

Efficient bacterial transcription of DNA nanocircle vectors with optimized single-stranded promoters

Tatsuo Ohmichi*, Angèle Maki, and Eric T. Kool†

Department of Chemistry, University of Rochester, Rochester, NY 14627; and Department of Chemistry, Stanford University, Stanford, CA 94305-5080

Communicated by John I. Brauman, Stanford University, Stanford, CA, November 2, 2001 (received for review July 10, 2001)

We describe experiments aimed at establishing whether circular single-stranded DNAs can form promoters for bacterial transcription from small folded motifs. *In vitro* selection experiments were carried out on circular 103-nt DNA libraries encoding 40-nt randomized sequences as well as self-processing hammerhead ribozymes. Rounds of rolling circle transcription, reverse transcription-PCR, and recyclicalization were carried out to optimize transcription efficiency. Sequences were identified that are 80-fold more actively transcribed than the initial library by *E. coli* RNA polymerase (RNAP). The selected motifs were found to be more active than canonical *E. coli* promoters in the same context. Experiments also demonstrated that a single-stranded pseudopromoter identified by this selection can be transplanted to other circular DNA contexts and retain transcriptional activity. Results suggest that the promoter is localized in a short (≈ 40 nt) hairpin, which is smaller than canonical *E. coli* promoters. To test whether this pseudopromoter was active in bacterial cells, a synthetic DNA nanocircle vector encoding a ribozyme targeted to a site in the *marA* drug resistance gene was constructed to contain an optimized single-stranded promoter. It is shown that this DNA circle can act as a “Trojan horse” in *E. coli*, being actively transcribed by the cellular RNAP and producing ribozymes that cleave a sequence in the *marA* drug resistance gene. The use of optimized single-stranded promoters in combination with synthetic nanocircle DNA vectors represents a potentially useful way to engender the synthesis of biologically active RNAs in living cells.

rolling circle transcription | hammerhead ribozyme | *in vitro* selection

Recent work has shown that very small circular single-stranded DNAs (ssDNAs) can be efficiently transcribed by phage and bacterial RNA polymerases (1–7). Transcription proceeds by a “rolling-circle” mechanism, generating long RNA repeats encoded by the circular DNA. This rolling circle transcription (RCT) has been demonstrated for circles in the size range 18 to ≈ 110 nt, or roughly 2–10 nanometers in diameter (6, 7), and the long product RNAs have been imaged under the atomic force microscope (8–10). Such DNA circles can be used to encode concatameric or self-cleaving monomeric ribozymes of the hammerhead (2), hairpin (3), and hepatitis delta motifs (4). Although evidence in one case suggests that transcription by an RNA polymerase of unstructured circular ssDNAs can initiate with GTP at any cytosine in a circle (2, 11), it was also observed that secondary structures in circular DNAs can strongly inhibit or enhance transcription in some cases.

It has been known for some time that RNA polymerases can initiate transcription in single-stranded “bubble” regions within duplex DNA (12–14), and in single-stranded DNA homopolymers, particularly in cytosine-rich sequences (15–18). However, to our knowledge, no broad range of sequences or structures have been surveyed for such activity. We sought to investigate whether there are specific sequences and/or structures of single-stranded DNA that would act as promoters for a bacterial RNA polymerase. Here we report the identification of such sequences and structures by an *in vitro* selection strategy. We describe a motif that is formed in only ≈ 40 nucleotides of contiguous sequence, or considerably smaller than a previously reported

naturally occurring single-stranded promoter in *E. coli* (19). It appears to be general, in that it can be transplanted from one context to another and still direct efficient transcription by the bacterial RNA polymerase. We further demonstrate that this promoter can be used in a DNA nanocircle vector (a “rolling circle” vector built from ssDNA) to direct transcription of an active hammerhead ribozyme in *E. coli* cells.

Materials and Methods

Preparation of Oligonucleotides and Circular DNA Library. An initial circular ssDNA library containing 63 nt of fixed sequence and 40 nt of randomized sequence was generated by sequential enzymatic ligations of 5'-phosphorylated 56-nt and 47-nt oligonucleotides by using T4 DNA ligase (New England Biolabs) and 16-nt splint oligonucleotides as described (2). The 5'-phosphorylated sequences were: 5'-pTTC GTC TG-N₄₀-TCT TTC AG-3' and 5'-TTT CGT CCT CAC GGA CTC ATC AGA ATG GCA ACA CAT TGA CTG AGG AG-3'. Details are given in supporting *Materials and Methods*, which is published as supporting information on the PNAS web site, www.pnas.org.

