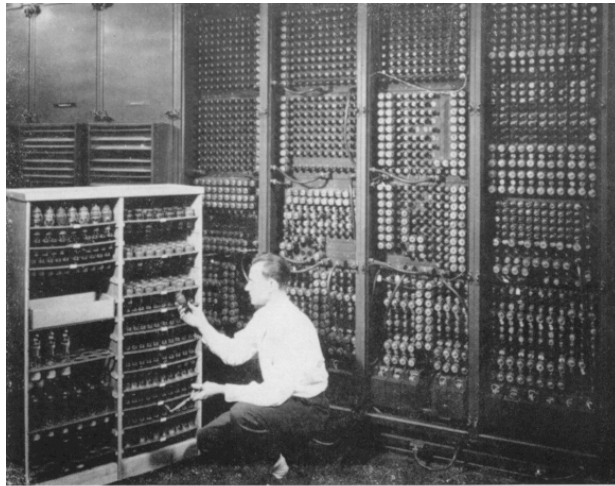
Why build bacterial computers?

# Evolution of Computers



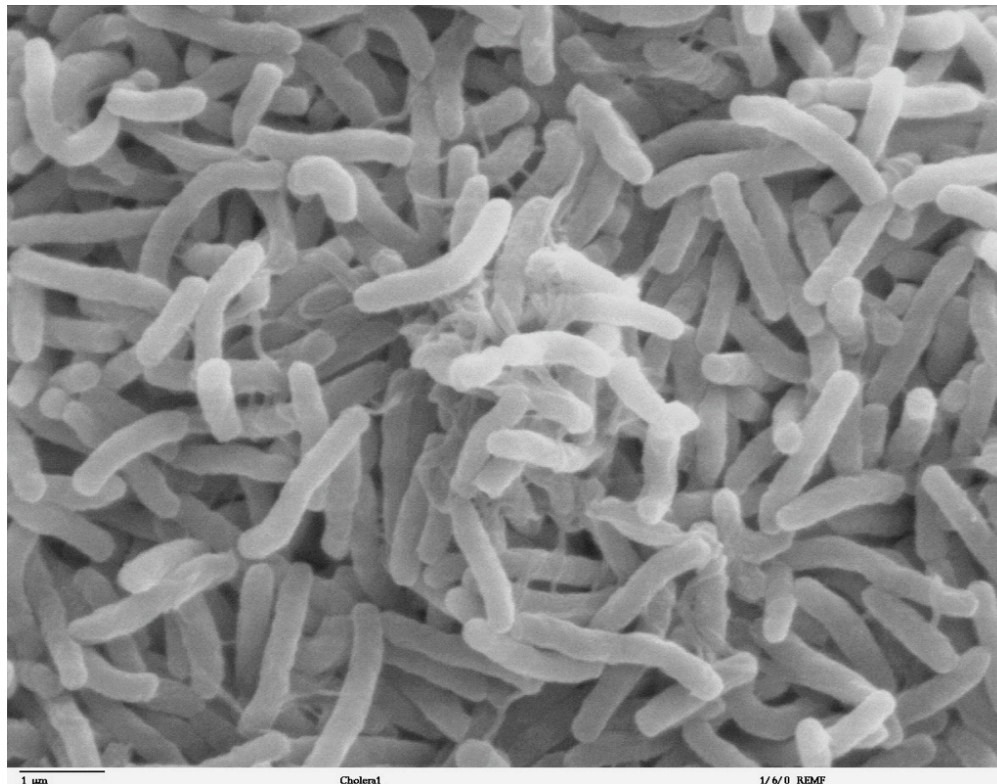
# Evolution of Computers

## iPhone in 2011



# Evolution of Bacterial Computers

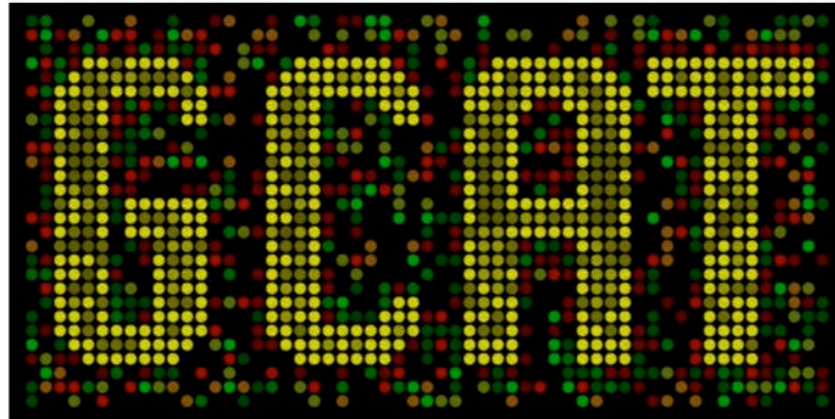
*E. coli* in 2011



Living Hardware  
in 2021



# Intro Bio Student Collaborators



# Intro Bio Student Collaborators



# Intro Bio Student Collaborators

Biologists need is a registry of functional promoters (RFP).

Intro students find promoter of interest.

Oligator converts into oligos.

Clone new promoter.

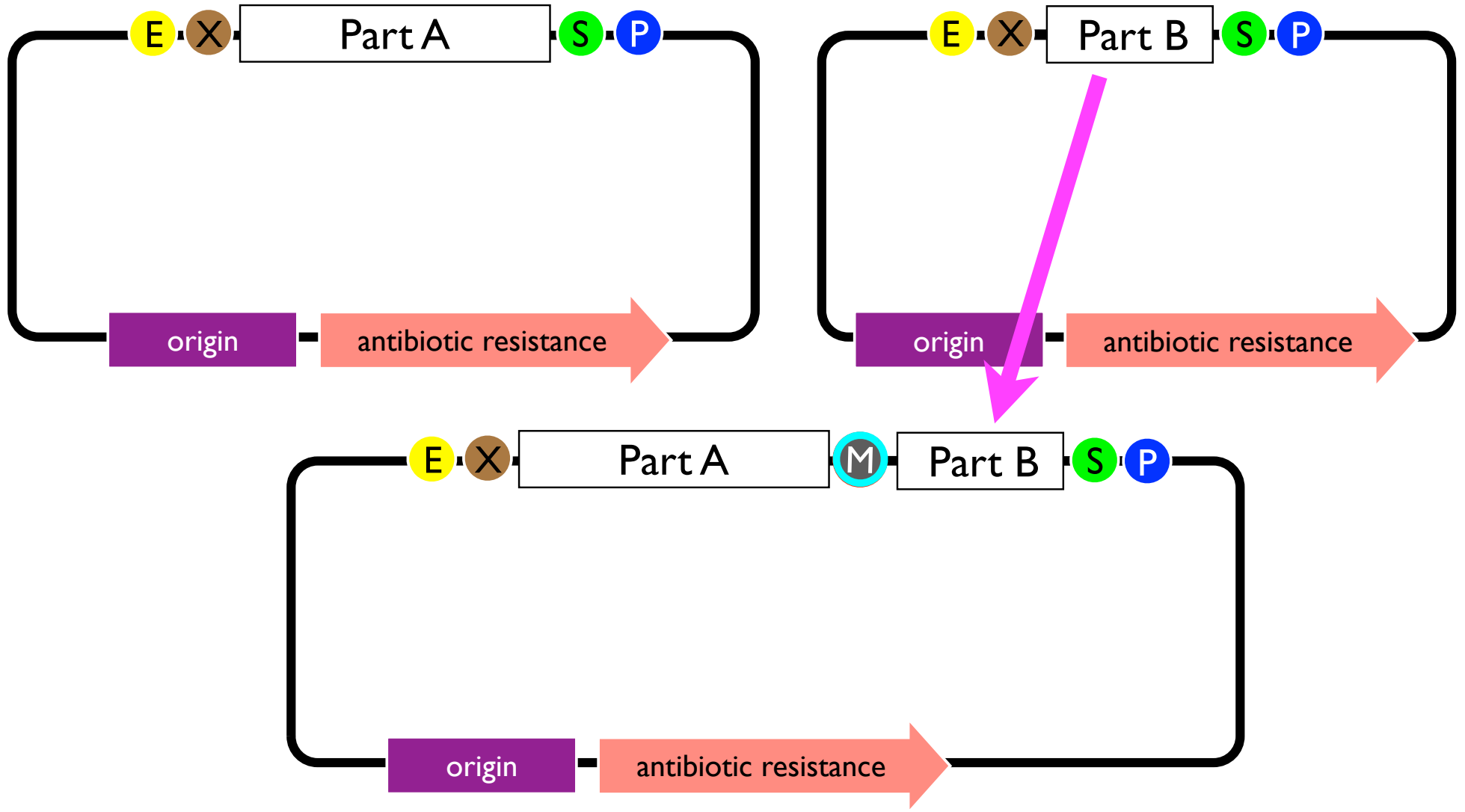
Transform and test new promoter.

Submit to Registry of Functional Promoters (national resource).