***In Vitro* Selection.** Conditions for initial RCT reaction were: 1 μ M circular DNA, 2 units *E. coli* RNA polymerase holoenzyme (Boehringer Mannheim), 0.5 mM ATP, CTP, and GTP, 60 μ M UTP, and 0.30 μ Ci (1 Ci = 37 GBq) of [α -³²P]UTP in 25 mM Tris·HCl (pH 8.1) buffer containing 20 mM NaCl, 15 mM MgCl₂, 0.4 mM spermine·HCl, 100 μ g/ml acetylated BSA, 10 mM DTT, and 12.5 units/ml RNase inhibitor (Promega), in a total reaction volume of 15 μ l. After a 1.5-h incubation at 37°C, the reaction was terminated by adding an equal volume of stop solution (30 mM Na₂EDTA/8 M urea/0.02% bromophenol blue/0.02% xylene cyanol). Self-processed 103-nt product RNAs were purified by 10% denaturing PAGE gels run at 4°C.

After elution from the gels, the selected 103-nt ssRNAs were reverse transcribed (Invitrogen) with a 5'-phosphorylated 18-nt primer (pGAC TGA GGA GTT CGT CTG) for 1 h at 42°C. After buffer exchange with Bio-spin column (Bio-Rad), the cDNA products were PCR amplified with *Taq* polymerase for 15 cycles (temperature cycle: 94°C, 1 min; 55°C, 1 min; 72°C, 1 min) in the presence of 100 pmol each of two primers: 5'-pGAC TGA GGA GTT CGT CTG-3' and 5'-biotin-AAT GTG TTG CCA TTC TGA-3'. The PCR products were extracted with phenol/CHCl₃, ethanol precipitated, and blunted with T4 DNA polymerase (LifeTechnologies, Grand Island, NY).

The blunt-ended PCR products were immobilized on magnetic beads with streptavidin (Dynal, Great Neck, NY) in the presence of 80 μ M binding buffer [1 M NaCl/10 mM Tris·HCl

Abbreviations: RT, reverse transcription; CAT, chloramphenicol transferase; RCT, rolling circle transcription; ssDNA, single-stranded DNA.

*Present address: High Technology Research Center, Konan University, 8-9-1 Okamoto, Higashinada-ku, Kobe 658-8501, Japan.

†To whom reprint requests should be addressed at: Department of Chemistry, Stanford University, Stanford CA 94305-5080. E-mail: kool@stanford.edu.

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked “advertisement” in accordance with 18 U.S.C. §1734 solely to indicate this fact.

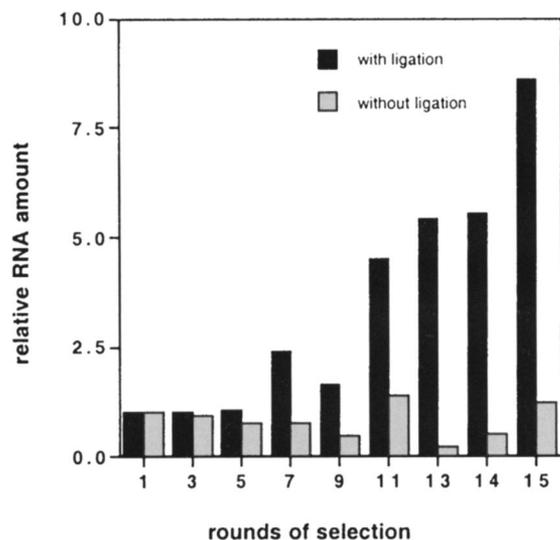


Fig. 2. Improvement of transcription activity over successive rounds of *in vitro* selection. RNA amount was measured for each successive population at 37°C after 1.5 h. Dark and light bars correspond to the relative RNA amounts (>80-nt product) for the successive population with and without ligation, respectively. The reaction conditions are described in *Materials and Methods*.

sequences that are efficient at being amplified by RT-PCR. To encourage optimization of transcription, we kept transcription times relatively short and self-processing and ligation times relatively long.

We carried out fifteen rounds of selection and amplification by this approach. Fitness of the library (measured by total amounts of RNA >80 nt produced) generally increased over the generations (Fig. 2). After fifteen rounds the fitness was increased several-fold over the original randomized set, and did not increase with further rounds (data not shown). Ligation to circular form was necessary for increasing fitness, because the unligated DNAs did not show increasing production of RNA (Fig. 2), consistent with a rolling circle mechanism of transcription. We then cloned the PCR fragments after round 15 and sequenced plasmids from 38 individual colonies. Selected sequences are shown in Fig. 3.

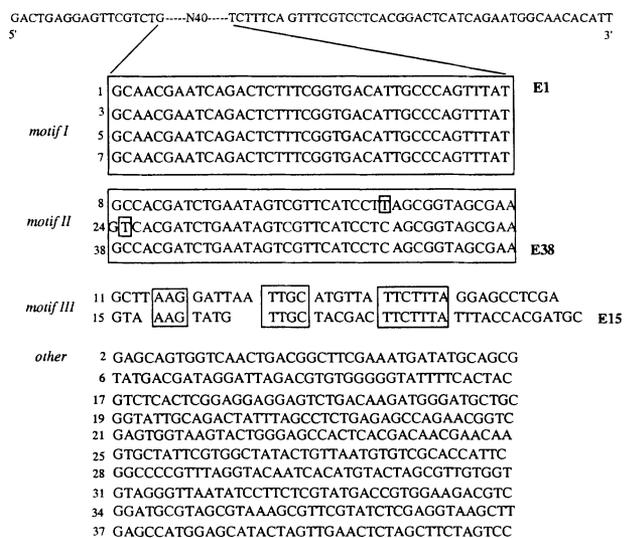


Fig. 3. Sequences of clones obtained following the fifteenth round of *in vitro* selection. Boxes indicate regions of high sequence similarity.

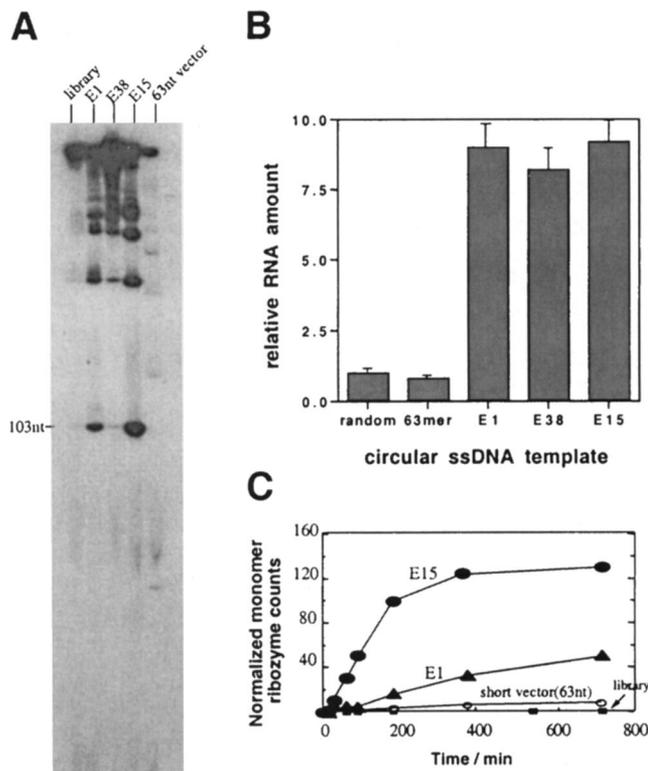


Fig. 4. Selected circular DNA motifs engender RNA synthesis *in vitro* with *E. coli* RNAP. (A) Autoradiogram of denaturing 10% polyacrylamide gel showing *in vitro* transcription of the 103-nt initial library, a control 63-nt nanocircle lacking the randomized domain, and selected individual nanocircles E1, E15, and E38 (after 1.5 h). (B) The relative total RNA amounts (all lengths >80 nt) for the 103-nt initial library, 63-nt nanocircle lacking the randomized domain, and E1, E15, and E38. (C) Time course of the production of monomeric ribozyme for the 103-nt initial library (■), 63-nt nanocircle lacking the randomized domain (○), E1 (▲), and E15 (●).

Selection Yields Strongly Transcribed Motifs in Circular ssDNAs. The results from sequencing revealed three main sequence motifs that were represented more than once, as well as ten sequences represented only once (Fig. 3). The presence of multiple examples of some of the motifs suggests that selection had indeed enriched the population with cases having superior fitness. Overall, there is a small bias toward T-rich sequences (28% T) and toward a deficiency in C (22% C). It is possible that this reflects the known preference of *E. coli* RNA polymerase (RNAP) for initiation with ATP opposite T, although some of the best-transcribed sequences are not T-rich (e.g., E38, see below). Alternatively, it may reflect some nucleotide bias in the initial library.