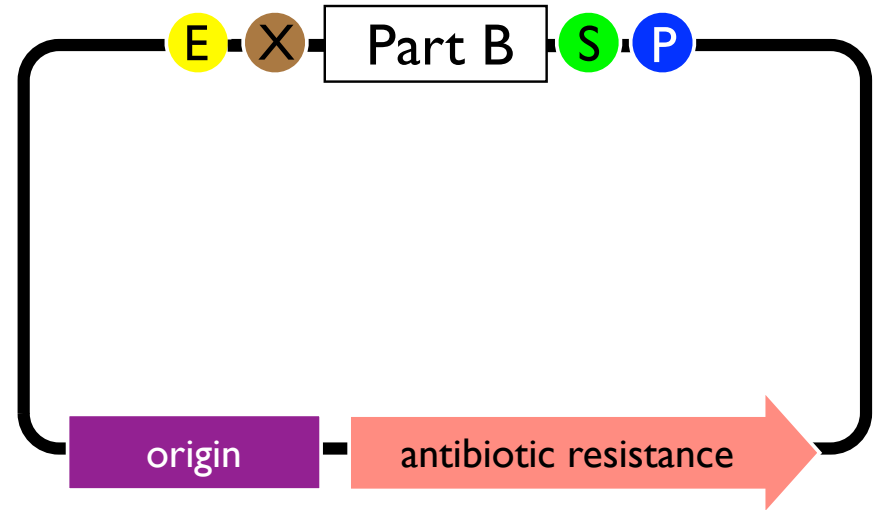
But cloning DNA is not easy....



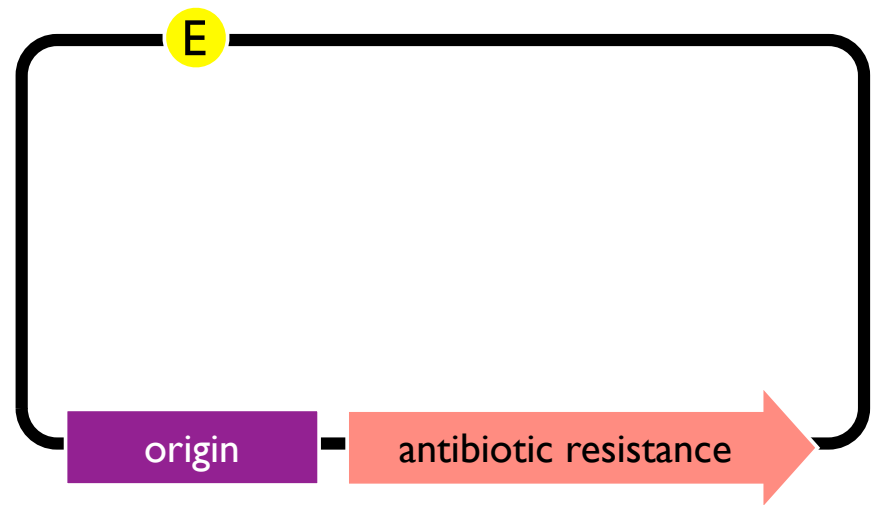
# Details of Cloning



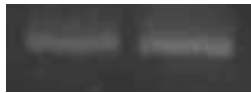
# Gel Purification

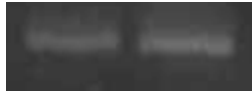


# Gel Purification



# Gel Purification



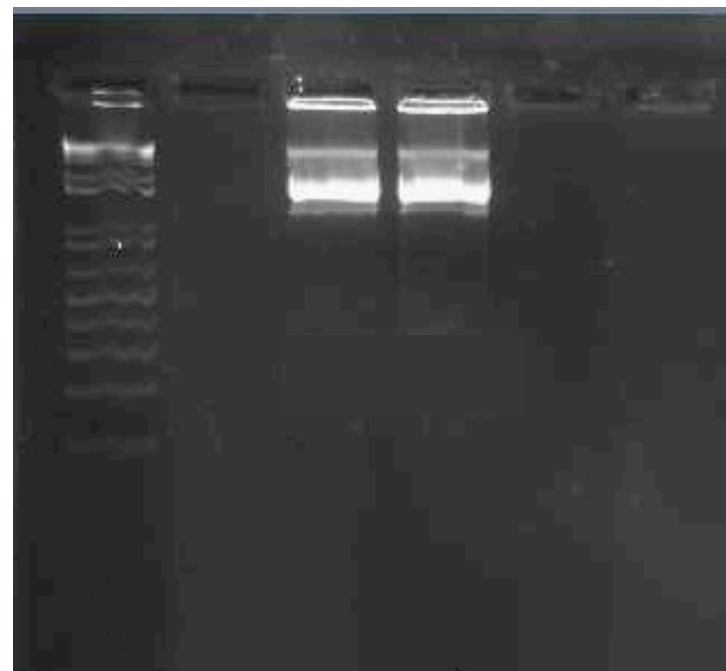
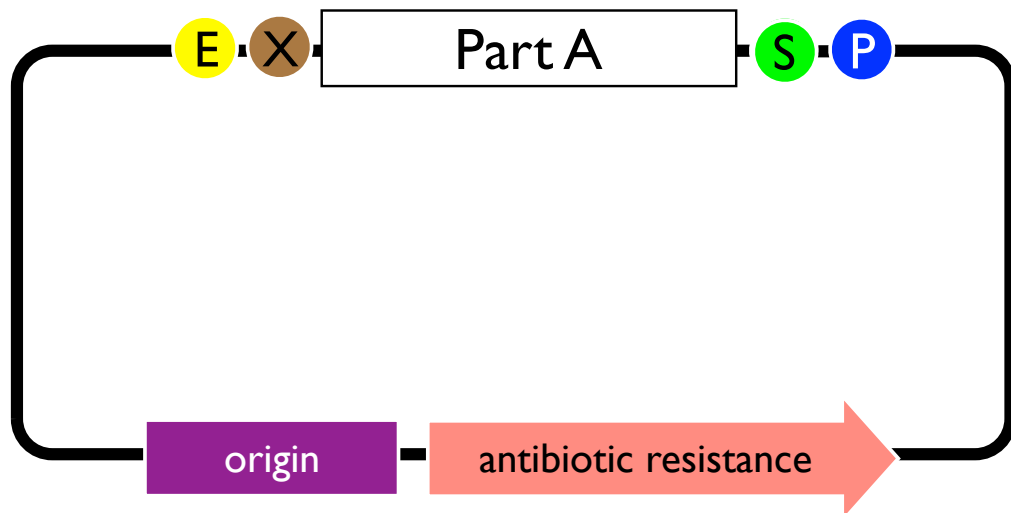


# Gel Purification

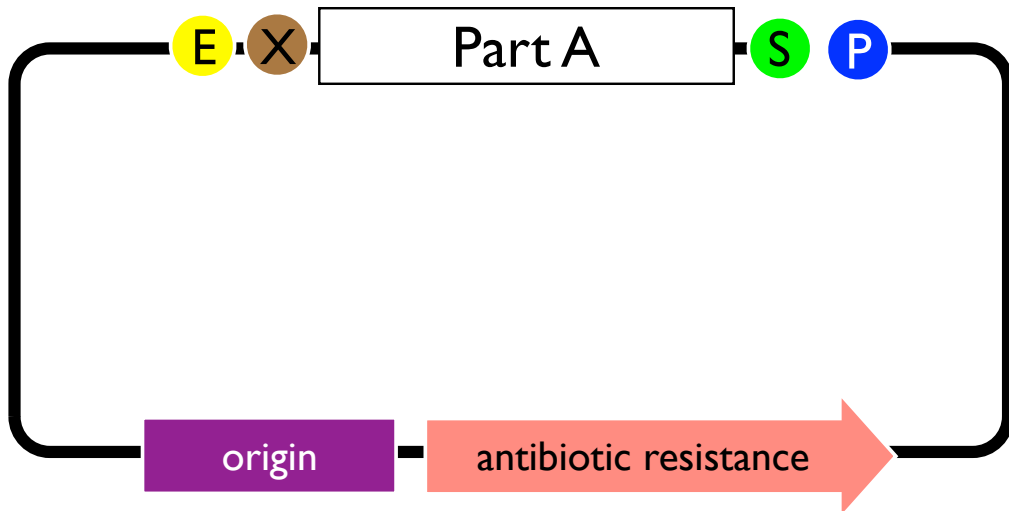




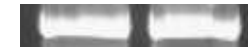
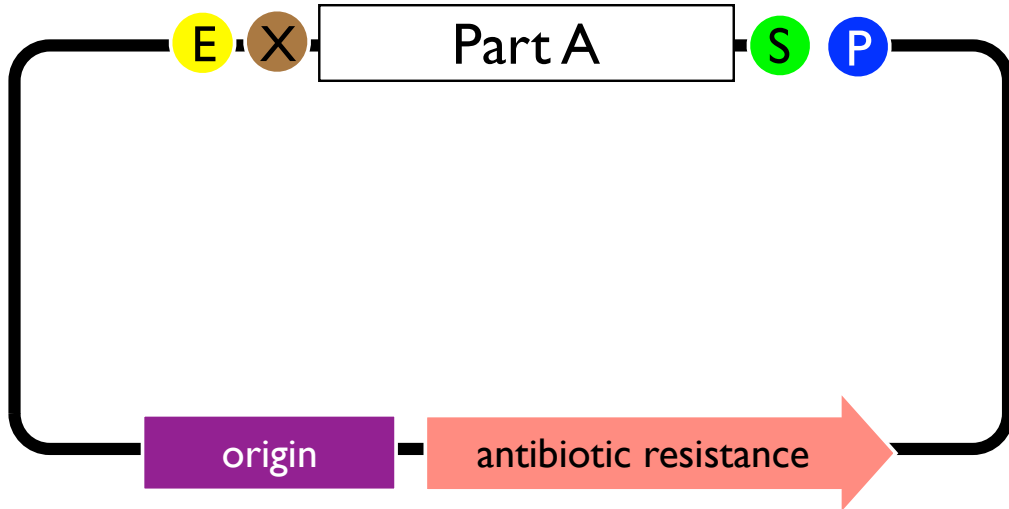
# Gel Purification