We evaluated fitness of individual selected motifs by separately examining one member of each of the three most common motifs. The individual DNA circles (labeled E1, E15, and E38 in Fig. 3) were constructed in the same way as the library and were then evaluated for their ability to engender RNA synthesis *in vitro* with *E. coli* RNA polymerase. The results showed (Fig. 4A and B) that all three were considerably better transcribed than either the initial randomized nanocircle library, or than a 63-mer nanocircle lacking the randomized domain altogether. In addition to enhancement of total RNA synthesis, the rate of production of monomeric ribozymes was also enhanced for E15 and E1 (Fig. 4C), with that for E15 being the greatest. Motif E38 produced RNA that was relatively slow at self-processing (Fig. 4A) but produced the most total RNA, being 80-fold more strongly transcribed than the original circular library.

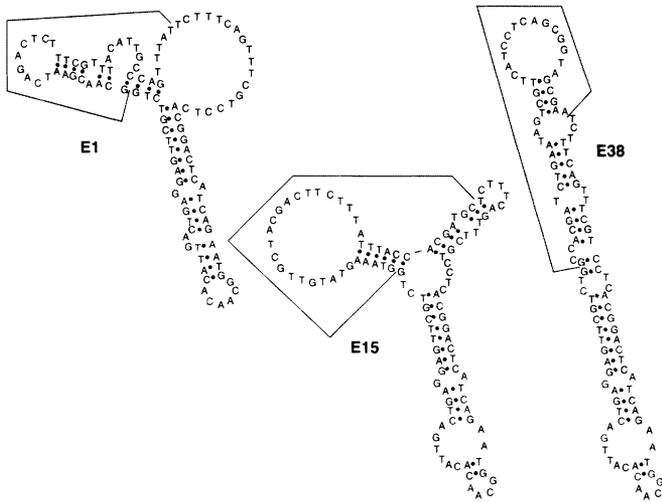


Fig. 5. Sequences and predicted secondary structures of E1, E15, and E38. Boxed part indicates selected sequence from original randomized 40-nt domain; unboxed part encodes ribozyme.

Finally, we asked whether canonical *E. coli* promoters would be active in the same context. We prepared a circle containing the 63-nt ribozyme constant domain along with a 42-nt *E. coli* *tac* promoter bottom strand (20). Transcription of this circle alone (bottom strand only) or in the presence of an equimolar amount of a 42-nt top strand complement showed that neither was transcribed well in comparison to the E15 sequence (data not shown). Based on amount of monomer ribozyme produced, the E15 circle produced 20-fold more RNA than the case with both strands of the *tac* promoter, and 40-fold more than with the bottom strand of the promoter alone. This finding is consistent with a previous observation that RCT can produce more RNA than DNAs having canonical promoters (1).

We evaluated the predicted secondary structures for the selected cloned circular DNAs (21). There was no apparent structural similarity predicted for the randomized domains, save for the fact that all formed bulged stem-loop structures without exceptionally strong secondary structure. There appeared to be no correlation between predicted free energy of the folds and transcription ability. This finding may reflect the fact that there is likely more than one way to increase the amount of monomer RNA produced. For example, increased initiation efficiency, increased processivity during transcription, increased self-processing ability, and increased ability of the DNA to be ligated to circular form may all contribute to fitness. Predicted structures for E1, E15, and E38 (Fig. 5) are all significantly different (21), which may not be surprising if different winning strategies are involved; however, we have no confirmation of what secondary structures are formed *in vivo*.

As mentioned above, one naturally occurring single-stranded promoter was previously identified that is active in *E. coli* (19). We thought it possible that some of our selected small motifs might also be active, or at least present and therefore potentially active, in *E. coli*. However, a search of the *E. coli* genome showed no significant similarities with our selected motifs.