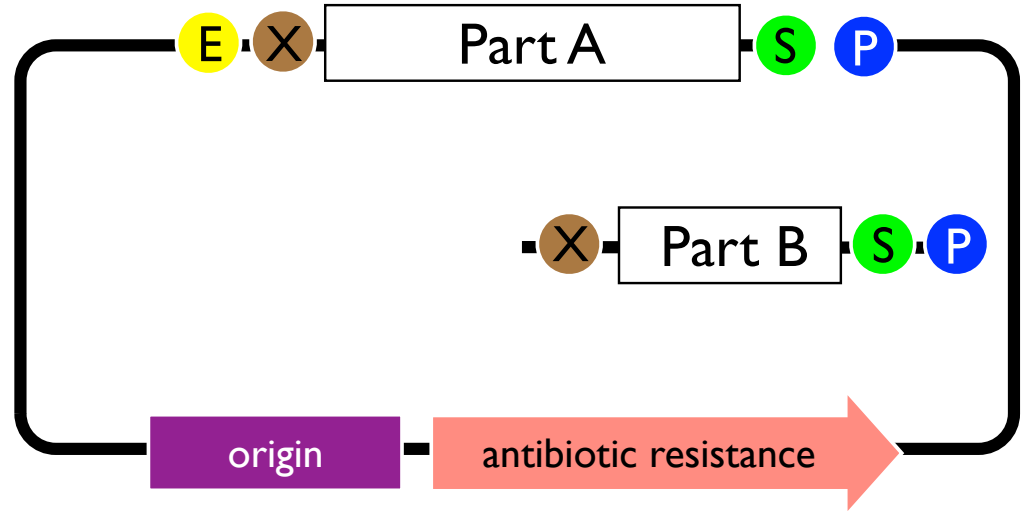
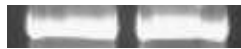
# Gel Purification



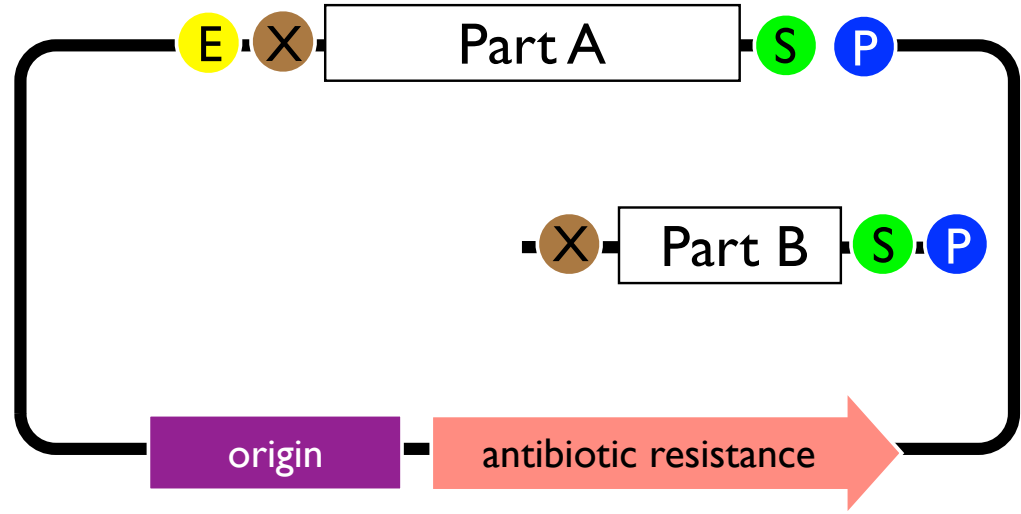
# Gel Purification



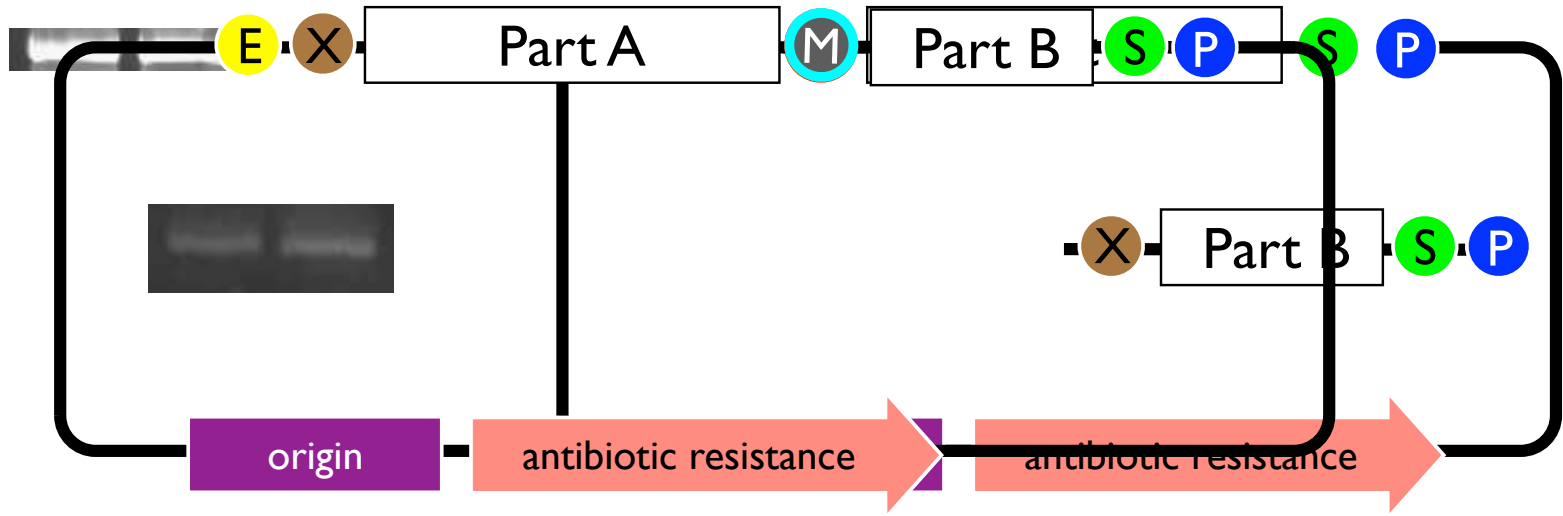
# Gel Purification



# Ligation

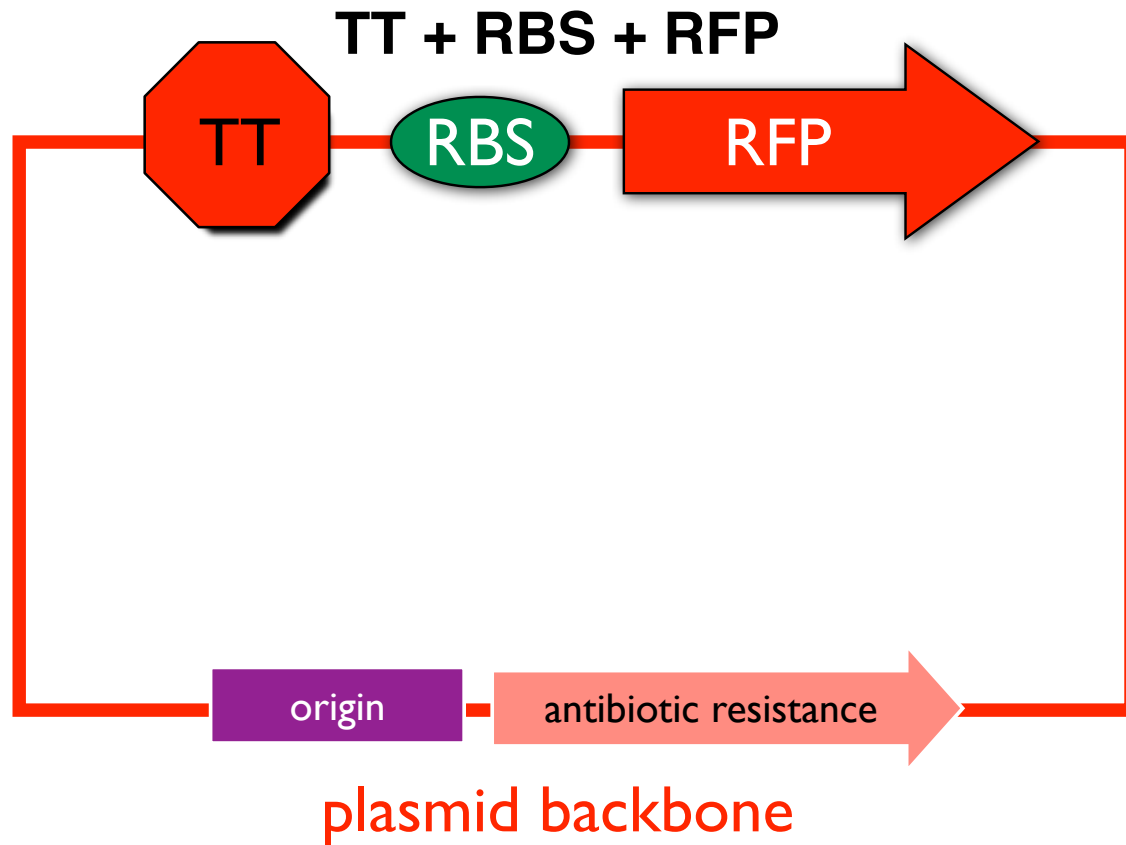
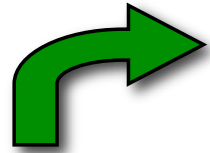


# Ligation



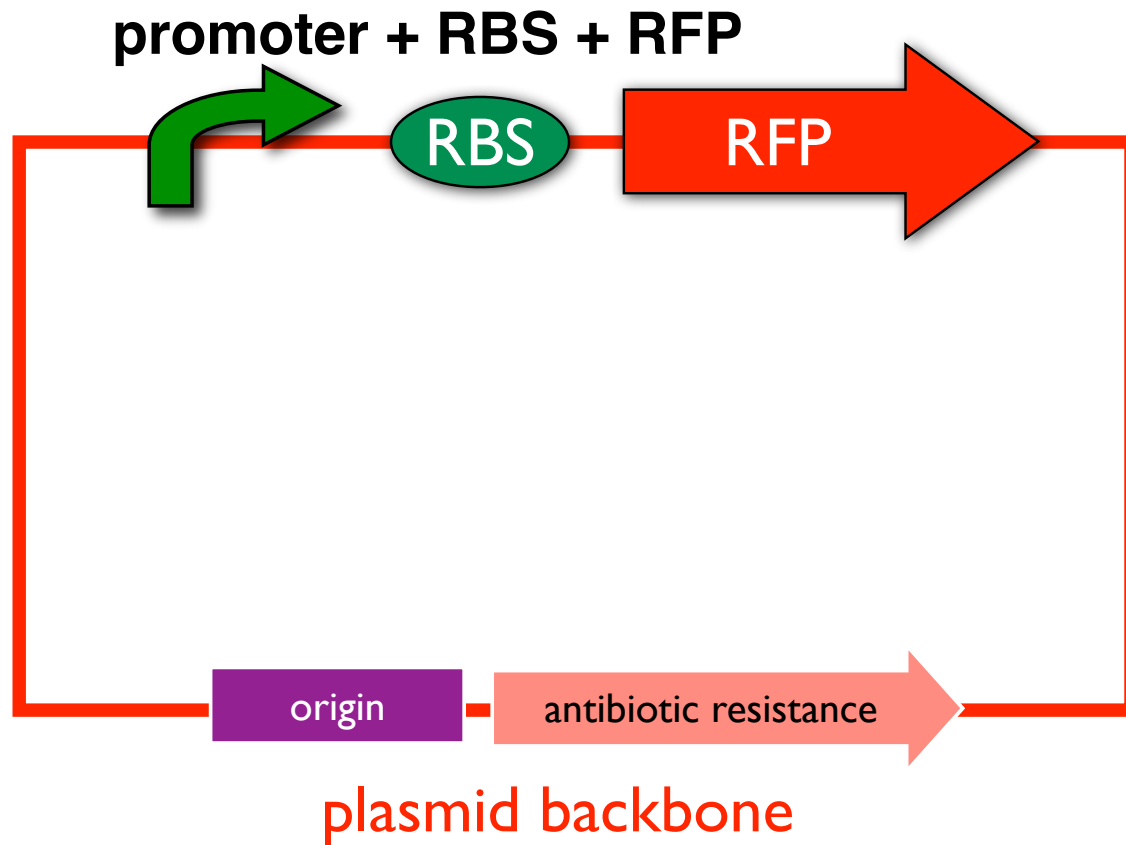
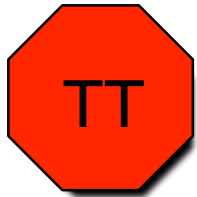
How can we clone DNA  
without all the hassle?

# Golden Gate Assembly Method





# Golden Gate Assembly Method



# Eco RI

GAATTC  
CTTAAG

palindrome

type II

# Eco RI

**G**AATTC  
CTTAAG

palindrome

type II

# Eco RI

GAATTC  
CTTAAG

A diagram showing the recognition sequence for the EcoRI restriction enzyme. The sequence is GAATTC on the top strand and CTTAAG on the bottom strand. A purple line indicates the cleavage site, which is between the G and A on the top strand and between the A and G on the bottom strand. The purple line starts at the G on the top strand, goes down, then right, then down again, then right, then down again, ending at the A on the bottom strand.

type II

# Eco RI

G

CTTAA

AATTC

G

type II

# Bsa I

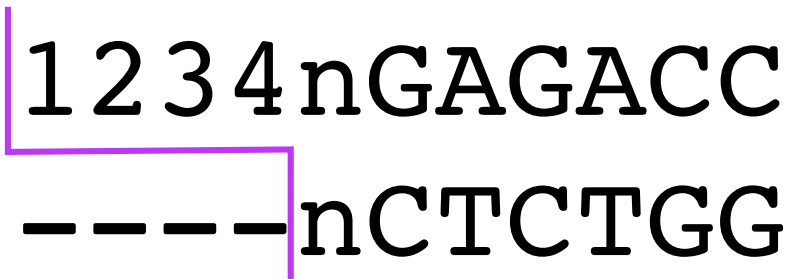
GAGACC  
CTCTGG

not a  
palindrome

type II

# Bsa I

1 2 3 4 n G A G A C C  
- - - - n C T C T G G

A diagram showing the recognition sequence for the Bsa I restriction enzyme. The top strand is 1 2 3 4 n G A G A C C, and the bottom strand is - - - - n C T C T G G. A purple line connects the first four positions (1-4) of the top strand to the first four positions (n-1) of the bottom strand, indicating a staggered cut. The 'n' represents a variable number of nucleotides between the recognition sequence and the cleavage site.

type II<sub>s</sub>

# Bsa I

1 2 3 4 n G A G A C C

n C T C T G G

-----

type II<sub>s</sub>



# Bsa I

GGTCTCn|-----  
CCAGAGn1234|

type II

# Bsa I

GGTCTCn

CCAGAGn1234

----

type IIs

# Bsa I

cuts  
left

1 2 3 4 n GAGACC  
-----n CTCTGG

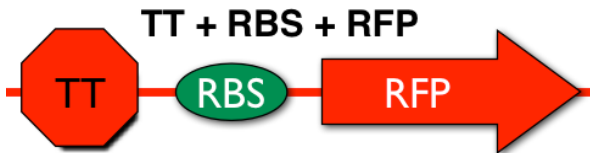
GGTCTCn----- cuts  
right

CCAGAGn 1 2 3 4

Bsa I



Bsa I



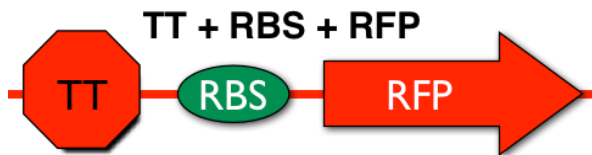
**CGACT****GAGACC** (**TT**) **GGTCTCa**  
**aCTCTGG** (**TT**) **CCAGAGt** **CGCC**