Transplantation of Promoter Motif. As a further test of transplantability of a selected pseudopromoter motif, we constructed a new nanocircle vector encoding a different hammerhead ribozyme (the sequence is given in supporting *Materials and Methods*). This new ribozyme is targeted to *marA* RNA, which encodes multiple antibiotic resistance in certain pathogenic strains of *E. coli* (22), and is predicted to cleave it between nucleotides 40 and 41. This new ribozyme (*marA*103) is different from the previous E15 by

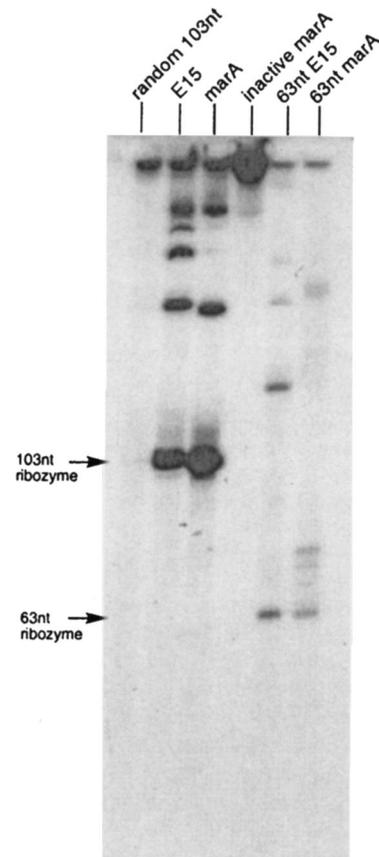


Fig. 6. Assessment of transplantability of E15 selected motif to a new nanocircle encoding *marA* ribozyme. Autoradiogram of denaturing 10% polyacrylamide gel showing *in vitro* transcription of the 103-nt initial library, nanocircle E15, the new *marA* nanocircle, *marA* nanocircle with inactivated ribozyme, and two 63-nt nanocircle controls.

13 nucleotides in sequence. We measured transcription efficiency of this new 103-nt nanocircle with *E. coli* RNAP, comparing it to the previous E15 vector as well as to a 63-mer circle lacking the selected promoter-like domain (*marA*63). Results showed (Fig. 6) that it retained full activity as compared with the E15 vector and, indeed, exceeded that activity by a significant factor. Once again, its activity was much higher than the 63-mer lacking the promoter-like sequence. Thus we conclude that at least conservative switches can be made in local structural and sequence context, with full retention of transcriptional activity. Finally, an A → C mutation in the conserved hammerhead domain of *marA*103 inactivated the cleavage activity of the ribozyme but did not diminish transcription, producing a long concatameric RNA (Fig. 6, 4th lane).

A Selected Nanocircle Vector Is Active Against *marA* RNA in *E. coli*. We then evaluated whether this *marA*103 nanocircle vector might be transcribed in bacterial cells. If it were transcribed, one expects that multimeric strings of ribozymes would be produced by RCT, and that these would subsequently self-process. The resulting monomeric ribozymes, or the concatameric precursor, could then potentially cleave *marA* RNA *in trans* (2, 23). To evaluate this possibility we prepared a reporter plasmid construct encoding a 51-nt segment of *marA* RNA in the upstream end of the CAT gene (the construct is diagrammed in Fig. 10, which is published as supporting information on the PNAS web site, www.pnas.org). Experiments showed that the extra 17 aa did not abolish CAT activity in the chimeric protein. With this construct,

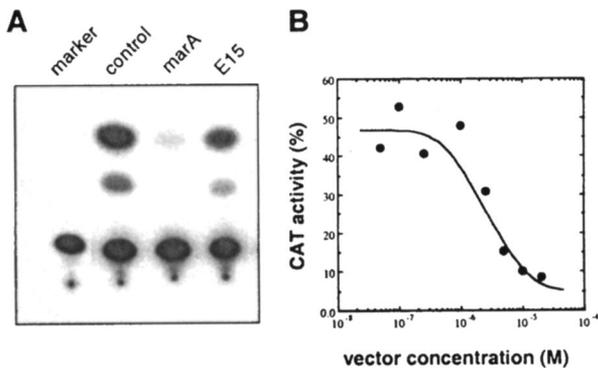


Fig. 7. Effect of nanocircle vectors on the inhibition of CAT activity. (A) Thin-layer chromatogram showing levels of CAT expressed in the presence of 10 μ M *marA* vector and E15 vector. The control lane is with no nanocircle vector. (B) Concentration dependence of down-regulation of CAT activity with *marA* vector.

cleavage of the *marA* segment of RNA would then be expected to lead to down-regulation of CAT activity by diminishing the amount of translatable CAT mRNA.

We introduced the *marA*103 nanocircle vector (NCV) and control DNAs into bacteria containing the reporter plasmid by heat shocking the cells. We measured CAT activity (by radio-labeled thin layer chromatography) as a function of nanocircle concentration in the medium during heat shock. Results show that the *marA* NCV was able to down-regulate CAT activity markedly (Fig. 7A) as compared with the control with no vector, and to the case with the E15 vector, which should be transcribed strongly but encodes a non-*marA* ribozyme (Fig. 8). Varying the amount of the *marA*103 NCV revealed a concentration-dependent down-regulation of CAT activity (Fig. 7B), reaching 50% of maximal activity at *ca.* 7 μ M nanocircle concentration and reaching *ca.* 80% inhibition at \approx 10 μ M and above. Control experiments revealed that when a single A \rightarrow C mutation, carried out to inactivate the ribozyme cleavage, was made, activity dropped significantly (Fig. 8), indicating that ribozyme cleavage is important. It also appears that some of the observed down-regulation of CAT may be due to simple antisense activity

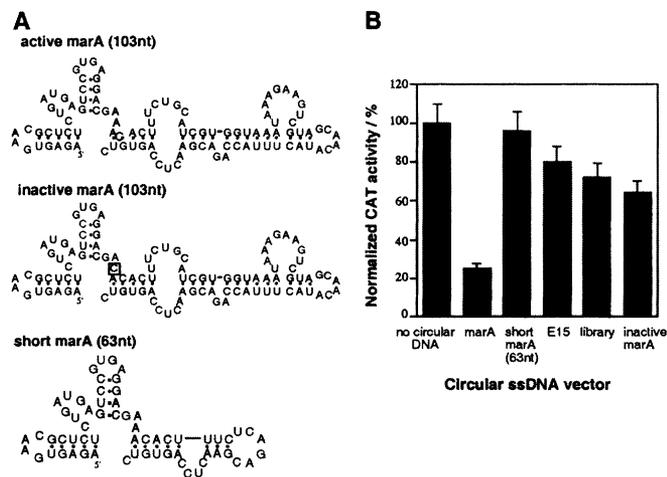


Fig. 8. (A) Sequences and predicted secondary structures of the monomer ribozymes: active and inactive *marA*, and short *marA*. The inactivating A \rightarrow C mutation is boxed in the first ribozyme. (B) Effect of 10 μ M various nanocircle vectors on the inhibition of CAT activity. The plotted data were averaged from three independent experiments.

of the ribozyme RNA, because this inactive *marA* NCV did show some down-regulation activity. Overall, the data suggest that the main activity of the *marA* monomeric (or multimeric) ribozymes arises from their ability to cleave the intended target. A separate control was carried out with the 63-mer nanocircle lacking the promoter-like domain (Fig. 8); this circle was inactive over the same concentration range. This result confirms that the greater amounts of RNA generated because of the promoter are needed to show trans-cleavage activity.

Finally, to independently check for cleavage of the intended target RNA in the bacterial cells, we carried out RT-PCR experiments with total isolated RNA, using pairs of primers on either side of the expected cleavage site, comparing the relative amounts of full-length RNA versus the expected shortened fragment [as judged by their corresponding 101- and 60-nt PCR fragments (see Fig. 11, which is published as supporting information on the PNAS web site, www.pnas.org)]. The results show a 96% decrease in full-length RNA in the presence of the *marA*103 NCV as judged by the drop in amount of 101-nt PCR fragment. Measurement of vector effects on the shorter 60-nt PCR fragment shows that the *marA* NCV causes a smaller 55% decrease, possibly due to an antisense mechanism. The results are consistent with cleavage between the two limiting primers spanning the intended site, providing further evidence in support of the expected ribozyme-mediated cleavage, decreasing CAT activity.

Discussion

The use of strongly transcribed nanocircle vectors represents a potentially useful way to deliver biologically active RNAs into bacteria. This method is quite distinct from the standard approach of delivering RNAs encoded on plasmids. Nanocircles may be easily used to engender repeating RNAs that would be difficult to encode on a plasmid stably. Although we used such a vector to encode a hammerhead ribozyme, it seems quite possible that other classes of ribozymes, as well as other short RNA motifs (such as antisense RNAs, hairpins, or dsRNAs), may be encoded (3, 4). The nanocircle vectors are not replicated and thus the encoded RNAs are transient in the cell, which may have advantages in certain applications where a drug-like approach to gene inhibition is useful, but disadvantages when permanent transformation is desired. Previous studies have shown that RCT can produce greater amounts of RNA than canonical RNA polymerase promoters, and the present comparisons also support this possibility. If that is the case, then nanocircle vectors may offer a unique way to engender more RNA in a cell than otherwise possible (A. Seyhan, A. M. Diegelman, E.T.K., and J. M. Burke, unpublished observation).