██████████  
██████████ **GCTG**

**GCGG** ██████████  
██████████



■■■■■ **CGAC** tGAGACC (**TT**) GGTCTCaGCGG ■■■■■  
■■■■■ GCTGaCTCTGG (**TT**) CCAGAGt**CGCC** ■■■■■



CGACtGAGACC (TT) GGTCTCa  
aCTCTGG (TT) CCAGAGtCGCC

██████████  
██████████ GCTG

GCGG ██████████  
██████████



CGAC (promoter)  
(promoter) CGCC

CGACtGAGACC (TT) GGTCTCa  
aCTCTGG (TT) CCAGAGtCGCC

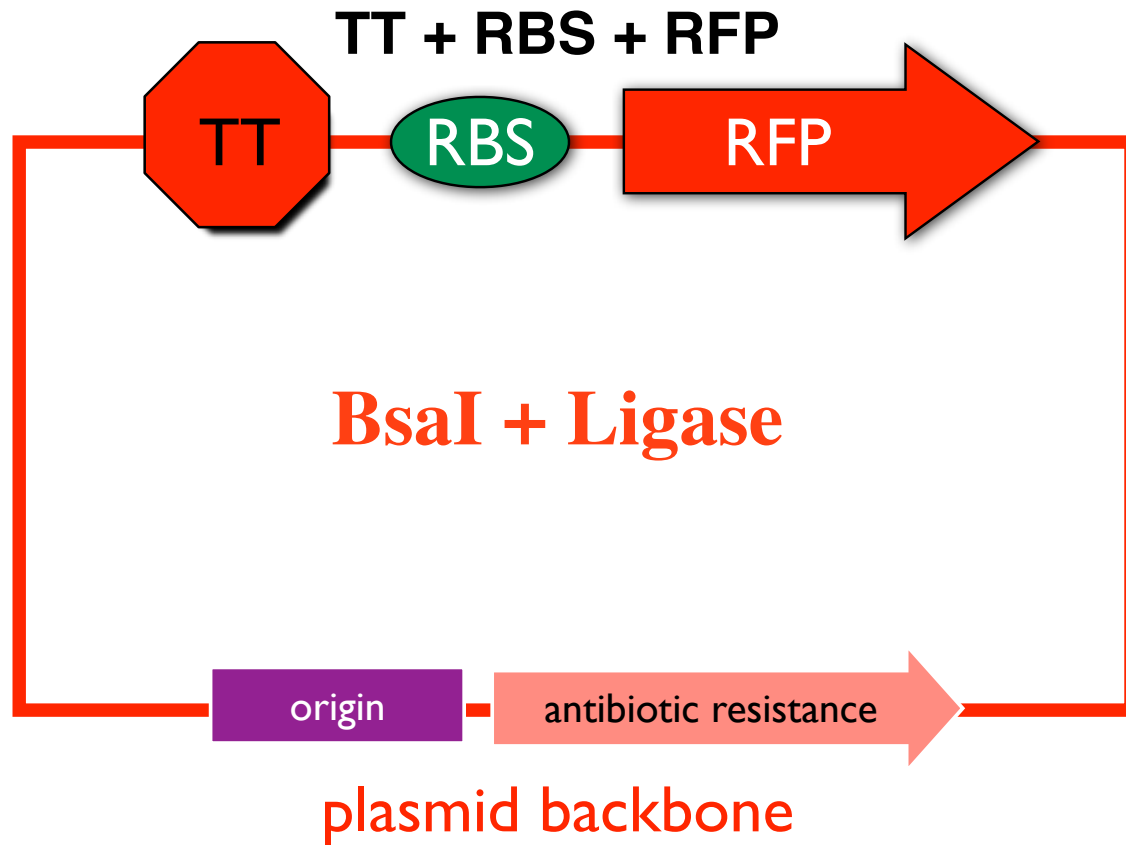
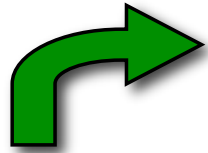
CGAC (promoter) GCGG  
GCTG (promoter) CGCC

promoter + RBS + RFP

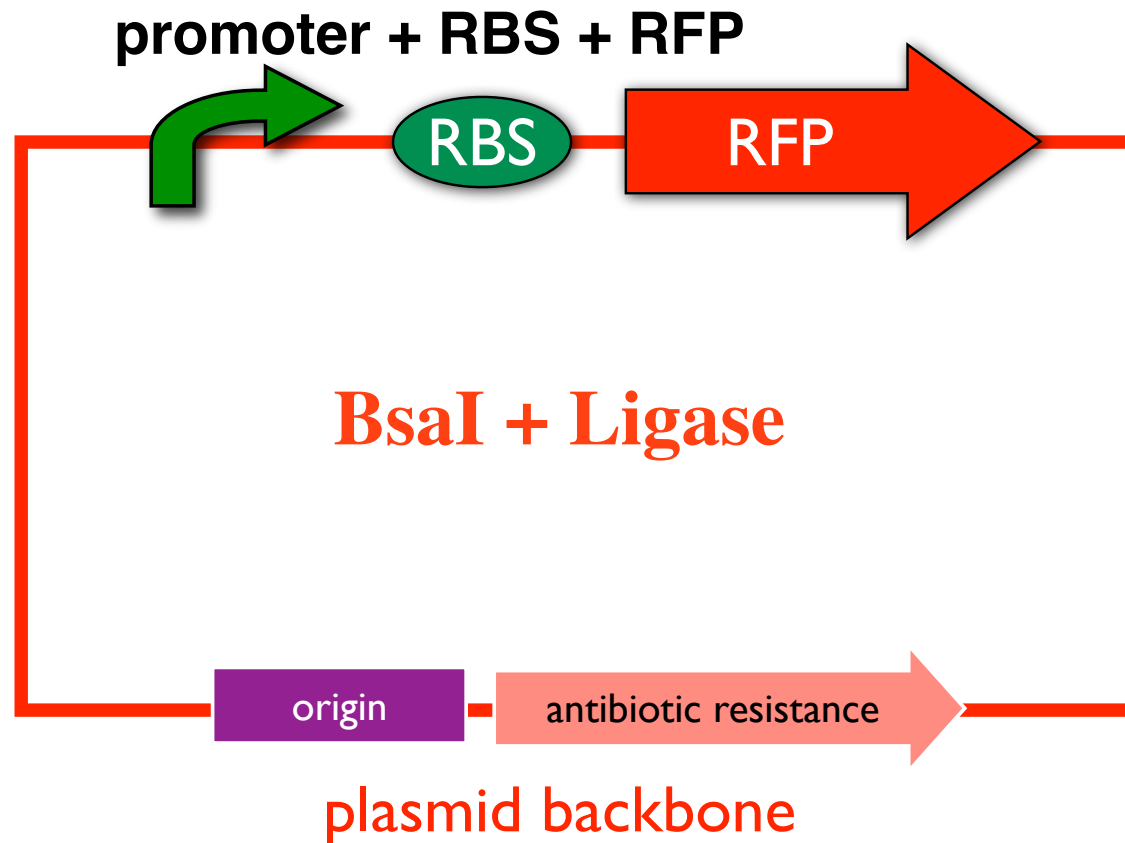
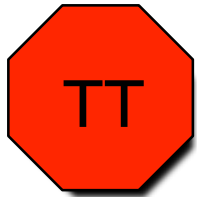




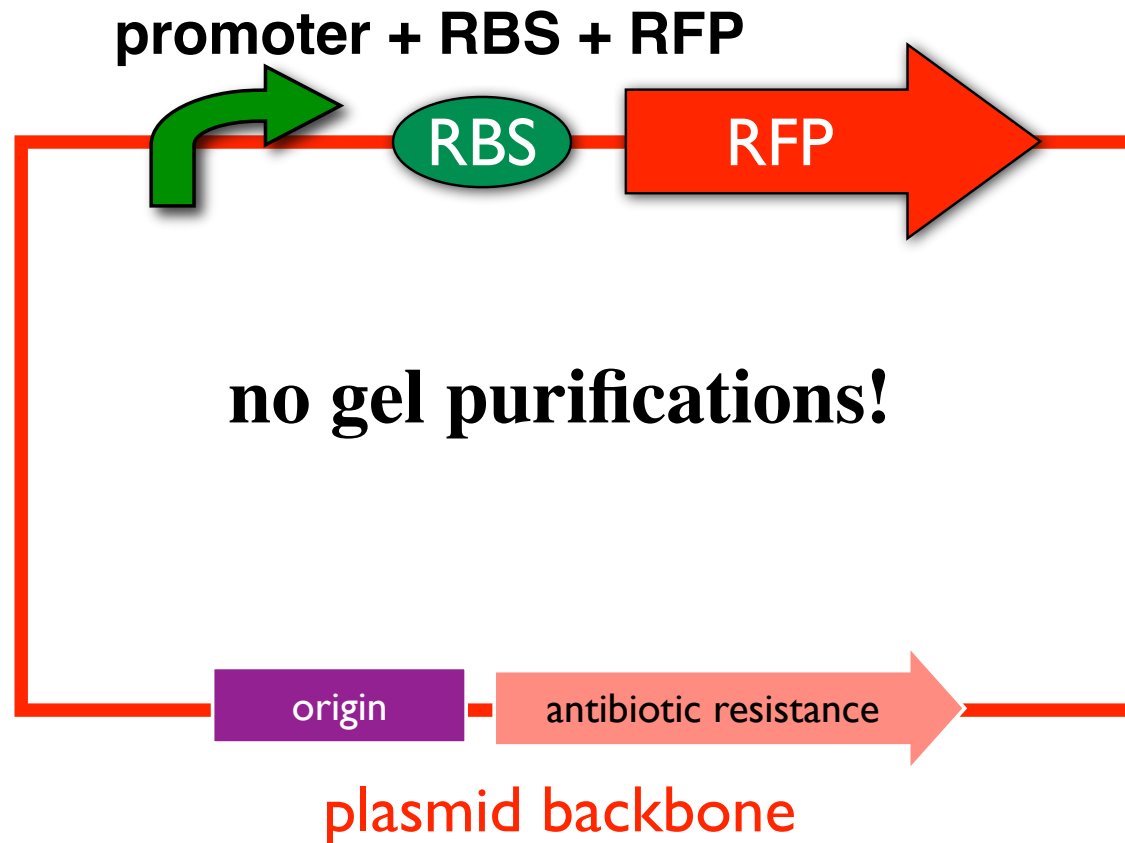
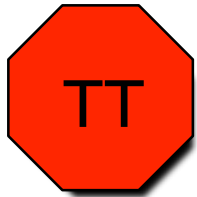
# GGA Ligation Method