The data allow us to conclude that at least conservative switches can be made in local structural and sequence context, with full retention of transcriptional activity. It remains to be seen whether such a pseudopromoter can be active in transcription of other contexts, such as with other ribozymes or other RNA structural motifs. In cases where transplantability is limited, one could possibly carry out similar selections in those other contexts.

It is worth noting that although the transplanted E15 sequence does promote transcription, it may not necessarily do so by efficiently directing initiation at a specific site. Examination of the transcription products of E15 or the other selected circles shows no strong bands between the self-processed ribozyme bands, which would necessarily exist if transcription were initiated at one site unique from the cleavage site. Thus it is possible that initiation is not rate limiting here, and that selection has generated sequences that are especially good at enhancing processivity rather than initiation. The transcription products seem to suggest that initiation occurs at several sites in the circle. More studies will be required to determine what structural

features, beyond stem-loop and bulged duplexes, are important for strong transcription. Most if not all of the members of the randomized library also contain mismatched (looped) regions, and yet the selected winner is 80-fold more strongly transcribed than the library. It will be of significant interest in future studies to identify how small changes in sequence and structure of some of the selected motifs affect transcription initiation and elongation efficiency. Also of interest would be to examine explicitly the role of the sigma factor in initiation on these sequences; we used a commercial holoenzyme preparation in which presence of sigma subunit was confirmed, although exact stoichiometries were not apparently quantitated.

Previous reports have described optimization by *in vitro* selection of double-stranded naturally occurring promoters for phage and bacterial RNA polymerases (24, 25). The present experiments are distinct from this by involving only single-

stranded sequence and using no naturally occurring sequence as the starting point. These new promoter-like motifs are to our knowledge the first cases of new single-stranded promoters for a bacterial polymerase. The *in vitro* selection method we used has the particular advantage that winners are self-amplified by their own RCT, which can potentially lead to rapid gains in population even from rare motifs. We have recently taken advantage of this by selecting for single-stranded promoters for a phage RNA polymerase, complementing the present bacterial polymerase study (Ohmichi and Kool, unpublished work). Future studies will be directed at identifying possible single-stranded promoter motifs for other RNA polymerases.

This work was supported by a grant from the U.S. Department of Defense Breast Cancer Research Program, administered by the U.S. Army.

1. Daubendiek, S. L., Ryan, K. & Kool, E. T. (1995) *J. Am. Chem. Soc.* **117**, 7818–7819.
2. Daubendiek, S. L. & Kool, E. T. (1997) *Nat. Biotechnol.* **15**, 273–277.
3. Diegelman, A. M. & Kool, E. T. (1998) *Nucleic Acids Res.* **26**, 3235–3241.
4. Diegelman, A. M. & Kool, E. T. (1999) *Chem. Biol.* **6**, 569–576.
5. Diegelman, A. M., Daubendiek, S. L. & Kool, E. T. (1998) *BioTechniques* **25**, 754–758.
6. Frieden, M., Pedroso, E. & Kool, E. T. (1999) *Angew. Chem. Int. Ed. Engl.* **38**, 3654–3657.
7. Kool, E. T. (1998) *Acc. Chem. Res.* **31**, 502–510.
8. Kasas, S., Thomson, N. H., Smith, B., Hansma, H. G., Zhu, X., Guthold, M., Bustamente, C., Kool, E. T., Kashlev, M. & Hansma, P. K. (1997) *Biochemistry* **36**, 461–468.
9. Thomson, N. H., Smith, B. L., Almqvist, N., Schmitt, L., Kashlev, M., Kool, E. T. & Hansma, P. K. (1999) *Biophys. J.* **76**, 1024–1033.
10. Hansma, H., Golan, R., Hsieh, W., Daubendiek, S. L. & Kool, E. T. (1999) *J. Struct. Biol.* **127**, 240–247.
11. Daubendiek, S. L. (1997) Ph.D. Thesis (Univ. of Rochester, Rochester, NY).
12. Aiyar, S. E., Helmann, J. D. & deHaseth, P. L. (1994) *J. Biol. Chem.* **269**, 13179–13184.
13. Mollegaard, N. E., Buchardt, O., Egholm, M. & Nielsen, P. E. (1994) *Proc. Natl. Acad. Sci. USA* **91**, 3892–3895.
14. Daube, S. S. & von Hippel, P. H. (1992) *Science* **258**, 1320–1324.
15. Chamberlin, M., McGrath, J. & Waskell, L. (1970) *Nature (London)* **228**, 227–230.
16. Chamberlin, M. & Ring, J. (1973) *J. Biol. Chem.* **248**, 2245–2250.
17. Kadesch, T. R. & Chamberlin, M. J. (1982) *J. Biol. Chem.* **257**, 5286–5295.
18. Krupp, G. (1988) *Gene* **72**, 75–89.
19. Masai, H. & Arai, K. (1997) *Cell* **89**, 897–907.
20. DeBoer, H. A., Comstock, L. J. & Vasser, M. (1983) *Proc. Natl. Acad. Sci. USA* **80**, 21–25.
21. SantaLucia, J., Jr. (1998) *Proc. Natl. Acad. Sci. USA* **95**, 1460–1465.
22. Cohen, S. P., Hacheler, H. & Levy, S. B. (1993) *J. Bacteriol.* **175**, 1484–1492.
23. Ohmichi, T. & Kool, E. T. (2000) *Nucleic Acids Res.* **28**, 776–783.
24. Xu, J., McCabe, B. C. & Koudelka, G. B. (2001) *J. Bacteriol.* **183**, 2866–2873.
25. Breaker, R. R., Banerji, A. & Joyce, G. F. (1994) *Biochemistry* **33**, 11980–11986.