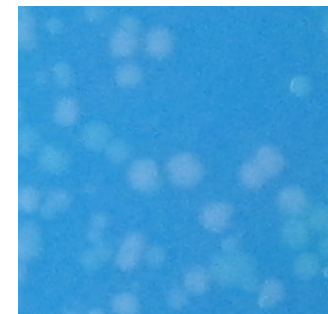
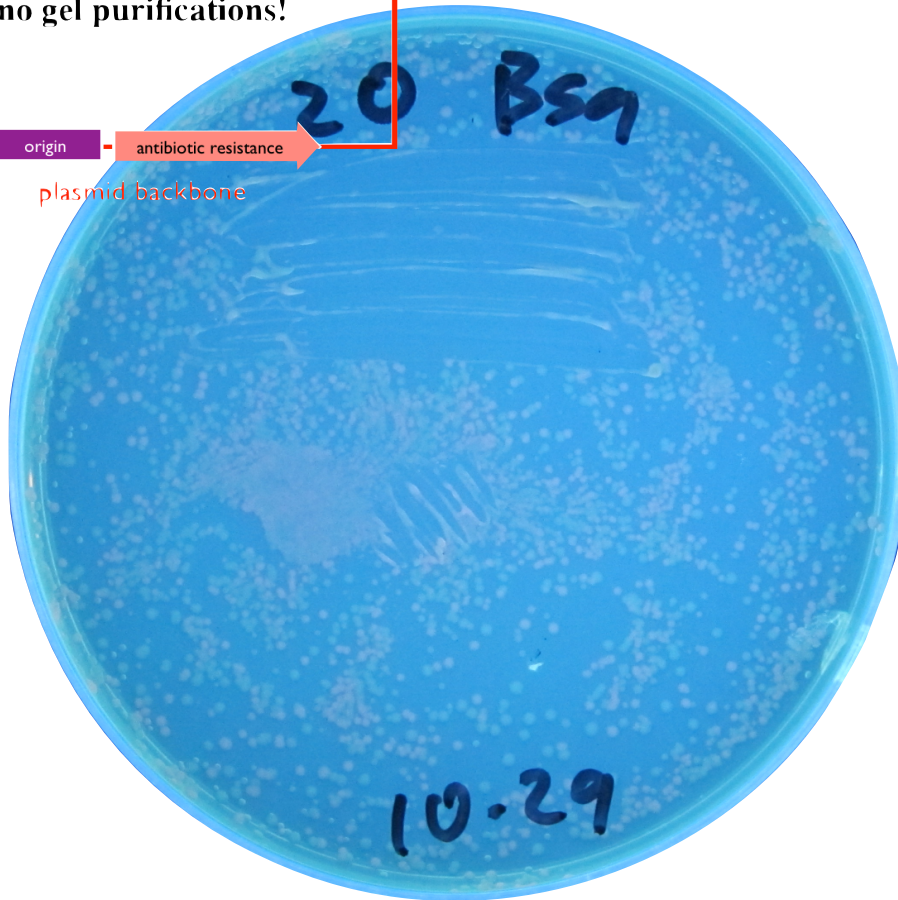
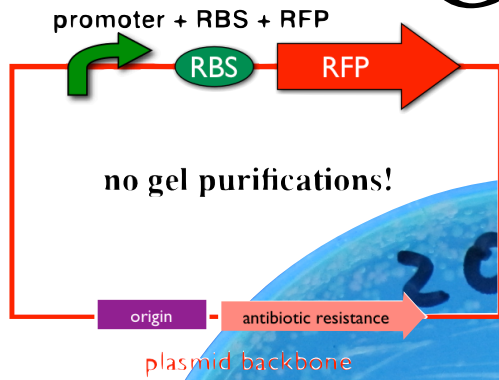
# GGA Ligation Method



# GGA Ligation Method



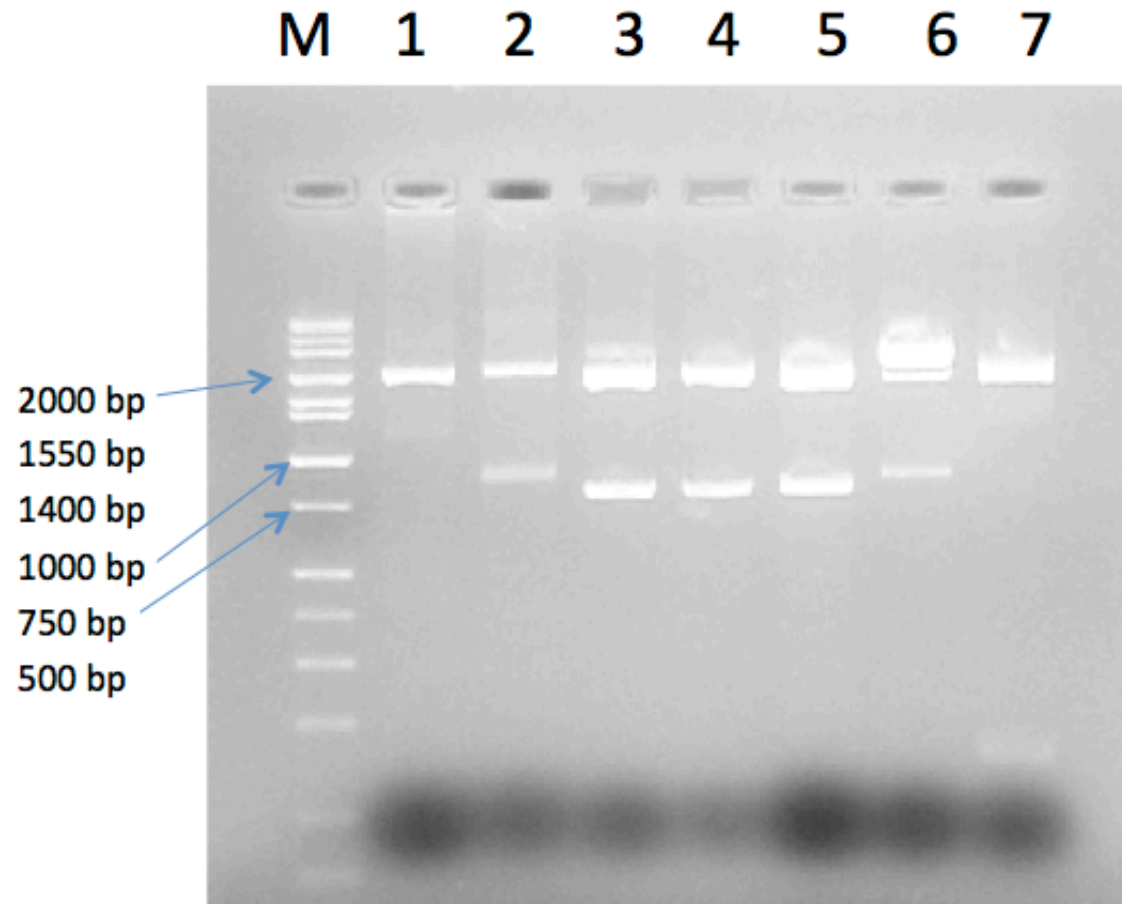
# GGA Ligation Method



# GGA Ligation Confirmation

All digested with R+P

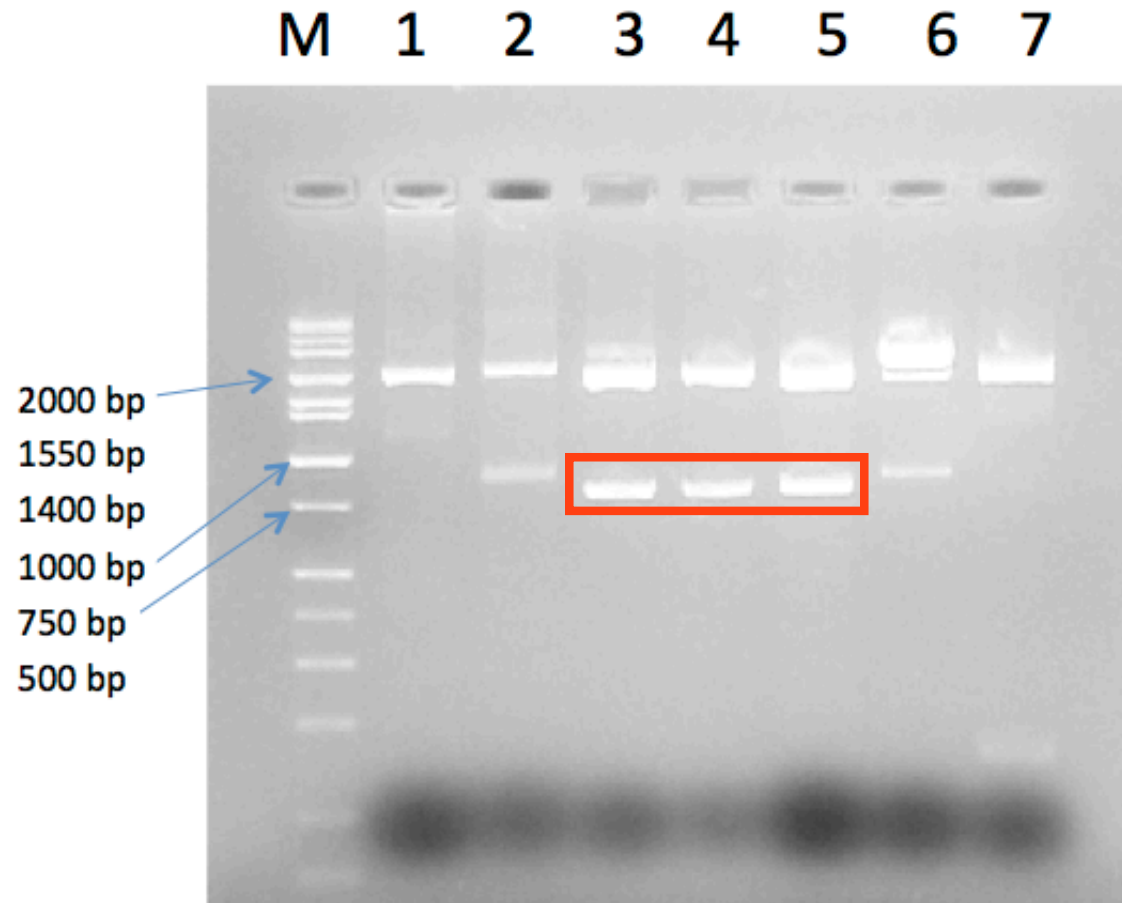
- 1 J119022
- 2 J100028
- 3 20 cycle red
- 4 20 cycle red
- 5 20 cycle red
- 6 20 cycle white
- 7 20 cycle white



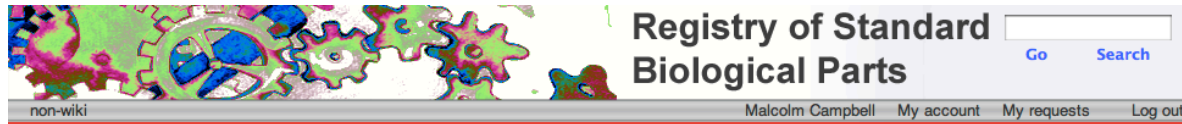
# GGA Ligation Confirmation

All digested with R+P

- 1 J119022
- 2 J100028
- 3 20 cycle red
- 4 20 cycle red
- 5 20 cycle red
- 6 20 cycle white
- 7 20 cycle white



# Registry of Functional Promoters



Registry of Standard Biological Parts

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## Campbell M Lab Parts

### Favorite Campbell M Lab Parts

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Name	Type	Description	Designer	Length
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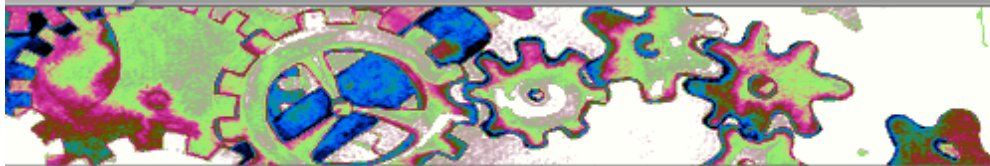
### Campbell M Lab Parts Sandbox

[Edit](#)

Name	Type	Description	Designer	Length
BBa_J100000	Coding	Cre with 8bp restriction sites and 1-Clause 2-SAT Problem Inserted	Eric Sawyer	1069
BBa_J100001	Composite	pTet+RBS+Cre2SAT1Clause+pLpp+iRNA CCACU	Eric Sawyer	1357
BBa_J100002	Composite	pTet+RBS+Cre2SAT1Clause+pLpp+iRNA CCGUC	Eric Sawyer	1357
BBa_J100003	Generator	pTet+RBS+Cre2SAT1Clause	Eric Sawyer	1149
BBa_J100004	Reporter	pTet+LoxP+RBS+RFP+LoxP	Eric Sawyer	870
BBa_J100005	Other	Palindromic Stop Sequence	Eric Sawyer	221
BBa_J100006	Intermediate	LoxP+Stop Sequence+LoxP	Eric Sawyer	305
BBa_J100007	Intermediate	pLac+RBS+LoxP+Stop Sequence+LoxP	Eric Sawyer	533
BBa_J100008	Composite	pLpp-iRNA CCACU-pLpp-iRNA CUAGU	Eric Sawyer	408
BBa_J100009	Composite	pLpp-iRNA CCACU-pLpp-iRNA CCGUC	Eric Sawyer	408
BBa_J100010	Composite	pLpp-iRNA CUAGU-pLpp-iRNA CCGUC	Eric Sawyer	408
BBa_J100011	Composite	pLpp-iRNA CCACU-pLpp-iRNA CUAGU-pLpp-iRNA CCGUC	Eric Sawyer	616
BBa_J100012	Intermediate	RBS-RFP-RBS	Eric Sawyer	747
BBa_J100013	Coding	LuxI with 1 Clause 2-SAT Problem	Eric Sawyer	638
BBa_J100014	Coding	LuxI with 2 Clause 2-SAT Problem	Eric Sawyer	652
BBa_J100015	Composite	1 Clause 2-SAT Problem with Frameshifted LuxI and a GFP Reporter	Eric Sawyer	2757
BBa_J100016	Composite	2 Clause 2-SAT Problem with Frameshifted LuxI and a GFP Reporter	Eric Sawyer	2771
BBa_J100017	Composite	TT+pLux+RBS+LuxI(2-SAT 2 clause)+RBS+GFP+pLac+RBS+LuxR+iRNAs	Eric Sawyer	3395
BBa_J100018	Protein_Domain	First Half of AspC gene	Catherine Doyle	448
BBa_J100019	Protein_Domain	First half of ivE gene	Julia Fearrington	457
BBa_J100020	Protein_Domain	Second Half of AspC	Catherine Doyle	869
BBa_J100021	Protein_Domain	First Half of PyrE	Catherine Doyle	488
BBa_J100022	Protein_Domain	Second Half of PyrE	Catherine Doyle	280
BBa_J100025	Protein_Domain	First half of CAT gene	James Harden	434
BBa_J100026	Protein_Domain	second half ivE gene	Julia Fearrington	574
BBa_J100027	Protein_Domain	second half of TyrB	James Harden	288
BBa_J100028	Other	placeholder insert for Bsal Golden Gate Assembly of promoter	Malcolm Campbell	877
BBa_J100029	Regulatory	The promoter of rpoDPhs	Maggie Baay	76
BBa_J100030	Regulatory	phoA is an inducible promoter induced by phosphate starvation.	Scott Hall	76
BBa_J100031	Regulatory	Constitutive promoter C on Gene 1 of T7, transcribes RNA Pol.	Caroline Vrana	100
BBa_J100032	Regulatory	proUP3 promoter	Molly Marshall	90
BBa_J100033	Regulatory	dnakP1 promoter: Heat shock inducible	Chris Peek	101
BBa_J100034	Regulatory	groE promoter	Margaret Stebbins	44
BBa_J100036	Regulatory	Promoter induced by DNA damage	Erich Baker	52
BBa_J100039	Regulatory	GalP1 Promoter-Induced By Galactose	Anaiah Toby	75
BBa_J100040	Coding	LuxI with 3 clause 2-SAT problem	Eric Sawyer	684
BBa_J100041	Composite	LuxI/GFP with 3 clause 2-SAT problem	Eric Sawyer	2803
BBa_J100042	Coding	LuxI with 3 clause 3-SAT problem	Eric Sawyer	702
BBa_J100043	Composite	LuxI/GFP with 3 clause 3-SAT problem	Eric Sawyer	2821
BBa_J100044	Coding	LuxI with 4 clause 2-SAT problem	Eric Sawyer	704
BBa_J100045	Composite	LuxI/GFP with 4 clause 2-SAT problem	Eric Sawyer	2823
BBa_J100046	RNA	lpp+iRNA CCAUC (10 bp anticodon loop)	Eric Sawyer	201
BBa_J100047	Protein_Domain	TyrB2	Julia Fearrington	930
BBa_J100048	Protein_Domain	TyrB1	Julia Fearrington	930
BBa_K091231	Composite	LuxR producer and XOR gate	Malcolm Campbell	2772
BBa_K091232	Composite	LuxR producer and RFP(rev) + RBS(rev) + pLux (for)	Malcolm Campbell	1916



# Student Sample



## Registry of Standard Biological Parts

  
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[BBa J100033 Main Page](#) [Part Design](#) [Physical DNA](#) [Hard Information](#) [Experience](#) [Tools](#)

## Part:BBa\_J100033

Designed by Chris Peek Group: Campbell\_M\_Lab (2011-09-01)



Regulatory

[DNA Planning](#)

[Experience:](#)

[Get This Part](#)

### dnakP1 promoter: Heat shock inducible

dnakP1 is naturally off, but is induced when E. coli is heat shocked, resulting in transcription downstream from this promoter.