LINKED CITATIONS

- Page 1 of 2 -



You have printed the following article:

Efficient Bacterial Transcription of DNA Nanocircle Vectors with Optimized Single-Stranded Promoters

Tatsuo Ohmichi; Angèle Maki; Eric T. Kool

Proceedings of the National Academy of Sciences of the United States of America, Vol. 99, No. 1. (Jan. 8, 2002), pp. 54-59.

Stable URL:

<http://links.jstor.org/sici?sici=0027-8424%2820020108%2999%3A1%3C54%3AEBTODN%3E2.0.CO%3B2-8>

This article references the following linked citations. If you are trying to access articles from an off-campus location, you may be required to first logon via your library web site to access JSTOR. Please visit your library's website or contact a librarian to learn about options for remote access to JSTOR.

[Endnotes]

¹³ Peptide Nucleic Acid#DNA Strand Displacement Loops as Artificial Transcription Promoters

Neils Erik Mollegaard; Ole Buchardt; Michael Egholm; Peter E. Nielsen

Proceedings of the National Academy of Sciences of the United States of America, Vol. 91, No. 9. (Apr. 26, 1994), pp. 3892-3895.

Stable URL:

<http://links.jstor.org/sici?sici=0027-8424%2819940426%2991%3A9%3C3892%3APNASDL%3E2.0.CO%3B2-G>

¹⁴ Functional Transcription Elongation Complexes from Synthetic RNA-DNA Bubble Duplexes

Shirley S. Daube; Peter H. von Hippel

Science, New Series, Vol. 258, No. 5086. (Nov. 20, 1992), pp. 1320-1324.

Stable URL:

<http://links.jstor.org/sici?sici=0036-8075%2819921120%293%3A258%3A5086%3C1320%3AFTECF%3E2.0.CO%3B2-L>

²⁰ A Flow- and Time-Dependent Index of Ischemic Injury after Experimental Coronary Occlusion and Reperfusion

Laurence W. V. DeBoer; Robert E. Rude; Robert A. Kloner; Joanne S. Ingwall; Peter R. Maroko; Michael A. Davis; Eugene Braunwald

Proceedings of the National Academy of Sciences of the United States of America, Vol. 80, No. 18, [Part 1: Biological Sciences]. (Sep. 15, 1983), pp. 5784-5788.

Stable URL:

<http://links.jstor.org/sici?sici=0027-8424%2819830915%2980%3A18%3C5784%3AAFATIO%3E2.0.CO%3B2-I>

NOTE: *The reference numbering from the original has been maintained in this citation list.*

LINKED CITATIONS

- Page 2 of 2 -



²¹ **A Unified View of Polymer, Dumbbell, and Oligonucleotide DNA Nearest-Neighbor Thermodynamics**

John SantaLucia

Proceedings of the National Academy of Sciences of the United States of America, Vol. 95, No. 4. (Feb. 17, 1998), pp. 1460-1465.

Stable URL:

<http://links.jstor.org/sici?sici=0027-8424%2819980217%2995%3A4%3C1460%3AAUVOPD%3E2.0.CO%3B2-0>