### Sequence and Features

Format:	<a href="#">Subparts</a>	<a href="#">Ruler</a>	<a href="#">SS</a>	<a href="#">DS</a>	Search:	Length: 101 bp	Context: Part only	<a href="#">Get selected sequence</a>				
1	11	21	31	41	51	61	71	81	91			
1	aaatttctgc	gcaaaagcac	aaaaaat	ttt	tgc	atctccc	ccttgatgac	gtggtttacg	acccattta	gtagtcaacc	gcagtgagtg	agtctgcaa
	tttaaagacg	cgtttctgtg	tttttaaaa	acgtagaggg	ggaactactg	caccaa	atgc	tggggtaaat	catcagttg	cgtcactcac	tcagacg	ttt
101	a											
	t											

Assembly Compatibility: [10](#) [12](#) [21](#) [23](#) [25](#)



# Student Sample

Registry of Standard Biological Parts

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BBa\_J100033 Main Page Part Design Physical DNA Hard Information Experience Tools

## Part:BBa\_J100033:Experience

Designed by Chris Peek Group: Campbell\_M\_Lab (2011-09-01)



DNA Planning

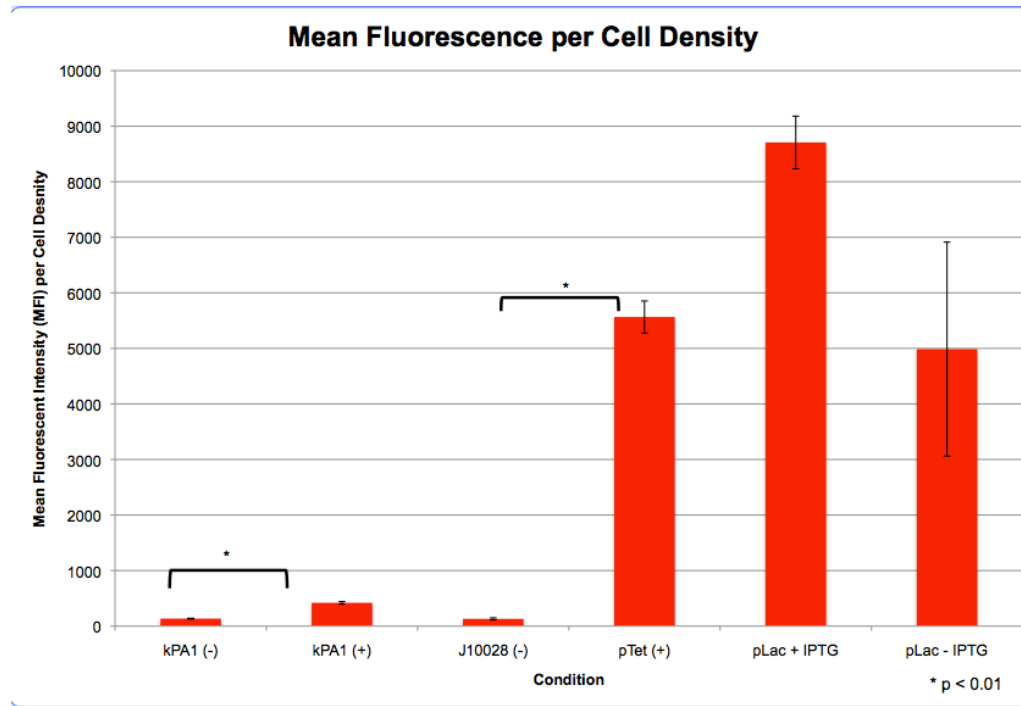
Experience:

[Get This Part](#)

This experience page is provided so that any user may enter their experience using this part. Please enter how you used this part and how it worked out.

### Applications of BBa\_J100033

[\[edit\]](#)



A: Experimental:

cells containing dnaKP1 without heat shock (incubated at 37°C) B: Experimental: cells containing dnaKP1 with heat shock (incubated at 40°C) C: Negative control: part i100028 without pTet promoter D: Positive control: part i100028 with pTet promoter (always on) E: pLac promoter (part i715039) with inducer (IPTG) F: pLac

# What's Next?

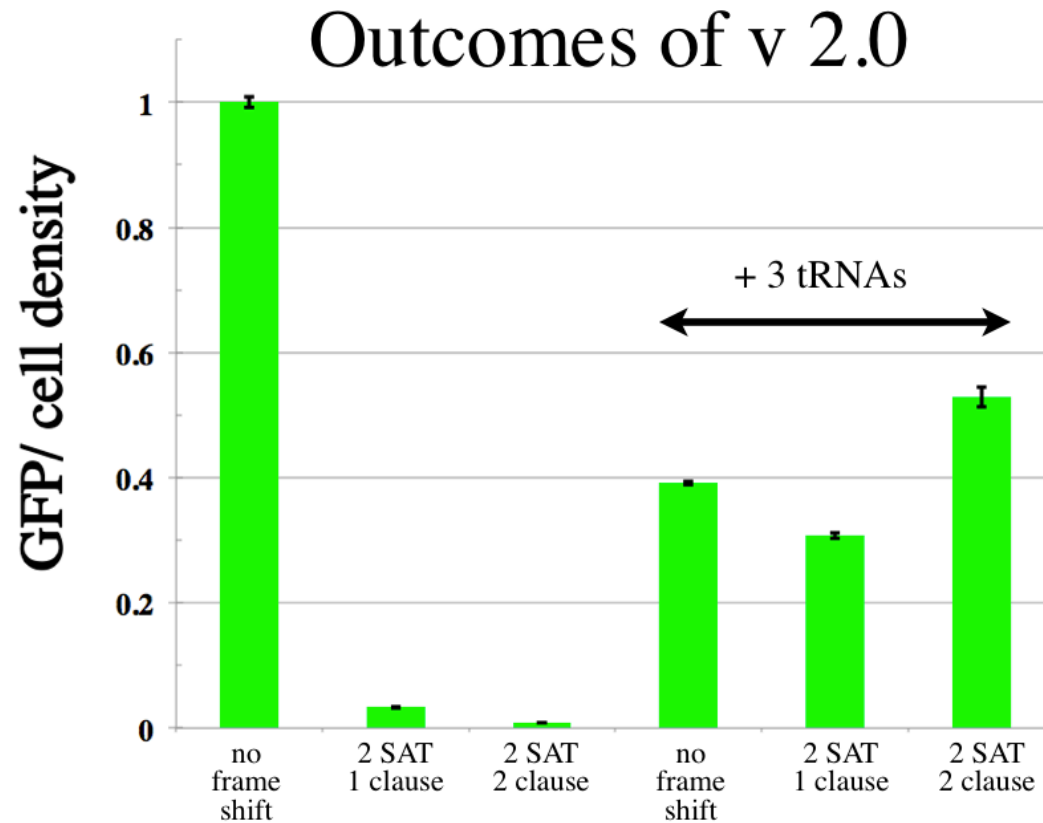
Improve the plasmid to increase phenotype.

Simplify ligation/digestion protocol.

Unleash intro bio students for real research!

# Three Rules for Our Lab

1. Everyone has to learn.



# Three Rules for Our Lab

1. Everyone has to learn.
2. Everyone has to have fun.



# Three Rules for Our Lab

1. Everyone has to learn.
2. Everyone has to have fun.
3. We try to contribute to the body of science.



# Acknowledgements

**Faculty:** Laurie Heyer, Jeff Poet, Todd Eckdahl, Karmella Haynes, Pat Sellers, Mark Barsoum

**Students:** Romina Clemente, Clif Davis, A.J. Grant, Mary Gearing, Kin Lau, Olivia Ho-Shing, Shamita Punjabi, Eric Sawyer, Ashley Schooner, Siya Sun, Shashank Suresh, Bryce Szczepanik, Leland Taylor, Annie Temmink, Alyndria Thompson, Will Vernon, Oyinade Adefuye, Will DeLoache, Jim Dickson, Andrew Martens, Amber Shoecraft, Mike Waters, Jordan Baumgardner, Tom Crowley, Lane Heard, Nick Morton, Michelle Ritter, Karen Acker, Bruce Henschen, Jessica Treece, Matt Unzicker, Amanda Valencia, Lance Harden, Sabriya Rosemond, Samantha Simpson, Erin Zwack, Marian Broderick, Adam Brown, Trevor Butner, Lane Heard, Eric Jessen, Kelley Malloy, Brad Ogden, Kelly Davis, Alicia Allen, James Barron, Robert Cool, Kelly Davis, Will DeLoache, Erin Feeney, Andrew Gordon, John Igo, Aaron Lewis, Kristi Muscalino, Madeline Parra, Pallavi Penumetcha, Karlesha Roland, Max Win, Xiao Zhu, Kristen DeCelle, Matt Gemberling, Oscar Hernandez, Andrew Drysdale, Nick Cain, Tamar Odel, and Jackie Ryan.

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Genome Consortium for Active Teaching (GCAT)  
Davidson College James G. Martin Genomics Program  
MWSU SGA, Foundation & Summer Research Institute



# The End

